

This Page Is Inserted by IFW Operations
and is not a part of the Official Record

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

As rescanning documents *will not* correct images,
please do not report the images to the
Image Problem Mailbox.

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
31 May 2001 (31.05.2001)

PCT

(10) International Publication Number
WO 01/38489 A2

(51) International Patent Classification⁷:

C12N

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.

(21) International Application Number: PCT/US00/32227

(22) International Filing Date:

22 November 2000 (22.11.2000)

(25) Filing Language:

English

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

(26) Publication Language:

English

(30) Priority Data:
09/448,215 23 November 1999 (23.11.1999) US

Published:

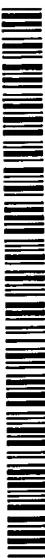
— Without international search report and to be republished upon receipt of that report.

(71) Applicant: CORNELL RESEARCH FOUNDATION,
INC. [US/US]; Suite 105, 20 Thornwood Drive, Ithaca, NY
14850 (US).

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(72) Inventors: YODER, Olen, C.; 44 Hungerford Road,
Ithaca, NY 14850 (US). TURGEON, Barbara, C.; 608
Mitchell Street, Ithaca, NY 14850 (US). LU, Shun-Wen;
604 Winston Court, Apt. #4, Ithaca, NY 14850 (US).

(74) Agents: GOLDMAN, Michael, L. et al.; Nixon Peabody
LLP, Clinton Square, P.O. Box 31051, Rochester, NY
14603 (US).



WO 01/38489 A2

(54) Title: PEPTIDE SYNTHETASE GENE CPS1

(57) Abstract: The present invention relates to genes cloned from the plant pathogens *Cochliobolus heterostrophus*, *Alternaria solani*, *Fusarium graminearum*, and *Pyrenophora teres*, that encode a CPS1 peptide synthetase required for fungal pathogenesis. The nucleic acid molecules in a vector, a host cell, or a plant is also disclosed. The invention further provides a protein or polypeptide encoded by the CPS1 genes. Other aspects of the invention relate to a method of imparting disease resistance to a plant by overexpressing a protein of the present invention in a plant and a method for identifying inhibitors of a CPS1 protein in a sample.

PEPTIDE SYNTHETASE GENE CPS1**CROSS REFERENCE TO RELATED APPLICATIONS**

This application is a continuation in part application of U.S. Application Serial No. 09/448,215, filed November 23, 1999.

5 STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH

The present invention was made with support from the United States Government under Grant No. 96-35303-3198 from the USDA/NRI. The United States Government may have certain rights in the invention.

FIELD OF THE INVENTION

10 The present invention relates to nucleic acid molecules encoding peptide synthetase homologs of *Cochliobolus heterostrophus*, *Pyrenophora teres*, *Fusarium graminearum*, and *Alternaria solani* and uses thereof.

BACKGROUND OF THE INVENTION

There are approximately 30 species included in the genus 15 *Cochliobolus*, nearly all of which are pathogens of wild grasses or cereals (Yoder et al., "Cochliobolus spp. And Their Host-Specific Toxins, in Carroll, eds., The Mycota Vol. 5: Plant Relationships, Part A, Berlin:Springer-Verlag, pp. 145-166 (1997)). *Cochliobolus heterostrophus* represents the most widely distributed species in the genus and can be found in many tropical and subtropical areas in the 20 world. As a natural pathogen of corn, *C. heterostrophus* causes a disease frequently called leaf spot of maize in the old literature (Drechsler, "Leafspot of Maize Caused by *Ophiobolus Heterostrophus* n. sp., The Ascigerous Stage of a *Helminthosporium* Exhibiting Bipolar Germination," J. Agr. Res., 31:701-726 (1925); Drechsler, "Phytopathological and Taxonomic Aspects of *Ophiobolus*, 25 *Pyrenophora*, *Helminthosporium*, and a New Genus, *Cochliobolus*," Phytopathol., 24:953-983 (1934); Yu, "Studies on *Helminthosporium* Leaf Spot of Maize," 3:273-318 (1933); Orillo, "Leafspot of Maize Caused by *Helminthosporium maydis*," 36:327-395 (1952). In the United States, *C. heterostrophus* is usually found in the warmer southern states, thus, the disease is commonly known as 30 Southern Corn Leaf Blight (Hooker, "Cytoplasmic Susceptibility in Plant

- 2 -

Disease," Ann. Rev. Phytopathol., 12:167-179 (1974)). For many years, Southern Corn Leaf Blight was only known as an endemic disease and was not considered to be major economic importance in the United States. But in 1970, it suddenly broke into a severe epidemic that destroyed 15% of the U.S. corn crop and caused 5 losses estimated at more than \$1 billion. This serious damage made Southern Corn Leaf Blight one of the most widely known crop diseases in the U.S.

Prior to the outbreak of the disease, only one race of *C. heterostrophus* (race O) was known in the field. In late 1969 when the disease became an epidemic, a new race of the fungus was identified from infected corn 10 leaves collected in severely diseased areas. It was soon designated as race T because of its high virulence on T-cytoplasm corn and the ability to produce a phytotoxin called T-toxin, which specifically affects T-corn. In contrast, race O does not produce T-toxin and is mildly virulent on both T-cytoplasm and N-cytoplasm (normal cytoplasm) corn (Hooker et al., "Physiological Races of 15 *Helminthosporium maydis* and Disease Resistance," Plant Dis. Repr., 54:1109-1110 (1970); Scheifele, "Cytoplasmically Inherited Susceptibility to Diseases as Related to Cytoplasmically Controlled Pollen Sterility in Maize," 25:110-138 (1970); Smith et al., "Physiologic Races of *Helminthosporium maydis*," 54:819-822 (1970); Yoder et al., "Segregation of Pathogenicity Types and Host-Specific 20 Toxin Production in Progenies of Crosses Between Races T and O of *Helminthosporium maydis* (*Cochliobolus heterostrophus*)," Phytopathology, 65:273-276 (1975); Yoder, "Evaluation of the Role of *Helminthosporium maydis*, Race T Toxin in Southern Corn Leaf Blight, in Tomiyama, eds., Biochemistry and Cytology of Plant Parasite Interaction, New York, New York:Elsevier, pp. 16-24 25 (1976); Yoder, "Toxins in Pathogenesis," Ann. Rev. Phytopathol., 18:103-129 (1980)). T-cytoplasm stands for Texas male sterile cytoplasm, a unique cytoplasm with a trait for maternally inherited male sterility, characterized by the failure to produce pollen (Levings, "The Texas Cytoplasm of Maize: Cytoplasmic Male Sterility and Disease Susceptibility," Science, 250:942-947 (1990)). T- 30 cytoplasm corn was widely used for hybrid seed production and breeding to avoid hand or mechanical emasculation in the 1950s and the 1960s. It was the coexistence of large acreages of intensively planted T-cytoplasm corn and the

sudden appearance of race T of *C. heterostrophus* that resulted in the epidemic of the disease in 1970. This discovery first opened the door to understanding pathogenesis by *C. heterostrophus*.

Early genetic analysis suggested that both T-toxin production and high virulence on T-cytoplasm corn are controlled by a single genetic locus defined as *Tox1* (Leach et al., "Dominance at the *Tox1* Locus Controlling T-Toxin Production by *Cochliobolus heterostrophus*," *Physiol. Plant Pathol.*, 21:327-333 (1982)). This was demonstrated by crosses between race T and race O in which only parental phenotypes segregated in a 1:1 ratio (*Tox⁺*:*Tox⁻*); all T-toxin producing progeny are highly virulent on T-cytoplasm corn while all T-toxin nonproducing progeny are weakly virulent (Yoder et al., "Segregation of Pathogenicity Types and Host-Specific Toxin Production in Progenies of Crosses Between Races T and O of *Helminthosporium maydis* (*Cochliobolus heterostrophus*)," *Phytopathology*, 65:273-275 (1975); Leach et al., "Dominance at the *Tox1* Locus Controlling T-Toxin Production by *Cochliobolus heterostrophus*," *Physiol. Plant Pathol.*, 21:327-333 (1982)). Further investigation by comparison of electrophoretic karyotypes and chromosome-specific DNA hybridizations indicated that *Tox1* is tightly linked to a reciprocal translocation breakpoint and is associated with as much as a megabase of DNA (mostly highly repeated and A+T-rich) that is missing in race O (Bronson, "Ascospore Abortion in Crosses of *Cochliobolus heterostrophus* Heterozygous for the Virulence Locus *Tox1*," *Genome*, 30:12-18 (1988); Tzeng et al., "A Restriction Fragment Length Polymorphism Map and Electrophoretic Karyotype of the Fungal Maize Pathogen *Cochliobolus heterostrophus*," *Genetics*, 130(1):81-96 (1992); Chang et al., "A Reciprocal Translocation and Possible Insertion(s) Tightly Associated with Host-Specific Virulence in *Cochliobolus heterostrophus*," *Genome*, 39(3):549-557 (1996)). Surprisingly, recent analyses of several *Tox⁻* mutants revealed that *Tox1* is not a single locus but rather two loci, each on a different translocated chromosome (Yoder et al., "Molecular Determinants of the Plant/Fungus Interaction," in Kohmoto, eds., *Host-Specific Toxin: Biosynthesis, Receptor and Molecular Biology*, Tottori, Japan:Faculty of Agriculture, Tottori Univ., pp. 23-32 (1994); Turgeon et al., "Function and Chromosomal Location of the *Cochliobolus*

- 4 -

heterostrophus Tox1 Locus," Can. J. Bot., 73 (suppl. 1 Sect. E-H):S1071-S1076 (1995)). These two *Tox1* loci have been designated *Tox1A* and *Tox1B* (Yoder et al., "Cochliobolus spp. and Their Host-Specific Toxins," in Carroll, eds., The Mycota Vol. 5: Plant Relationships, Part A, Berlin:Springer-Verlag, pp. 145-166 (1997)). Two genes *PKS1* and *DEC1* have been cloned from the two loci respectively; both are required for biosynthesis of T-toxin and are found only in race T isolates of *C. heterostrophus* (Yang, "The Molecular Genetics of T-Toxin Biosynthesis by *Cochliobolus heterostrophus*," Ph.D. Thesis, Cornell University (1995); Yang et al., "A Polyketide Synthase is Required for Fungal Virulence and Production of the Polyketide T-Toxin," Plant Cell, 8(11):2139-2150 (1996); Rose et al., "A Decarboxylase Required for Polyketide Toxin Production and High Virulence by *Cochliobolus heterostrophus*," 8th Int. Symp. Mol. Plant-Microbe Int., Knoxville, p. J-49 (1996)).

Genetic analysis also suggested that T-toxin is required by *C. heterostrophus* for its high virulence on T-cytoplasm corn. This hypothesis was first tested by the generation of induced T-toxin deficient mutants using different mutagenesis procedures. All mutants with a tight *Tox⁻* phenotype cause disease symptoms that are indistinguishable from those caused by race O when tested on both T and N-cytoplasm corn, suggesting that T-toxin is indeed a virulence factor (Yang et al., 1992; Lu et al., "Tagged Mutations at the *Tox1* Locus of *Cochliobolus heterostrophus* Using Restriction Enzyme-Mediated Integration," Proc. Natl. Acad. Sci. USA, 91:12649-12653 (1994); Rose et al., "A Decarboxylase Required for Polyketide Toxin Production and High Virulence by *Cochliobolus heterostrophus*," 8th Int. Symp. Mol. Plant-Microbe Int., Knoxville, p. J-49 (1996)). This conclusion was firmly supported by the site-specific disruption of the *PKS1* or *DEC1* in the wild type race T genome; disruptants lost the ability to produce T-toxin and caused race O type symptoms on both T-corn and N-corn (Yang et al., "A Polyketide Synthase is Required for Fungal Virulence and Production of the Polyketide T-Toxin," Plant Cell, 8(11):2139-2150 (1996); Rose et al., "A Decarboxylase Required for Polyketide Toxin Production and High Virulence by *Cochliobolus heterostrophus*," 8th Int. Symp. Mol. Plant-Microbe Int., Knoxville, p. J-49 (1996)). These experiments have given a very

- 5 -

clear resolution for the role of T-toxin in pathogenesis. They also implied that pathogenesis by *C. heterostrophus* must involve additional pathogenicity factors because race O which does not produce T-toxin and race T-derived *Tox-* mutants are effective pathogens on corn.

- 5 A number of fungal molecules have been identified as general pathogenicity or virulence factors in several plant pathogenic fungi (Yoder et al., "Molecular-Genetic Evaluation of Fungal Molecules for Roles in Pathogenesis in Plants," *J. Genet.*, 75(3):425-440 (1996)). These include potential penetration factors such as melanin (Guillen et al., "Linkage Among Melanin Biosynthetic
- 10 Mutations in *Cochliobolus heterostrophus*," *Fungal Genet. NewsL*, 41:41-42 (1994)), cutinase (Oeser et al., "Pathogenesis by *Cochliobolus heterostrophus* Transformants Expressing a Gene Encoding Cutinase from *Nectria haematocephala*," *Mol. Plant-Microbe Int.*, 7:282-288 (1994)) and polygalacturonase and xylanase (Lyngholm et al., "Mutants of *Cochliobolus heterostrophus* Deficient in Extracellular Enzymes," *Fungal Genet. NewsL*, 42:46-47 (1995)) or possible mechanisms involved in colonization such as phytotoxin detoxification (Schäfer et al., "One Enzyme Makes a Fungal Pathogen, But Not a Saprophyte, Virulent on a New Host Plant," *Science*, 246:247-249 (1989)) or components of signal transduction pathways (Horwitz et al., "A G Protein Alpha
- 15 Subunit Gene From the Corn Pathogen *Cochliobolus heterostrophus* is Involved in Two Complex Developmental Pathways: Mating and Appressorium Formation (unpublished) (1997)). Although *C. heterostrophus* is known to produce a nonhost specific toxin called ophiobolin (or cochliobolin), a C₂₅ sesterterpenoid compound, which is toxic to many organisms, including plants, bacteria, fungi and
- 20 nematodes, there is no evidence that ophiobolins are involved in pathogenesis by *C. heterostrophus* or other phytopathogenic fungi. No other pathogenesis-related toxins have been isolated from *C. heterostrophus* so far, but studies on closely related *Cochliobolus* species and other phytopathogenic fungi suggest that pathogenesis by this group of fungi also involves peptide toxins.
- 25
- 30 Four peptide phytotoxins (victorin, HC-toxin, AM-toxin, and enniatins) have been characterized as pathogenicity or virulence factors. They are

- 6 -

all small cyclic peptides (4-6 residues), containing unusual amino acids or hydroxy acids, and they can be either host specific or non-host specific in terms of plant toxicity. A number of peptide phytotoxins are believed to be synthesized nonribosomally. Early in the 1960s, several biochemists working on the bacterial peptide antibiotics gramicidin and tyrocidine found that these polypeptides can be synthesized in RNAase-treated particle-free extracts of *Bacillus brevis* that are known to produce the same antibiotics; adding protein-synthesis inhibitors to the extracts does not affect this process. This indicated the existence of a peptide biosynthetic system in which ribosomes and mRNAs are not needed. Further studies revealed that in this system, peptides are synthesized on a protein-template and this template itself is a multifunctional enzyme or a complex of several such enzymes, collectively called peptide synthetases, catalyzing the biosynthetic process (Laland et al., "The Protein Thiotemplate Mechanism of Synthesis for the Peptide Antibiotics Produced by *Bacillus Brevis*," Essays in Biochemistry, 7:31-57 (1973); Lipmann, "Bacterial Production of Antibiotic Polypeptides by Thiol-Linked Synthesis on Protein Templates," Adv. Microbiol. Physiol., 21:277-266 (1980)).

Peptide synthetases can catalyze biosynthesis of a variety of peptides. In terms of bioactivity, they can be antibiotics, enzyme inhibitors, plant or animal toxins and immunosuppressants (Stachelhaus et al., "Modular Structure of Peptide Synthetases Revealed by Dissection of the Multifunctional Enzyme GrsA.," Journal of Biological Chemistry, 270(11):6163-6169 (1995)). In terms of chemical structure, they can be either linear (i.e. ACV, the penicillin precursor and gramicidin) or cyclic (most are). The latter can be further classified into three subgroups: 1) The "standard" cyclic peptides (i.e. gramicidin S, tyrocidine, HC-toxin and cyclosporin); 2) cyclic lactones (i.e. destruxin); 3) cyclic depsipeptides (i.e. beauvericin and enniatin). There have been over 300 different carboxy compounds that can be activated by peptide synthetases.

Although the first peptide synthetase, Gramicidin S synthetase, was purified and used for the cell-free synthesis of the peptide early in the 1960s (Tomino et al., "Cell-Free Synthesis of Gramicidin S," Biochem., 6:2552-2560 (1967)), the first bacterial peptide synthetase gene, *tycA*, which encodes the

- 7 -

tyrocidine synthetase 1 in *B. brevis* was not cloned until almost twenty years later (Marahiel et al., "Cloning of the Tyrocidine Synthetase 1 Gene from *Bacillus-brevis* and Its Expression in *Escherichia-coli*, Mol. Gen. Genet., 201(2):1986 (1985)). Since then, more than twenty peptide synthetase genes have been

5 reported for both bacteria and filamentous fungi, but only fourteen have complete nucleotide sequences published. All are larger than 3.3 kb and range between 3.3-19.5 kb for bacterial genes and 9.4-45.8 kb for fungal ones. Interestingly, all fungal peptide synthetase genes reported lack introns, even the cyclosporin A synthetase gene *simA*, which has a 45.8 kb of open reading frame (the largest

10 genomic ORF so far recorded). Although biosynthesis of bacterial peptides differs from that of fungal ones in terms of the number of multifunctional enzymes involved, the genes encoding these enzymes are similar to each other in both function and structure. Comparison of nucleotide sequences reveals one or more highly conserved regions at certain positions in each peptide synthetase

15 gene. These regions formerly called "amino acid activating domains" (Stachelhaus et al., "Modular Structure of Peptide Synthetases Revealed by Dissection of the Multifunctional Enzyme GrsA," Journal of Biological Chemistry, 270(11):6163-6169 (1995)), now called "amino acid activating modules" (Marahiel, "Protein Templates for the Biosynthesis of Peptide

20 Antibiotics," Chem. Biol., 4(8):561-567 (1997)) consist of a set of domains (formerly called "modules") believed to have specific functions such as recognition, activation and thioesterification of individual constituent amino or hydroxy acids, and in some cases methylation and racemation for modification of certain residues before incorporation into the peptide chain (Stachelhaus et al.,

25 "Modular Structure of Peptide Synthetases Revealed by Dissection of the Multifunctional Enzyme GrsA," Journal of Biological Chemistry, 270(11):6163-6169 (1995)). The most convincing evidence supporting this assignment is that in most cases, the number of conserved functional units in each gene or gene cluster is equal to the number of amino acids in the respective peptide. This one-for-one

30 match is very clear between three of four fungal peptides and their biosynthetic genes. The total number of modules in three of four bacterial gene clusters also matches the number of amino acids in the respective peptides.

- 8 -

Sequence alignment of amino acid-activating modules reveals strictly conserved sequence motifs that contain active residues for module functions. These motifs are called "core sequences" (Marahiel, "Multidomain Enzymes Involved in Peptide Synthesis," FEBS Lett., 307(1):40-43 (1992)). A 5 minimal amino acid-activating module must contain six core sequences, whose functions (except for core 1) have been proposed based on mutational analysis of several peptide synthetases. Core sequences 1-5 are grouped into an amino acid adenylation domain and core 6 is a thioester formation domain (Figure 1A). All bacterial peptide synthetase genes contain "type I modules" – the minimal amino 10 acid activating modules which were previously called "type I domains" (Stachelhaus et al., "Modular Structure of Peptide Synthetases Revealed by Dissection of the Multifunctional Enzyme GrsA," Journal of Biological Chemistry, 270(11):6163-6169 (1995)). Two fungal genes, *acvA* and *HTS1* also have this modular structure. In addition to the type I module, two fungal genes, 15 *esyn1* and *simA*, contain type II modules, in which an insertion (about 400 amino acids) is found between cores 5 and 6 of a normal type I module. This region contains a motif (VLE/DXGXGXG (SEQ. ID. No. 1)), highly conserved in S-adenosyl-methionine (SAM)-dependent methyltransferases, hence, it is referred to as a *N*-methylation domain (Figure 1A). Additional evidence for 20 methyltransferase activity of this module is that the number and position of type II modules in *esyn1* and *simA* exactly match that of *N*-methylated amino acids in ennatin and cyclosporin sequences (Figure 1B).

Although the modular structure described above is highly conserved among most peptide synthetase genes, some variations have been found 25 in the latest cloned peptide synthetase gene *safB*, which is the first gene in the saframycin Mx1 synthetase gene cluster (Pospiech et al., "A New *Myxococcus xanthus* Gene Cluster for the Biosynthesis of the Antibiotic Saframycin Mx1 Encoding a Peptide Synthetase," Microbiology, 141(8):1793-1803 (1995)). *safB* contains two type I amino acid activating modules. One module has all six highly 30 conserved core sequences, but another, believed to activate alanine (the first amino acid in the linear tetrapeptide precursor of saframycin Mx1), lacks core 5 and has a weakly conserved core 1 (Pospiech et al., "Two Multifunctional Peptide

- 9 -

Synthetases and an O-methyltransferase Are Involved in the Biosynthesis of the DNA-Binding Antibiotic and Antitumour Agent Saframycin Mx1 from *Myxococcus xanthus*," *Microbiology*, 142(4):741-746 (1996)) (Figure 1A). This suggests that some of the motifs in the amino acid adenylation domain are
5 dispensable or not critical for domain function. It also raises the possibility that other variations might be found in yet unknown peptide synthetase genes.

Although *C. heterostrophus* has been a model eukaryotic plant pathogen since the 1970s, most molecular genetic analyses conducted in this system have focused on production of the polyketide T-toxin by race T isolates of
10 the fungus. Solid evidence now indicates that T-toxin is a host-specific virulence factor in Southern Corn Leaf Blight (Yoder et al., "Molecular-Genetic Evaluation of Fungal Molecules for Roles in Pathogenesis in Plants," *J. Genet.*, 75(3):425-440 (1996); Yoder et al., *Cochliobolus spp.* and Their Host-Specific Toxins, Carroll eds., *The Mycota Vol. 5: Plant Relationships, Part A*, Berlin:Springer-Verlag, pp. 145-166 (1997)). It is clear, however, that *C. heterostrophus* needs additional factors, presumably general factors for pathogenesis to corn plants, since race O, which does not produce T-toxin, can be an effective corn pathogen. Attempts to identify additional general factors required by *C. heterostrophus* for pathogenesis have been unsuccessful. Cloning and characterizing additional *C.*
15 *heterostrophus* genes that control biosynthesis of novel fungal molecules involved in critical pathogenic processes may be important because it offers potential targets for the design of products that might interfere with the corn plant infection process. The present invention is directed to achieving these objectives.

SUMMARY OF THE INVENTION

25 The present invention relates to an isolated nucleic acid molecule encoding CPS1 peptide synthetase homologs. The DNA molecule comprises a nucleotide sequence which hybridizes to a DNA molecule having a sequence as set forth in at least one of SEQ. ID. No. 2, SEQ ID No. 41, SEQ ID No. 43, or SEQ ID No. 45.

30 Another aspect of the present invention relates to a method for identifying inhibitors of a CPS1 protein or polypeptide which involves providing

- 10 -

the CPS1 protein or polypeptide, contacting the protein with potential inhibitor compounds, determining peptide synthetase activity, and selecting compounds which decrease the peptide synthetase activity.

Still another aspect of the present invention relates to a method of
5 imparting disease resistance to a plant by overexpressing a CPS1 protein or
polypeptide in the plant cell.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 provides the structure of amino-acid activating modules identified in peptide synthetase genes (adapted from Stachelhaus and Marahiel, 10 1995; Pospiech 1995; Marahiel, 1997). Figure 1A shows the domain arrangements in two types of modules. Structural variations in the first module (safB1) of the gene *safB* are also indicated below type I. Figure 1B shows the correlation between module types and the nature of residues in two fungal peptides. Open box: type I module; filled box: type II module. Each peptide 15 sequence is given below.

Figure 2 depicts the recovery of DNA flanking the REMI vector insertion site (arrows) in mutant R.C4.2696. Circled numbers indicate restriction enzyme sites used for recovery of each plasmid. p214B7 contains 4.2 kb flanking DNA (3.4 left; 0.7 right); p214M1 contains 0.1 kb left flank that overlaps with 20 p214B7 and 1.1 kb right flank that overlaps with p214S1, which contains 3.2 kb flanking DNA on the left only.

Figures 3A-B illustrate the extension of the tagged sequence by targeted integration and plasmid rescue. Figure 3A shows a general scheme illustrating chromosome walking strategy (only one direction is indicated). "X" 25 indicates a restriction enzyme site used for recovery of vector (indicated by "T") with flanking genomic DNA ("L" for left flank; "R" for right flank; R' for a fragment from "R" that is subcloned into a subsequent transformation vector). Each integration site is indicated by a vertical arrowhead pointed at the selectable marker on the vector. Overlapped flanking DNA is represented by bars drawn in 30 the same pattern. Figure 3B shows two strategies used for construction of a

- 11 -

targeted integration vector. Left: A genomic DNA fragment (indicated by the letter "a") is subcloned into a vector directly and the subsequent integration occurs by a single crossover ("a" is duplicated). Right: "a" is subcloned into a vector via multiple steps (see Figure 6 for an example) that allows linearization with restriction enzyme "Y" and the subsequent integration occurs by a double crossover resulting in a simple insertion.

Figure 4 shows how the targeted integration vector p214SNP was constructed. The sites for restriction enzyme digestion are indicated by arrows. p214SNP was originally designed for a double crossover integration after digested with *Bst*XI and *Bcl*I, but it was found later that the *E. coli* strain (DH5 α) used for amplification of the plasmid is Dam $^+$ which blocks the *Bcl*I site (TGA TCA to TG m TCA). As an alternative, the plasmid was linearized with *Bst*XI only and used for transformation.

Figure 5 depicts the recovery of genomic DNA flanking the targeted integration vector p214SNP in transformant #118. The vector integrated into the target site by a single crossover that resulted in duplicated *Hind*III-*Sac*I fragments (indicated by letter b and b'). The second genomic DNA fragment carried by the vector is also duplicated in the genome (*Nar*I-*Hind*III, indicated by letter a and a'). Genomic DNA was digested with *Bgl*II (which does not cut the vector) or *Bcl*I (which cuts once in fragment a' on the vector) as indicated by numbers in a circle. The two recovered plasmids carry sequences that extend the right flank only by 170 and 800 bp respectively beyond the genomic DNA on p214S1.

Figure 6 shows how the targeted integration vector p118BSP was constructed. The sites for restriction enzyme digestion on the vectors are indicated by arrows. p118B14 contains two duplicated *Nar*I-*Hind*III-*Sac*I fragments (numbered region, a + b = a' + b') but only one is present in p118BSP. p118BSP was linearized with *Bgl*II and transformed into a wild type C. *heterostrophus* strain.

- 12 -

Figure 7 depicts the recovery of genomic DNA flanking the targeted integration vector p118BSP in transformant #9. The vector integrated into the target site by a double crossover. Genomic DNA was digested with *Pst*I and religated. The recovered plasmid p9P2 includes the entire pUC18 sequence on p118BSP and 4.6 kb of genomic DNA that contains all of ORF1 (*CPS*1), including the stop codon (TAG) and 3.0 kb of genomic region 3' of the stop codon.

Figure 8 shows how the targeted integration vector p118BCS was constructed. The sites for restriction enzyme digestion are indicated by arrows.

Figure 9 depicts the recovery of genomic DNA flanking the targeted integration vector p118BSP in transformant #12. The vector integrated into the target site by a single crossover that resulted in a duplicated *Ssp*I-*Ssp*I fragment in the transformant genome (indicated by the letter "a"). Genomic DNA was digested with *Hind*III (arrows) and ligated. The recovered plasmid p12H6 contains the entire p118BSP sequence and a 2.1 kb genomic DNA (*Sac*I-*Hind*III) on the left region that overlaps with the sequence carried on p9P2 (see Figure 7).

Figures 10A-B are photographs which show that the REMI mutant R.C4.2696 grows like wild type in culture (Figure 10A) and produces normal appressoria (Figure 10B). In Figure 10A, plates containing complete medium (CM) were inoculated with a conidia-bearing mycelium plug and incubated at 22°C under warm white light (F40/350BL) (Sylvania Inc., Danvers, MA). The photograph in Figure 10A was taken 6 days after inoculation. Left to right: Mutant R.C4.2696; wild type. In Figure 10B, conidia of mutant (left) or wild type (right) were placed in a drop of water and incubated at 32°C for 6 hrs. No significant difference in percentage of appressorium-forming conidia (arrows) was detected.

Figures 11A-B are photographs which show that the REMI mutant R.C4.2696 produces wild type levels of T-toxin (Figure 11B) but has reduced virulence on T-cytoplasm corn (Figure 11A). In Figure 11A, two week old T-cytoplasm corn plants were inoculated with conidial suspensions of (left to right)

- 13 -

mutant R.C42696, wild type race T, and a Tox⁺ mutant progeny from a cross between R.C4.2696 and a race O tester, and symptoms (represented by three diseased leaves for each strain) were recorded 5 days after inoculation. Leaves infected by wild type race T collapsed (arrow) but those infected by mutants 5 remained alive. In Figure 11B, a plate containing T-toxin-sensitive *E. coli* cells was inoculated with agar blocks bearing mycelia of three strains (in the same order as in Figure 11A) and incubated at 32°C overnight (the inoculum at bottom is race O control). No significant differences in T-toxin production (indicated by halos) among the three strains were detected.

10 Figure 12 is a photograph which displays that the REMI mutant R.C4.2696 produces lesions much smaller than wild type on N-cytoplasm corn. Two week old N-cytoplasm corn plants were inoculated with conidial suspensions at the same concentration and symptoms were recorded 7 days after inoculation. The mutant (right) produces the same number of lesions as wild type (left) but the 15 size of lesions was dramatically reduced compared to wild type (arrows).

Figures 13A-B show that the REMI mutant R.C4.2696 has a 60% reduction of virulence compared to wild type. In Figure 13A, lengths of 100 typical lesions from corn leaves inoculated with wild type race O and a mutant progeny R45 (*Tox*⁻, *hygB*^R) carrying the R.C4.2696 mutation were measured 7 20 days after inoculation and values plotted. Figure 13B shows the statistical analysis that 86% of the mutant lesions are less than 4 mm in length (average size 3.5 mm), 60% reduced compared to that of wild type (8.5 mm).

Figures 14A-B illustrate the genetic analysis showing that a tagged, 25 single site mutation is responsible for the mutant phenotype. Figure 14A is a photograph of a plant assay where N-cytoplasm corn was inoculated with parents and progeny indicated in Figure 14B (a complete tetrad from the second cross is shown here; random spore assay for the two crosses gave the same results). Figure 14B is a table which shows progeny segregation data. In both crosses, progeny segregated 1 : 1 for parental type only and all hygromycin B resistant 30 progeny showed the same small lesion phenotype (represented by progeny 1-4 in Figure 14A) as the mutant parent (parent 1 in Figure 14A); all hygromycin B

- 14 -

sensitive progeny show the large lesion phenotype (represented by progeny 5-6 in Figure 14A) of the wild type parent (parent 2 in Figure 14A).

Figure 15 is a restriction map of the cloned sequences surrounding the tagged site. A 11.3 kb genomic region (thick line) was cloned and completely sequenced. The original REMI insertion point in the mutant R.C4.2696 is indicated by a vertical arrow. The asterisks indicate two targeted integration sites in the wild type genome. Two open reading frames (in opposite directions), ORF1 (*CPS1*, 5.4 kb) and ORF2 (*TES1*, 1.1 kb) are indicated by open boxes below the map (the positions of putative introns are indicated by vertical bars).
5 Locations of seven overlapping plasmid clones used for sequencing are indicated by thin lines on the top of the map (filled triangles represent the vector sequence in each clone). Sequencing strategy is indicated by arrows above each clone line.
10

Figure 16 graphically depicts the G+C content of the cloned sequences surrounding the tagged site. The 11.3 kb of genomic DNA sequence (Figure 15) was examined as 200 bp fragments and G+C content of each fragment was calculated and plotted. Regions A and C have normal G+C content; regions B and D show high G+C content. The overall percentage of G+C in each region is indicated by underlined numbers. The positions of ORF1 (*CPS1*) and ORF2 (*TES1*) are indicated by arrows and the percentage of G+C of each ORF is given
15
20 below (in parentheses).

Figure 17 displays the nucleotide sequence of *CPS1*. 5,725 base pairs from the 11.3 kb sequenced region (Figure 15) are shown. The deduced amino acid sequence of *CPS1* protein is given below the DNA sequence. The position of start codon ATG (bold and underlined) is designated +1 and the open reading frame stops at position 5381 (TAG, in bold and underlined). Five putative "CAAT" boxes (bold and indicated by asterisks) are found at the positions -36, -58, -67, -172 and -309. Three putative introns (in lowercase with 5' and 3' splice sequences in bold; branch sites are underlined) are located at positions 2070-2114, 3542-3592 and 4197-4249. Conserved core sequences are shaded. The GXSXG motif is boxed. A putative polyadenylation signal is located at position 5604 (bold and overlined).
25
30

- 15 -

Figure 18 displays the nucleotide sequence of *TES1*. 1,901 base pairs 5' of *CPS1* are shown. The deduced amino acid sequence of the *TES1* protein is given below the DNA sequence. The CT motif (bold and indicated by asterisks) is found at position -46. A "AT"-rich region (bold and italicized) is 5 found at position -12. The position of ATG start codon (bold and underlined) is designated +1 and the open reading frame (transcribed in the opposite direction from *CPS1*) stops at position 1153 (TGA, in bold and underlined). One putative intron (in lowercase, border sequences are bold; branch sites are underlined) is located at position 518-566. A putative polyadenylation signal is located at 10 position 1345 (bold and overlined). The putative active site (VHS) is shaded.

Figures 19A-C are schematic representations which show the characterization of modular structure of *CPS1*. Peptide synthetase and thioesterase are indicated by open boxes; shaded boxes inside indicate functional domains and modules; vertical bars in the shaded boxes indicate highly conserved 15 core sequences. Figure 19A illustrates the general structure of bacterial and fungal peptide synthetases (adapted from Marahiel 1997, which is herein incorporated by reference). A peptide synthetase gene cluster is shown on the top. There can be one or more amino acid activating module (cyclosporine synthetase has 11) in each protein; some peptide synthetases have thioesterase domains (TE), 20 which can be either integrated into modules or encoded by a separate gene. Each synthetase can have type I, type II or both modules. A type I (minimal) module is enlarged to show organization of core sequences and domains. Some peptide synthetases also have condensation or epimerization domains. Figure 19B illustrates the organization of saframycin Mx1 synthetase containing 4 amino acid 25 activating modules (Pospiech et. al., "Two Multifunctional Peptide Synthetases and an O-methyltransferase are Involved in the Biosynthesis of the DNA-Binding Antibiotic and Antitumour Agent Saframycin Mx1 from *Myxococcus xanthus*," *Microbiology*, 142(4):741-746 (1996)). SafB1 from the first module is enlarged. Core sequences 1 and 5 in safB1 are weakly conserved (indicated by dashed 30 vertical bars). The remaining domains are typical of type I as shown in Figure 19A. SafC is a putative O-methyltransferase. Figure 19C illustrates the organization of *CPS1*. Sequence analysis revealed two amino acid activating

- 16 -

modules (CPS1A and CPS1B), both of which have high similarity to safB1 except that core 2 is weakly conserved. A thioesterase domain is found at the C-terminal region of CPS1B. Three vertical arrows indicate the positions of targeted gene disruptions in the wild type genome that yielded the mutant phenotype. TES1 is a 5 thioesterase encoded by a separate gene (*TES1*).

Figures 20A-F are comparative alignments of core amino acid sequences in CPS1A and CPS1B with those of other peptide synthetases. Figures 20A-E are comparative amino acid sequence alignments of amino-acid-activating domains (cores 1-5); Figure 20F is a comparative amino acid sequence alignment 10 of the thioester formation domain (core 6). In each subfigure, the first column shows the names of peptide synthetases; the second indicates the position of the first residue aligned in the original amino acid sequence of each protein; the last column on the right indicates the number of amino acids between two cores (Figures 20A-E, in parentheses) or the distance between two adjacent amino-acid-activating modules (Figure 20F, in parentheses). There is an extra column on the 15 right in Figure 20F, showing the total number (underlined) of residues in each amino-acid-activating module in which the aligned core sequence is located. The consensus of each core sequence is on the top, which includes identical or similar residues found in all peptide synthetases or with only a few exceptions (active site 20 is also indicated by asterisks). SafB1: the first module in saframycin Mx1 synthetase B of *Myxococcus xanthus* (Genbank accession U24657); GrsA: gramicidin S synthetase A of *Bacillus brevis* (SWISS PROT accession P14687); HTS1A and HTS1B: the first two modules in HC-toxin synthetase of 25 *Cochliobolus carbonum* (Q01886); EsynA and EsynB: two modules in enniatin synthetase of *Fusarium scirpi* (EMBL accession Z18755); ACVA and ACVB: the first two modules in ACV synthetase of *Aspergillus nidulans* (SWISS PROT P19787); CsypA and CsypB: the first two modules in cyclosporine synthetase of *Tolyphocladium nivale* (EMBL Z28383).

Figure 21 is a comparative alignment of amino acid sequences of 30 active sites of thioesterase domains (TE) in CPS1 with those of other peptide synthetases. ACV: ACV synthetase (Swiss-PROT accession P19787); GrsB: gramicidin S synthetase B (P14688); GrsT: the thioesterase encoded by *grsT*

- 17 -

(P14686) in gramicidin S synthetase gene cluster; SrfA: surfactin synthetase A-3 (Q08787); TycC: tyrocidine synthetase C (Genbank accession AF004853); TycF: the thioesterase encoded by *tycF* (AF004853) in the tyrocidine synthetase gene cluster. The highly conserved residues (GXSXG) are indicated by asterisks. The 5 number on the left of each amino acid sequence indicates the original position of the first residue; the number on the right (in parentheses) indicates the distance between the last residue shown to the end of each protein.

Figure 22 is a comparative alignment of the amino acid sequence of the *TES1* protein (CH-TES1) with that of other type II thioesterases. HS-TEII: 10 *Homo sapiens* thioesterase II (EMBL accession X86032); EC-TESB: *E. coli* acyl-coA thioesterase II (Genbank accession M63308); MT-TESB: *Mycobacterium tuberculosis* homolog to *E. coli* acyl-coA thioesterase II (EMBL Z95387). The identical residues in all four proteins are in bold. The putative active site VHS motif is indicated by asterisks. The numbers on the right column indicate the 15 original position of the last residue of the line in each protein sequence. The entire protein sequence of each TES was aligned using the Jotun Hein Method. Amino acids corresponding to the positions 142-171, 236-265 and 356-367 in CH-TES1 have no significant similarity among the four proteins.

Figures 23A-B are photographs displaying a plant assay (Figure 20 23A) and a gel blot (Figure 23B). Targeted gene disruption suggests that *CPS1* is involved in fungal pathogenesis. In Figure 23A, N-cytoplasm corn was inoculated with (left to right): The REMI mutant R.C4.2696; wild type race T; wild type race O and five disruptants obtained using the linearized p214B7 as vector. All 25 disruptants give the small lesions similar to the original mutant. In Figure 23B, total genomic DNA was digested with *Bg*II and probed with both 5' and 3' end flanking DNA fragments carried on p214B7 separately (strain order is the same as above). A single band (4.2 kb) is present in both wild type race T and race O but replaced by a 9.3 kb band (increased by the size of the vector, 5.1 kb) in all strains that showed the mutant phenotype in Figure 23A.

30 Figure 24 displays a gel blot analysis showing targeted integration of the chromosome walking vector p214SNP into the wild type genome. Lane 1:

- 18 -

wild type race O. Lanes 2-8: seven transformants obtained by transforming wild type race O (strain C5) with p214SNP. Genomic DNAs were digested with *Bgl*II and probed with 3.2 kb flanking genomic DNA fragment cloned on p214S1 which gives three bands in wild type (4.2, 2.1 and 0.5 kb, indicated by arrows on the left). The targeted site is in the 2.1 kb fragment which is missing in all disruptants (indicated by an arrow on the right). Other two bands are intact as predicted. All disruptants showed the same mutant phenotype in the plant assay as shown in Figure 23. Five disruptants (lanes 2, 3, 5, 7 and 8) resulted from a single crossover integration. Integration in two other disruptants (lanes 4 and 6) has not been determined. One of the disruptants (#118, lane 2) was used to recover the plasmid p118B14 and p118BC4.

Figure 25 displays a gel blot analysis showing targeted integration of the chromosome walking vector p118BSP into the wild type genome. Lane 1: wild type race O; Lane 2: wild type race T; Lanes 3, 4 and 5: three transformants obtained by transforming wild type race O (strain C5) with p118BSP. Genomic DNAs were digested with *Pst* I (which cuts pUCATPH) and probed with the 3.2 kb flanking genomic DNA fragment cloned on p214S1 which gives two bands in wild type (6.6 and 2.5 kb, indicated by arrows on the left). The targeted site is in the 6.6 kb region which is missing in all three disruptants (indicated by arrows on the right). The 2.5 kb band is intact as predicted. The third band (part of vector plus 4.6 kb genomic DNA) does not hybridize to the probe. All disruptants showed the same mutant phenotype in the plant assay as shown in Figure 23. Two disruptants (lanes 3 and 4) resulted from a double crossover integration. Integration in the third disruptant (lane 5) has not been determined. One of the disruptants (#9, lane 3) was used to recover the plasmid p9P2.

Figure 26 displays a gel blot showing the detection of *CPS1* homologs in *C. victoriae* and *C. carbonum*. Genomic DNAs were digested with *Bgl*II and probed with the 3.4 kb *CPS1* fragment cloned on p214B7 (Figure 2) which includes most of the 4.2 kb *Bgl*II fragment of *CPS1* (Figure 15). Lanes 1 and 2: *C. heterostrophus* race T (C4) and race O (C5), both of which hybridized to the 4.2 kb fragment. Lanes 3: *C. victoriae* (Hvw). Lanes 4, 5 and 6: *C. carbonum* race 1 (26R13), race 2 (YugY) and race 3 (BZ1209). Note that both *C. victoriae*

- 19 -

and *C. carbonum* (three races) hybridized to a 5.0 kb fragment and the hybridization signals of all three species showed about the same intensity.

Figure 27 is a high-performance liquid chromatography (HPLC) profile of culture extracts from wild type *C. victoriae* (HvW, top left) and three transformants (Tx7, Tx2 and Tx9). The major peak for victorin C is indicated by arrowheads. No significant differences in victorin production were detected between the wild type and transformants. Three other transformants (Tx4, Tx5 and Tx8) gave the same results.

Figure 28A-B are photographs which display a plant assay (Figure 28A) and a gel blot (Figure 28B), showing targeted disruption of the *CPSI* homolog in *C. victoriae*. Figure 28A shows *C. victoriae* transformants (Tx) with reduced or wild type pathogenicity to susceptible oats. Oat seeds were inoculated with conidial suspensions of (left to right) wild type, Tx7, Tx2 and Tx9 (the last pot on the right is the uninoculated control). Two transformants (Tx7 and Tx2) showed dramatically reduced pathogenicity as indicated by the substantial growth of the oat plants. These two transformants resulted from a homologous integration (lanes 2 and 3 in Figure 28B). Tx9, which killed all oat plants as wild type, resulted from a ectopic integration (lane 4 in Figure 28B). All three transformants produced wild type level of victorin as determined by HPLC analysis (Figure 27). Figure 28B shows disruption of *CPSI* homolog in the wild type genome. Genomic DNAs were digested with *Bgl*II and probed with the 3.2 kb *CPSI* fragment (*Kpn*I-*Sac*I) cloned on p214S1 (Figure 2) which hybridized to two fragments (4.2 and 2.2 kb, see Figure 15) in *C. heterostrophus* (lane 1), but to three fragments (5.0, 1.8 and 0.2 kb) in the *C. victoriae* (lane 2). In Tx7 (lane 3) and Tx2 (lane 4), one or two of the wild type fragments was replaced by a larger fragment (8.0 kb for Tx7 and 9.4 kb for Tx2) containing the transforming vector. The actual size increase by the vector integration can not be predicted because the presence of polymorphic bands in *C. victoriae* genome and the presence of duplicated *CPSI* fragments on the transforming vector (Figure 5). In Tx9 (lane 5), which caused wild type symptoms shown in Figure 28A, all three wild type bands (indicated by arrows) are intact, confirming an ectopic integration.

- 20 -

Figure 29 shows the REMI vector pUCATPH. This vector was constructed by insertion of a 2.4 kb *Sal*I fragment containing the selectable marker cassette (the largest arrow) from pDH25 (Cullen et al., 1987) into the *Sal*I site of polylinker site of pUC18 (between *lacI* and *lacZ*). Only six-base-pair restriction enzyme sites are shown on the map. Italicized sites are unique; three of them, *Hind*III, *Kpn*I, and *Sac*I (**bold**) have been used for REMI transformation. Non-cutting enzymes are listed below the map (enzymes that recognize six-base-pair sites are underlined). *amp*, Ampicillin resistance gene; *hygB*, hygromycin B resistance gene; *PtrpC*, *A. nidulans* *trpC* promoter; *TtrpC*, *A. nidulans* *trpC* terminator; *ori*, *Escherichia coli* origin of replication.

Figures 30A-30C are photographs of DNA gel blots showing DNA-DNA hybridization of *ChCPS1* to other fungal genera and species. In Figure 30A, the gel was loaded with *Cochliobolus* species (lanes 1-17) as follows: *C. heterostrophs* race T, race O; *C. carbonum* race 1, race 2; *C. victoriae* isolates FI3, HvW; *C. bicolor*, *C. dactyloctenii*, *C. chloridis*, *C. homomorphus*, *C. intermedius*, *C. melinidis*, *C. melinidis*, *C. peregranensis*, *C. perotidis*, *C. ravenelii* and *C. sativus*. Figure 30B is a photograph of a DNA gel blot from a gel loaded with other *Ascomycete* genera (lanes 1-14) as follows: *C. carbonum* race 1 (control), *Setosphaeria rostrata*, *Stemphyllium spp.*, *Pyrenophora tritici repentis*, *Bipolaris sacchari*, *Alternaria spp.*, *A. solani*, *Nectria haematococca*, *Fusarium oxysporum*, *Glomerella spp.*, *Magnaporthe grisea*, *F. moniliforme*, *F. moniliforme* (repeat) and *A. solani* (repeat). Figure 30C is a photograph of a DNA gel blot from a gel comparing *Candida albicans* to *C. heterostrophus* and closely related species (lanes 1-7): *C. heterostrophs* race T, *Bipolaris sacchari*, *Setosphaeria rostrata*, *Stemphyllium spp.*, *Pyrenophora tritici repentis*, *Alternaria spp.* and *Candida albicans* (arrowhead). Genomic DNAs were digested with *Hind*III (A, lanes 1-17; B, lanes 1-11; C, lanes 1-7), *Xba*I (B, lanes 12 and 14) or *Bgl*II (B, lane 13) and probed with the 3.2 kb fragment of *CPS1* from p214S1 (Fig. 2) at high stringency. Weak signals in lanes 3 and 17 (panel A) are due to insufficient DNA loading (confirmed by a repeat experiment).

Figure 31A is a structural comparison of the four *CPS1* homologs to *ChCPS1*. ORFs are indicated by the open boxes; shaded boxes inside indicate

- 21 -

functional domains; vertical bars indicate conserved motif sequences found in nonribosomal peptide synthetases (NRPS) as defined by Stachelhaus and Marahiel (Stachelhaus and Marahiel, 1995; Marahiel, 1997) (dashed bars indicate weak conservation). The black bulbs indicate the position of putative introns. Cores 1-5: 5 adenylation; core 6: thioaltung; TE: thioesterase. The distance between core sequences is not drawn in exact scale. The name of proteins is on the left of the ORF boxes and the number of amino acids on the right. The unidentified regions of *AsCPS1* and *PtCPS1* are indicated by dash-lined boxes. The similarity to *ChCPS1* (in the overlapping region only, see text for details) is given in the 10 parentheses under the protein names in the order: nucleotide identity/ amino acid identity/ amino acid similarity. The positions of the ChCPS1 amino acid 1040 is indicated by the open arrow; the positions 511 and 1269 (to the first and the last amino acids of AsCPS1 and PtCPS1) are indicated by filled triangles. Figure 31B is an amino acid alignment of the four CPS1 homologs to ChCPS1. 530 amino 15 acids aligned to the amino acids 511-1040 of ChCPS1 (shown in A) are shown. The identical residues are in uppercase and the similar residues in lowercase. Consensus of sequences similar to the typical NRPS signature motifs is underlined. The putative cyclization domain motif "DXXXXD/ EXXS/ A" is underlined.

20 Figure 32 is the nucleotide sequence of *FgCPS1*. 6,003 base pairs cloned using the plasmid rescue procedure are shown. The amino acid sequence of *FgCPS1* protein is given below the DNA sequence. The position of the start codon ATG (bold and underlined) is designated +1 and the open reading frame stops at position 5123 (TGA, bold and underlined). A "CT" motif (italicized and underlined) and two putative "CAAT" boxes (bold with asterisks) are found at positions -30, -204 and -302. A putative intron (in lowercase with 5' and 3' splice sequences in bold; branch sites underlined) is located at positions 4245-4290. 25 Conserved core sequences are shaded and the putative cyclization domain motif "DXXXXEXXA" (position 2323-2346) is underlined. A putative polyadenylation signal "AATAA" at position 5188 is bold and overlined.

30

Figure 33 shows the nucleotide sequence of *AsCPS1*. 2,369 base pairs amplified by PCR are shown. The amino acid sequence of *AsCPS1* protein is

- 22 -

given below the DNA sequence. The sequence is not complete. Two putative introns (in lowercase with 5' and 3' splice sequences in bold; branch sites underlined) are located at positions 540-584 and 2012-2059. Conserved core sequences are shaded and the putative cyclization domain motif “DXXXXDXXS” (positions 694-720) is underlined. The PCR primer binding sites at the 5' and 3' end are underlined.

Figure 34 shows the nucleotide sequence of *PtCPS1*. 2,320 base pairs amplified by PCR are shown. The amino acid sequence of PtCPS1 protein is given below the DNA sequence. The sequence is not complete. A putative intron (in lowercase with 5' and 3' splice sequences in bold; branch sites underlined) is located at positions 540-583. Conserved core sequences are shaded and the putative cyclization domain motif “DXXXXDXXS” (positions 693-719) is underlined. The PCR primer binding sites at the 5' and 3' end are underlined.

Figure 35A is a photograph of a DNA gel blot showing that the 15 2.2 kb wild type band (arrowhead) is disrupted in homologous transformants TxFgC8-4, -10, -11 and -5 but is intact in the ectopic transformant TxFgC8-H1, - H2 and -B1 [generated using *HindIII* (H) or *BglII*(B)-digested pFgC8-hygB]. Genomic DNAs were digested with *ClaI* and probed with pFgC8 which carries a 1.0 kb *FgCPS1* fragment. Figure 35B is a photograph of plants from a virulence 20 assay showing *F. graminearum* transformants (Tx) with reduced or wild type virulence to wheat. Wheat heads were inoculated with conidial suspensions (10⁴/ml) of (left to right) wild type, TxFgC8-4, -10, -H1, and -11 or with water only. Photograph was taken 7 days after inoculation. Note that most spikelets of wheat heads inoculated with homologous transformants looked “healthy” in contrast to those inoculated with ectopic transformants that were completely 25 “bleached” (indistinguishable from wild type).

DETAILED DESCRIPTION OF THE INVENTION

The present invention generally relates to an isolated DNA molecule from a plant pathogen encoding a CPS1 peptide synthetase. In one 30 embodiment, the DNA molecule has a nucleotide sequence which hybridizes to a DNA molecule having a sequence corresponding to SEQ. ID. No. 2 as follows:

- 23 -

	TGCCTGCGCC	TGTGCTTGTG	CCTGTGGAAT	GTCGGGGCCC	GCTGCTGCAT	AGCCTATCTG	60
	TACATACAAC	ACCATCCCAT	CCCGCTTCAC	CTGCCTTGCC	TCCCTCCTCG	TGCCACACAT	120
	CCGCCGCCA	CAACACCATG	GCTGCGACCA	ACCCCCAGCT	GCAGGGCCAAA	CTGCAGGAGC	180
	TGGACCACGA	GCTCGAGGAG	GGCGATATT	CACAAAAAGG	GTCCGTACTG	CTGCACCACC	240
5	ACGCCATCC	GCCTCTCTGC	GTGCGCTAAT	CAGTCGCATA	GCTATGAAAA	ACGTCGCACC	300
	GTGCTGCTGT	CGCAGTATCT	AGGGCCTGAC	TTTGTGCCC	AGTTGCAGGC	CGACCTGAAC	360
	CAGCAGAAC	CACCCCAACC	ATCCAGTGAG	GGCTCTCGCT	CCCGCACCGC	ATCCTTGCT	420
	ATTCCGTCG	GTCCGAGTCC	ATCACNGCGA	CCACAACCCC	CACATATCCA	GCTCCCCCGC	480
	CCCGACTCAT	ACCATGACGC	TTCCGCACAG	GGCCAATTGG	GCGCACCCAT	GCCATATGCC	540
10	AACGCCCTCG	CCGCTGCCTC	GGGGGGCTCG	CAGTACATGG	CATACCCGCC	CAGCCAAGTC	600
	GGCCGTTTC	AAGAGAACGA	GCTGGGCCTG	CGTACAAATT	CGCTCCAGCG	CAATTCCCTCA	660
	CAGCTGTCG	AAGGAAGCGA	GACGTTCATT	CCACGGCCTC	AAACGCCCTGA	ATACAACCAC	720
	TCGCGCGAGC	CCACCATGAT	GGGCAACTAC	GCCTTCAATC	CAGACAATCA	GCAAAGTTAT	780
	GATGGCCAAT	TTGGCTCTCC	GGGAGAGGCC	AGTCGAAGGA	GCACCATGCT	CGAGGTAAAC	840
15	CAGGGTTATT	TTTCCGACTT	CACAGGCCAG	CAGATGCAAG	ACAATCGCGA	CTCGTATGGG	900
	GGACCCAACC	GCTACTCGTC	GGGAGATGCC	TTTTCTCCTA	CCGCCGCGAT	TCCACCTCCC	960
	ATGATGAACC	CCAACGATCT	CCCCTTGGGC	GCTGCTGAAA	CCATGATGCC	GCTAGAGCCC	1020
	CGCGATCTGC	CTTTTGACGT	TTACGACCCCT	CACAACCCC	ATGTCAAAAT	GTCAAAGTTT	1080
	GACAACATTG	GCGCTGTCTT	GCGTCACCGA	AGTCGCACAC	AGCCAAGGAC	GAUTGCCTTC	1140
20	TGGGTCTTG	ACGCAAAAGG	CAAAGAGACG	GCGTCCATCA	CCTGGGAAAA	GGTGGCTAGT	1200
	CGCGCGGAAA	AGGTGGCCAA	AGTGATTGCG	GACAAGAGCA	ACCTCTATCG	AGGCAGCGT	1260
	GTGGCATTAG	TGTACAGGG	TACAGAAATC	ATTGATTTG	TCGTGGCGTT	GATGGGCTGC	1320
	TTCATTGCGG	GCGTTGTAGC	GGTACCCATC	AATAGCGTCG	ACGACTACCA	GAAACTCATT	1380
	CTTCTCCTAA	CGACAACCTCA	AGCTCATCTC	GCATTGACCA	CAGACAACAA	TCTCAAGGCC	1440
25	TTTCATCGTG	ACATTAGTC	GAACCGTCTG	AAATGGCCGA	GTGGGGTAGA	GTGGTGGAAAG	1500
	ACGAACGAGT	TTGGCAGCCA	CCACCCCAAG	AAACATGACG	ATACTCCAGC	TTTGCAAGTA	1560
	CCAGAGGTTG	CCTATATTGA	GTTCTCGCGT	GCACCTACTG	GTGACCTTCG	CGGTGTGGTG	1620
	CTTAGTCACC	GGACTATTAT	GCACCAAATG	GCCTGCATCA	GTGCCATGAT	TAGCACGATA	1680

- 24 -

	CCACCAACG CTCAGAGCCA AGACACGTT AGCACTAGCC TACGGGATGC AGAGGGAAAG	1740
	TTCGTTGCTC CAGCACCGTC CAGAAACCCC ACAGAAAGTGA TCCTCACGTA CCTCGACCCG	1800
	CGCGAAAGCG CTGGTCTCAT TCTCAGTGTC TTGTTGCGAG TTTATGGAGG CCACACCACC	1860
	GTATGGCTCG AGACAGCGAC CATGGAAACC CGGGGTCTAT ATGCACATCT CATCACCAAA	1920
5	TACAAGTCCA ACATACTGCT AGCGGATTAC CCAGGCCCTCA AGCGCGCTGC ATACAACCTAC	1980
	CAACAGGATC CAATGGCTAC AAGAAACTTC AAGAAAAACA CAGAACCCAA CTTCGCCTCC	2040
	GTGAAGATCT GTCTGATTGA CACGCTTACC GTCGACTGTG AATTCACGA AATTCTCGGA	2100
	GATCGATATT TCAGGCCACT GCGAAACCCCT AGAGCGCGAG AACTGATCGC GCCAATGCTC	2160
	TGCTTGCAG AACATGGTGG AATGATAATA TCTGTACGCG ACTGGCTAGG TGGAGAGGAG	2220
10	CGCATGGGCT GCCCGCTAAG CATAGCAGTA GAAGAGTCAG ATAATGATGA AGATGATACA	2280
	GAGGATAAGT ATGCAGCGGC AAATGGCTAC TCCAGTCTTA TTGGTGGTGG CACTACAAAG	2340
	AACAAAAAAGG AGAAGAAGAA GAAAGGCCCG ACAGAGCTTA CAGAAATCTT GCTGGACAAG	2400
	GAAGCTCTGA AGATGAACGA AGTCATTGTT CTGGCCATTG GAGAAGAAGC AAGCAAGCG	2460
	GCAAACGAGC CCGGCACCAT GCGAGTCGGT GCCTTGGAT ACCCCATACC GGATGCGACA	2520
15	CTAGCTATTG TAGACCCCTGA GACAAGTCTT CTATGTTCAC CATACTCGAT AGGCGAGATC	2580
	TGGGTAGATT CGCCTTCACT CTCTGGTGGC TTCTGGCAGC TGAGAAGCA TACAGAGACC	2640
	ATTTCCATG CTCGACCATA CCGTTTCGTT GANGTAGCC CTACGCCACA GTTGCTTGA	2700
	CTCGAGTTTC TCGGTACTGG ACTCCTCGGC TTTGTTGTAG ANGGAAAAT ATTTGTCCTT	2760
	GGACTGTACG AAGATCGCAT CAGACAGCGT GTTGAATGGG TAGAAAATGG TCAGCTTGA	2820
20	GCCGAGCATC GATACTTTT TGTGCAGCAC CTGGTCACAA GCATTATGAA GGCGTGCCA	2880
	AAAATTACG ACTGGTAAGT GAGCTGCCAA CAGAGCAAGG ACTGTCTAAC GTGTCATAGC	2940
	TCGTCGTTTG ATTCTTATGT AAATGGTGA TACCTGCCAA TCATTCTCAT CGAGACGCAG	3000
	GCCGCATCGA CTGCGCCCAC AAACCCAGGT GGACCACCC AACAAATGGA TATACCATT	3060
	TTGGATTCAC TATCTGAGAG GTGCATGGAG GTCCCTTACC AAGAGCATCA TTTACGGGT	3120
25	TACTGCGTGA TGATTACAGC ACCTAATACA CTTCCACGAG TCATCAAGAA CGGACGGCGA	3180
	GAAATTGGCA ATATGCTGTG TANGAGANAG TTTGACAATG GCTCTCTGCC CTGTGTNCAC	3240
	GTNAAGTTG GCATTGAGGG ATCAGTGCAG AACATTGCGC TCGGTGACGA TCCCGCTGGC	3300
	GGCATGTGGT CATTGAGGC ATCAATGGCA CGTCAGCAAT TCTTGATGCT CCAAGACAAG	3360

- 25 -

	CAATACTCTG GTGTCGATCA TCGCGAAGTC GTCATTGACG ACAGGACATC GACTCCACTC	3420
	AATCAGTTCT CGAATATCCA CGACCTGATG CAATGGCGTG TATCTCGGCA GGCGAGGAA	3480
	CTTGCCTACT GCACGTGCGA CGGTCGAGGA AAAGAGGGCA AAGGCCCAA TTGGAAGAAC	3540
	TTTGATCAA AGGTTGCGGG CGTAGCAATG TACCTCAAGA ACAAGGTCAA GGTCCAGGCC	3600
5	GGCGATCATC TCCTTCTGAT GTACACGCAT TCAGAAGAAAT TTGTTTATGC TGTCATGCA	3660
	TGTTTGTGC TTGGAGCTGT TTGCATACCA ATGGCGCAA TTGATCAGAA CCGGTTGAAT	3720
	GAGGATGCGC CGGCCTTGCT GCATATCCT GCAGATTCA AGGTCAAAGC CATTCTGTC	3780
	AACGCTGACG TTGACCATCT GATGAAGATC AAGCAAGTAT CGCAGCACAT CAAACAATCG	3840
	GCCGCTATCC TCAAGATCAG TGTGCCAAC ACATACAGCA CAACAAAGCC GCCAAAGCAA	3900
10	TCCAGTGGCT GCGCGACCT CAAGCTTACA ATTGACCGG CATGGATTCA GGCGGGTTTC	3960
	CCAGTGCTAG TCTGGACATA CTGGACGCC GATCAACGTC GTATGCAGT TCAGCTGGC	4020
	CATAGCCAAA TCATGGCACT GTGCAAGGTC CAAAAAGAAA CATGCCAAAT GACAAGTACA	4080
	CGACCAGTCC TTGGTTGTGT CCGGAGCACG ATAGGACTTG GTTCCCTCA CACTTGTCTC	4140
	ATGGGAATCT TCCTTGCCGC ACCCACATAC CTGGTGTAC CTGTTGACTT TGCACAAAAC	4200
15	CCTAATATTC TGTTCCAAAC GCTTCGCGG TACAAGATCA AGGATGCATA TGCAACGAGT	4260
	CAAATGTTGG ACCACGCCAT CGCACGCGGA GCTGGTAAGA GTATGGCTCT GCACGAGCTG	4320
	AAGAATCTCA TGATTGCGAC TGATGGAAGA CCACCGTTG ATGTTGTAA GTGAACATT	4380
	GTATGAGAGG ACTTCATGA TTGCTAACTC AATGCAGACC AAAGAGTGC G TGTGCACTTT	4440
	GCGCCAGCCA ACTTAGACCC AACCGCAATC AACACTGTCT ACTCACATGT ATTGAACCCA	4500
20	ATGGTAGCAT CACGATCATA CATGTGTATT GAGCCAGTCG AGCTCCATCT CGATGTGCAT	4560
	GCTCTGCGAC GCGGCCTCGT CATGCCCGTT GACCTGACA CAGAGCCCAA CGCTTGCTC	4620
	GTCCAAGACT CGGGCATGGT GCCAGTGAGC ACGCAAATAT CCATTGTCAA CCCAGAGACC	4680
	AACCAACTGT GCTTGAACGG CGAGTACCGC GAGATCTGGG TGCAGTCCGA GGCGAATGCT	4740
	TATAGCTTCT ACATGTCGAA AGAGCGCTTG GATGCAGAAC GCTTCATGG GAGGACGATT	4800
25	GACGGAGACC CAAATGTGCG ATATGTTCGT ACAGGGCATT TAGGATTTT GCACAGCGTG	4860
	ACACGGCCCA TTGGACCCAA CGGTGCACCT GTTGATATGC AGGTGCTTT CGTGCTTGG	4920
	AGCATAGGTG ACACTTTGAGTCAACCGA CTGAACCATT TCTCTATGGA CATTGAGCAG	4980
	TCTGTTGAAC GTTGTACCCG GAATATTGTC CCTGGAGGCT GGTACGTTTC TTCGATTGCC	5040

- 26 -

	TGTTATTTAG TAAATACTTA CTAACACTCT ACAGTGCTGT TTTCCAGGCA GGTGGGCTTG	5100
	TTGTTGTCGT TGTGGAATC TTCCGACGCA ACTTCCTCGC AAGCATGGTG CCTGTGATTG	5160
	TCAATGCAAT TTTGAACGAG CATCAGCTGG TCATTCACAT TGTCTCGTT GTGCAAAGG	5220
	GCGACTTCCA CCGGTCTCGT CTGGGCAGA AGCAACGCGG AAAGATTCTT GCAGGATGGG	5280
5	TCACACGGAA GATGCGCACA ATAGCCCAGT ACAGTATAACG GGATCCTAAT GGACAGGATT	5340
	CCCAGATGAT CACGGAAGAG CCTGGTCCAC GGGCTAGATG ACTGGAAGTA TGCTTGGCG	5400
	AATGGGCGGC CCAGCCAGTA TCAAGGCCGG GTGCACAAGA GCACCGAGTC TAATGGGCAT	5460
	GACAGCGACT ATGAATAATC TATCCCTTAC ACAGCAGCAA CAGCAGCAAT ACCAACAGCC	5520
	GGGTATGTAT GCTCAACAGC AAGGCATGCA CCCCCAGCAA CAACACCAAT TTAGCATGTC	5580
10	CAACACGCCA CCACAAGGTC CACCCCAAGG CGTAGAACTA CATGATCCTA GCGACCGCAC	5640
	ACCAACAGAC AACCGGCAC TTTCCCTTGC CGACCCCGT ATGCAGAACCC AGGGCCAAAT	5700
	GAACGAGACG GGCGCCTACG AACCCATGAA CTATCAAAAC GCGTATCATC CGCATCAACA	5760
	ACAATACGAA TCTGAAGACG GGGGGAGCAG ACTCAGCGC CCCGTGCCAG ACGTGCTGCG	5820
	GCCGGGTCCCT TCATCCGGGT CCATAGAGCA GCACGACCAA GCTAACAAACG ACAACAATAT	5880
15	GTGGAATAAT CGCGAGTACT ATGGTAACAG CCCATCGTAT GCAGGGCGGAT ACACGCAAGA	5940
	TGGCAATATC CACGAGCAGC AACAAACACGA TGAGTACACG AGTAATGCGT CATATGGCG	6000
	AAATCAAGGA GCAGGGCGGAG GCAGCGGGCG CGGTGGCGGT CTCCGAGTTG CAAATCGTGA	6060
	CAGCTCCGAC AGCGAGGGTG CAGATGACGA CGCTTGGAGA CGTGATGCC CCCGTGCCAG	6120
	CAATTTGCG GGCAGCGCTG CTGCTGCCTC CGCTGGAGCA CCTGCTGCTG GTGCTTCTTC	6180
20	TTCGCAGCCG GGCCATGCGC AGTAGACGGG ATATGCGTGA GTTTTTTTT AAATTCGTA	6240
	CATAGAGACC GTTGTATACG CAGGTTCAA ATTAGAAGAG CGAATATGCA TATCAGCTGT	6300
	TGTTCAATGT TCTAGTTGG GAAGGTTAAC CCCCCCCCCCT TCCCTTCCA AGACTTTCA	6360
	CTTGTGTTG TGATGTTAA ATCTGGAGAT TTCAAATCTA CATCTCGCTA TACATAGGTG	6420
	TTGTTGATA ACGTAGGGGG CAGAAGGGTA TCTCGTGATA TTAGACTGGG AGTTGCATGA	6480
25	ATCAAGGTGT TGAGCAAAAA AAGAGAGAGC GGTGAAGGGC GGGGGGATA GGTGGTGTGC	6540
	ACGTGGCTG	6549

In a preferred embodiment, the plant pathogen is *Cochliobolus heterostrophus*. In another preferred embodiment, the plant pathogen is

- 27 -

Cochliobolus carbonum, C. victoriae, C. sativus, C. specifer, C. homomorphus, C. dactyloctenii, Setosphaeria turcica, S. rostrata, or Bioplaris sacchari.

The peptide synthetase of SEQ. ID. No. 2 has a deduced amino acid sequence corresponding to SEQ. ID. No. 3 as follows:

5	Met Leu Glu Val Asn Gln Gly Tyr Phe Ser Asp Phe Thr Gly Gln	15
	Gln Met Gln Asp Asn Arg Asp Ser Tyr Gly Gly Pro Asn Arg Tyr	30
	Ser Ser Gly Asp Ala Phe Ser Pro Thr Ala Ala Ile Pro Pro Pro	45
	Met Met Asn Pro Asn Asp Leu Pro Leu Gly Ala Ala Glu Thr Met	60
	Met Pro Leu Glu Pro Arg Asp Leu Pro Phe Asp Val Tyr Asp Pro	75
10	His Asn Pro Asn Val Lys Met Ser Lys Phe Asp Asn Ile Gly Ala	90
	Val Leu Arg His Arg Ser Arg Thr Gln Pro Arg Thr Thr Ala Phe	105
	Trp Val Leu Asp Ala Lys Gly Lys Glu Thr Ala Ser Ile Thr Trp	120
	Glu Lys Val Ala Ser Arg Ala Glu Lys Val Ala Lys Val Ile Arg	135
	Asp Lys Ser Asn Leu Tyr Arg Gly Asp Arg Val Ala Leu Val Tyr	150
15	Arg Asp Thr Glu Ile Ile Asp Phe Val Val Ala Leu Met Gly Cys	165
	Phe Ile Ala Gly Val Val Ala Val Pro Ile Asn Ser Val Asp Asp	180
	Tyr Gln Lys Leu Ile Leu Leu Leu Thr Thr Thr Gln Ala His Leu	195
	Ala Leu Thr Thr Asp Asn Asn Leu Lys Ala Phe His Arg Asp Ile	210
	Ser Gln Asn Arg Leu Lys Trp Pro Ser Gly Val Glu Trp Trp Lys	225
20	Thr Asn Glu Phe Gly Ser His His Pro Lys Lys His Asp Asp Thr	240
	Pro Ala Leu Gln Val Pro Glu Val Ala Tyr Ile Glu Phe Ser Arg	255
	Ala Pro Thr Gly Asp Leu Arg Gly Val Val Leu Ser His Arg Thr	270
	Ile Met His Gln Met Ala Cys Ile Ser Ala Met Ile Ser Thr Ile	285
	Pro Thr Asn Ala Gln Ser Gln Asp Thr Phe Ser Thr Ser Leu Arg	300
25	Asp Ala Glu Gly Lys Phe Val Ala Pro Ala Pro Ser Arg Asn Pro	315
	Thr Glu Val Ile Leu Thr Tyr Leu Asp Pro Arg Glu Ser Ala Gly	330
	Leu Ile Leu Ser Val Leu Phe Ala Val Tyr Gly Gly His Thr Thr	345
	Val Trp Leu Glu Thr Ala Thr Met Glu Thr Pro Gly Leu Tyr Ala	360
	His Leu Ile Thr Lys Tyr Lys Ser Asn Ile Leu Leu Ala Asp Tyr	375

- 28 -

	Pro Gly Leu Lys Arg Ala Ala Tyr Asn Tyr Gln Gln Asp Pro Met	390
	Ala Thr Arg Asn Phe Lys Lys Asn Thr Glu Pro Asn Phe Ala Ser	405
	Val Lys Ile Cys Leu Ile Asp Thr Leu Thr Val Asp Cys Glu Phe	420
	His Glu Ile Leu Gly Asp Arg Tyr Phe Arg Pro Leu Arg Asn Pro	435
5	Arg Ala Arg Glu Leu Ile Ala Pro Met Leu Cys Leu Pro Glu His	450
	Gly Gly Met Ile Ile Ser Val Arg Asp Trp Leu Gly Gly Glu Glu	465
	Arg Met Gly Cys Pro Leu Ser Ile Ala Val Glu Glu Ser Asp Asn	480
	Asp Glu Asp Asp Thr Glu Asp Lys Tyr Ala Ala Asn Gly Tyr	495
	Ser Ser Leu Ile Gly Gly Thr Thr Lys Asn Lys Lys Glu Lys	510
10	Lys Lys Lys Gly Pro Thr Glu Leu Thr Glu Ile Leu Leu Asp Lys	525
	Glu Ala Leu Lys Met Asn Glu Val Ile Val Leu Ala Ile Gly Glu	540
	Glu Ala Ser Lys Arg Ala Asn Glu Pro Gly Thr Met Arg Val Gly	555
	Ala Phe Gly Tyr Pro Ile Pro Asp Ala Thr Leu Ala Ile Val Asp	570
	Pro Glu Thr Ser Leu Leu Cys Ser Pro Tyr Ser Ile Gly Glu Ile	585
15	Trp Val Asp Ser Pro Ser Leu Ser Gly Gly Phe Trp Gln Leu Gln	600
	Lys His Thr Glu Thr Ile Phe His Ala Arg Pro Tyr Arg Phe Val	615
	Xaa Gly Ser Pro Thr Pro Gln Leu Leu Glu Leu Glu Phe Leu Arg	630
	Thr Gly Leu Leu Gly Phe Val Val Glu Gly Lys Ile Phe Val Leu	645
	Gly Leu Tyr Glu Asp Arg Ile Arg Gln Arg Val Glu Trp Val Glu	660
20	Asn Gly Gln Leu Glu Ala Glu His Arg Tyr Phe Phe Val Gln His	675
	Leu Val Thr Ser Ile Met Lys Ala Val Pro Lys Ile Tyr Asp Cys	690
	Ser Ser Phe Asp Ser Tyr Val Asn Gly Glu Tyr Leu Pro Ile Ile	705
	Leu Ile Glu Thr Gln Ala Ala Ser Thr Ala Pro Thr Asn Pro Gly	720
	Gly Pro Pro Gln Gln Leu Asp Ile Pro Phe Leu Asp Ser Leu Ser	735
25	Glu Arg Cys Met Glu Val Leu Tyr Gln Glu His His Leu Arg Val	750
	Tyr Cys Val Met Ile Thr Ala Pro Asn Thr Leu Pro Arg Val Ile	765
	Lys Asn Gly Arg Arg Glu Ile Gly Asn Met Leu Cys Arg Arg Glu	780
	Phe Asp Asn Gly Ser Leu Pro Cys Val His Val Lys Phe Gly Ile	795
	Glu Arg Ser Val Gln Asn Ile Ala Leu Gly Asp Asp Pro Ala Gly	810
30	Gly Met Trp Ser Phe Glu Ala Ser Met Ala Arg Gln Gln Phe Leu	825

- 29 -

	Met Leu Gln Asp Lys Gln Tyr Ser Gly Val Asp His Arg Glu Val	840
	Val Ile Asp Asp Arg Thr Ser Thr Pro Leu Asn Gln Phe Ser Asn	855
	Ile His Asp Leu Met Gln Trp Arg Val Ser Arg Gln Ala Glu Glu	870
	Leu Ala Tyr Cys Thr Val Asp Gly Arg Gly Lys Glu Gly Lys Gly	885
5	Val Asn Trp Lys Lys Phe Asp Gln Lys Val Ala Gly Val Ala Met	900
	Tyr Leu Lys Asn Lys Val Lys Val Gln Ala Gly Asp His Leu Leu	915
	Leu Met Tyr Thr His Ser Glu Glu Phe Val Tyr Ala Val His Ala	930
	Cys Phe Val Leu Gly Ala Val Cys Ile Pro Met Ala Pro Ile Asp	945
	Gln Asn Arg Leu Asn Glu Asp Ala Pro Ala Leu Leu His Ile Leu	960
10	Ala Asp Phe Lys Val Lys Ala Ile Leu Val Asn Ala Asp Val Asp	975
	His Leu Met Lys Ile Lys Gln Val Ser Gln His Ile Lys Gln Ser	990
	Ala Ala Ile Leu Lys Ile Ser Val Pro Asn Thr Tyr Ser Thr Thr	1005
	Lys Pro Pro Lys Gln Ser Ser Gly Cys Arg Asp Leu Lys Leu Thr	1020
	Ile Arg Pro Ala Trp Ile Gln Ala Gly Phe Pro Val Leu Val Trp	1035
15	Thr Tyr Trp Thr Pro Asp Gln Arg Arg Ile Ala Val Gln Leu Gly	1050
	His Ser Gln Ile Met Ala Leu Cys Lys Val Gln Lys Glu Thr Cys	1065
	Gln Met Thr Ser Thr Arg Pro Val Leu Gly Cys Val Arg Ser Thr	1080
	Ile Gly Leu Gly Phe Leu His Thr Cys Leu Met Gly Ile Phe Leu	1095
	Ala Ala Pro Thr Tyr Leu Val Ser Pro Val Asp Phe Ala Gln Asn	1110
20	Pro Asn Ile Leu Phe Gln Thr Leu Ser Arg Tyr Lys Ile Lys Asp	1125
	Ala Tyr Ala Thr Ser Gln Met Leu Asp His Ala Ile Ala Arg Gly	1140
	Ala Gly Lys Ser Met Ala Leu His Glu Leu Lys Asn Leu Met Ile	1155
	Ala Thr Asp Gly Arg Pro Arg Val Asp Val Tyr Gln Arg Val Arg	1170
	Val His Phe Ala Pro Ala Asn Leu Asp Pro Thr Ala Ile Asn Thr	1185
25	Val Tyr Ser His Val Leu Asn Pro Met Val Ala Ser Arg Ser Tyr	1200
	Met Cys Ile Glu Pro Val Glu Leu His Leu Asp Val His Ala Leu	1215
	Arg Arg Gly Leu Val Met Pro Val Asp Pro Asp Thr Glu Pro Asn	1230
	Ala Leu Leu Val Gln Asp Ser Gly Met Val Pro Val Ser Thr Gln	1245
	Ile Ser Ile Val Asn Pro Glu Thr Asn Gln Leu Cys Leu Asn Gly	1260
30	Glu Tyr Gly Glu Ile Trp Val Gln Ser Glu Ala Asn Ala Tyr Ser	1275

- 30 -

	Phe Tyr Met Ser Lys Glu Arg Leu Asp Ala Glu Arg Phe Asn Gly	1290
	Arg Thr Ile Asp Gly Asp Pro Asn Val Arg Tyr Val Arg Thr Gly	1305
	Asp Leu Gly Phe Leu His Ser Val Thr Arg Pro Ile Gly Pro Asn	1320
	Gly Ala Pro Val Asp Met Gln Val Leu Phe Val Leu Gly Ser Ile	1335
5	Gly Asp Thr Phe Glu Val Asn Gly Leu Asn His Phe Ser Met Asp	1350
	Ile Glu Gln Ser Val Glu Arg Cys His Arg Asn Ile Val Pro Gly	1365
	Gly Cys Ala Val Phe Gln Ala Gly Gly Leu Val Val Val Val	1380
	Glu Ile Phe Arg Arg Asn Phe Leu Ala Ser Met Val Pro Val Ile	1395
	Val Asn Ala Ile Leu Asn Glu His Gln Leu Val Ile Asp Ile Val	1410
10	Ser Phe Val Gln Lys Gly Asp Phe His Arg Ser Arg Leu Gly Glu	1425
	Lys Gln Arg Gly Lys Ile Leu Ala Gly Trp Val Thr Arg Lys Met	1440
	Arg Thr Ile Ala Gln Tyr Ser Ile Arg Asp Pro Asn Gly Gln Asp	1455
	Ser Gln Met Ile Thr Glu Glu Pro Gly Pro Arg Ala Ser Met Thr	1470
	Gly Ser Met Leu Gly Arg Met Gly Gly Pro Ala Ser Ile Lys Ala	1485
15	Gly Ser Thr Arg Ala Pro Ser Leu Met Gly Met Thr Ala Thr Met	1500
	Asn Asn Leu Ser Leu Thr Gln Gln Gln Gln Gln Tyr Gln Gln	1515
	Pro Gly Met Tyr Ala Gln Gln Gln Gly Met His Pro Gln Gln Gln	1530
	His Gln Phe Ser Met Ser Asn Thr Pro Pro Gln Gly Pro Pro Gln	1545
	Gly Val Glu Leu His Asp Pro Ser Asp Arg Thr Pro Thr Asp Asn	1560
20	Arg His Ser Phe Leu Ala Asp Pro Arg Met Gln Asn Gln Gly Gln	1575
	Met Asn Glu Thr Gly Ala Tyr Glu Pro Met Asn Tyr Gln Asn Ala	1590
	Tyr His Pro His Gln Gln Gln Tyr Glu Ser Glu Asp Gly Gly Ser	1605
	Arg Leu Ser Gly Pro Val Pro Asp Val Leu Arg Pro Gly Pro Ser	1620
	Ser Gly Ser Ile Glu Gln His Asp Gln Ala Asn Asn Asp Asn Asn	1635
25	Met Trp Asn Asn Arg Glu Tyr Tyr Gly Asn Ser Pro Ser Tyr Ala	1650
	Gly Gly Tyr Thr Ser Gln Asp Gly Asn Ile His Glu Gln Gln His	1665
	Asp Glu Tyr Thr Ser Asn Ala Ser Tyr Gly Gly Asn Gln Gly Ala	1680
	Gly Gly Gly Ser Gly Gly Gly Gly Leu Arg Val Ala Asn Arg	1695
	Asp Ser Ser Asp Ser Glu Gly Ala Asp Asp Asp Ala Trp Arg Arg	1710
30	Asp Ala Leu Ala Gln Ile Asn Phe Ala Gly Gly Ala Ala Ala Ala	1725

- 31 -

Ser Ala Gly Ala Pro Ala Ala Gly Ala Ser Ser Ser Gln Pro Gly	1740
His Ala Gln	1743

In another embodiment of the invention, there is provided an isolated nucleic acid molecule encoding a CPS1 peptide synthetase homolog and which hybridizes to a nucleic acid molecule having a sequence corresponding to SEQ ID No. 41 as follows :

AAGAAGAAAG GGCGGACCGA GTTGACCGAA ATATTGCTAG ATAAGGAAGC ACTGAAGCTG	60
AACGAAGTTG TTGTTTGGC CATTGGAGAG GAAGTGGAGCA AGCGTGTCAA CGAACCCGGC	120
ACTATGAGAG TCGGTGCTTT TGGCTACCCG ATACCAGATG CGACGCTGGC CGTCGTCGAT	180
CCGGAAACTA ATCTTTTGTG TTCACCTAT TCCATAGGAG AGATCTGGGT AGACTCGCCA	240
TCATTGTCCG GAGGGTTTG GCAGCTGCAG AAGCACACTG AGACTATTTT CCACGCTCGG	300
CCATATCGTT TCGTAGAGGG CAGCCCCAAC CCGCAACTAC TCGAACTGGA GTTTCTACGC	360
ACTGGACTGC TCGGATGCGT GGTAGAAGGC AAAATCTTCG TATTAGGCCT GTACGAGGAC	420
CGGATTAGGC AGCGCGTTGA ATGGGTAGAG CACGGTCAGC TAGAAGCCGA ACATAGGTAT	480
TTCTTCGTGC AGCATCTTG CACCAGCATT ATGAAAGCTG TTCCAAAGAT TTACGACTGG	540
TAAGTGCTAT CGAATCTCTG GGTAATCAAC CTAACATTGC GCAGCTCGTC TTTCGATTCC	600
TATGTCACG GCGAATACTT ACCAATCATC CTTATCGAGA CACAGGCCGC ATCAACTGCT	660
CCCACAAATC CAGGGGGGCC ACCACAACAA CTTGACATTG CTTTCCTAGA CTCTCTTCT	720
GAGCGATGTA TGGAGGTACT GTATCAAGAA CACCACCTTC GGGTGTATTG TGTGATGATC	780
ACTGCACCGA ACACACTCCC GCGAGTCATC AAGAACGGTC GACGAGAAAT TGGAAACATG	840
CTTGCCGGA GAGAATTGTA CAATGGCTCG CTACCCCTGCG TTCACGTCAA GTTTGGCGTC	900
GAGAGGTCGG TCCAGAAATAT TCGCCTAGGT GATGACCCCTG CTGGCGGCAT GTGGTCTTAC	960
GAGGGCGTCGA TGGCACGCCA GCAGTTCTG ATGCTTCAAG ATAAGCAGTA CTCTGGAGTA	1020
40 GATCACAGAG AAGTCGTTAT TGACGACAGA ACGTCGACGC CGCTCAACCA GTTCTCCAAC	1080
ATTCATGACC TTATGCAATG GCGCGTACAA CGACAAGCTG AAGAGCTCGC CTACTGCACG	1140
45 GTAGATGGTC GAGGTAAAGA GGGCAAAGGC GTCAACTGGA AGAAGTTCGA CCAGAAGGTC	1200
GCAGGTGTCG CCATGTACCT GAAGAACAAAG GTCAAGGGTC AGACTGGTGA CCACCTGCTC	1260
50 TTGATGTACA CCCACTCGGA AGACTTTGTC TATGCCGTAC ACGCGTGTGTT CGTCCTTGGA	1320
GCTGTGTGTA TACCCATGGC ACCAATCGAC CAGAACAGGC TAAATGAAGA CGCGCCCCGCA	1380
CTACTACATA TCATTGCTGA CTTCAAGGTC AAGGCTATCC TCGTCAATGC TGGCGTAGAC	1440
55 CACCTGATGA AGGTCAAGCA AGTATCGCAG CACATCAAAC AGTCAGCAGT CATTCTCAAG	1500
ATCAACGTAC CGAACATACCTA TAACACCACAA AAACCACCTA AGCAGTCTAG TGGTTGCCGC	1560

- 32 -

	GATCTTAAGC TCACAATACG ACCTGCTTGG ATACAATCTG GTTCCCTGT TCTAGTATGG	1620
	ACATACTGGA CACCTGACCA GAGACGCATA GCTGTCAAT TAGGTCATAG CCAAATCATG	1680
5	GCGCTATGCA AAGTTCAGAA AGAAACGTGC CAGATGACGA GCACACGGCC CGTCCTTGGA	1740
	TGTGTTCGTA GCACGATCGG TCTTGCTTC ATACACACCT GTGTTATGGG TATCTTCCTC	1800
10	GCAGCGCCAA CTTACCTTGT GTCACCTGTC GATTTGCGC AAAACCCGAA CATCCTCTTC	1860
	CAGACCATGT CGAGATACAA GATCAAGGAC GCGTATGCGA CCAGCCAAAT GCTGGACCAC	1920
	GCTATTGCAC GAGGTGCTGG CAAGAACATG GCTCTGCACG AGCTCAAGAA CCTCATGATC	1980
15	GCGACTGACG GTCGGCCGCG CGTAGACGTC TGTAAGTGT GCGATCCTGT ATAAGCATCT	2040
	GAAATCTAAT TCTTGATAGA CCAGCGTGTG CGAGTACACT TCTCGCCAGC AAGTTGGAC	2100
20	CGAACGGCAA TCAACTGT TTACTCACAC GTACTGAATC CTATGGTCGC ATCGCGGTCA	2160
	TACATGTGCA TCGAACCCAT AGAACTACAT CTCGATGTG GTGCCCTTCG AAGAGGTCTC	2220
	ATCATGCCTG TCGACCCAGA CACGGAACCT GGTGCTCTCT TAGTCCAGGA CTCGGGTATG	2280
25	GTACCAAGTTA GTACACAAAT TTCAATCGTG AATCCAGAGA CAAACCCAGCT TTGCCTAGTC	2340
	GGCGAGTATG GCGAAATCTG GGTCCAACC	2370

Preferably, the CPS1 nucleic acid molecule which hybridizes to a
 30 nucleic acid molecule having a sequence as set forth in SEQ ID No. 41 is from the
 plant pathogen *Altenaria solani*. In another preferred embodiment, the CPS1 gene
 is from a plant pathogen such as *Altenaria alternatherae*, *A. alternata*, *A.
 amaranthi*, *A. araliae*, *A. brassicae*, *A. brassicicola*, *A. camelliae*, *A. cassiae*, *A.
 cheiranthi*, *A. cinerariae*, *A. gossypii*, *A. helianthi*, *A. helianthinficiens*, *A. mali*,
 35 or *A. raphani*.

The peptide synthetase product of SEQ ID NO:41 has a deduced
 amino acid sequence as follows (SEQ ID NO:42):

	KKKGPTELTE ILLDKEALKL NEVVVLALIGE EVSKRVNEPG TMRVGAFGYP IPDATLAVVD	60
40	PETNLLCSPY SIGEIWVDSP SLSGGFWQLQ KHTETIFHAR PYRFVEGSPT PQLLELEFLR	120
	TGLLGCVVEG KIFVGLGLYED RIRQRVEWVE HGQLEAEHRY FFVQHLVTSI MKAVPKIYDC	180
45	SSFDSYVNGE YLPIIILIEIQ AASTAPTNPG GPPQQLDIPF LDSLSERCME VLYQEHHLRV	240
	YCVMITAPNT LPRVIKNGRR EIGNMLCRRE FDNGSLPCVH VKFGVERSVO NIALGDDPAG	300
	GMWSYEASMA RQQFLMLQDK QYSGVDHREV VIDDRSTPL NQFSNIHDLM QWRVQRQAE	360
50	LAYCTVDGRG KEGKGVNWK FDQKVAGVAM YLKNKVKGQT GDHLLLHYTH SEDFVYAVHA	420
	CFVLGAVCIP MAPIDQNRLN EDAPALLHII ADFKVKAILV NAGVDHLMKV KQVSQHIKQS	480

- 33 -

	AVILKINVPN TYNTTKPPKQ SSGCRDLKLT IRPAWIQSGF PVLVWTYWTP DQRRIAVQLG	540
	HSQIMALCKV QKETCQMTST RPVLGCVRST IGLGFIHTCV MGIFLAAPTY LVSPVDFAQN	600
5	PNILFQTMSR YKIKDAYATS QMLDHAIARG AGKNMALHEL KNLMIATDGR PRVDVYQRVR	660
	VHFSPASLDR TAINTVYSHV LNPMVASRSY MCIEPIELHL DVGALARGLI MPVDPDTEPG	720
	ALLVQDSGMV PVSTQISIVN PETNQLCLVG EYGEIWVQ	760
10		

In another embodiment of the invention, there is provided an isolated nucleic acid molecule encoding a CPS1 peptide synthetase homolog and which hybridizes to a nucleic acid molecule having a sequence corresponding to SEQ ID No. 43 as follows :

15	CTCGAGGTTA GTAAAAGATC CCCGTTTGT CCACAAATCT CCATCTCCCT CTCATGCCT	60
	TTCTTGGCGC CTCAACCCGC TATTTTGAAAG ACAGTTTGT GTTGTGCGAT GCGACCAAAA	120
	ATCATCCTCT CAAGTTTTCA TCGCTGACCT GTTCTTGGC TAGGAAAGGA GATATCACAC	180
20	AGAAAGGGTA AGCTGTTTG CGTCCAGAGT ACTTACAATT GCTTCTCAAT TACTTACGCG	240
	CCGGCAGCTA CCAAAGCGA CGAACTCAAC TTTCTCCCA ATTCTCGGT GCACCTCCAC	300
25	CTCAGATTGC TGCTCTGCC GAGCCTCAGT CTGGCCTACG CATAACTCG CCCGATGACT	360
	CCGACCACCC TTCAGGCGAT GGCCATCGCG CTACCGCTA TGCGCTCTC GGTAGCAGCA	420
	GCGGTCCAAT CCCAGATTCA CCAGACTCAC CTATGTACCG ACCGACTCT GGTTATGCTC	480
30	CTTCAGAAC ACCAAGACCT TCTCCAGCAC AACCTCCACC TTCCCTGCTG CGCCCGGGGG	540
	GTTCTCTCGC TGGAGGATCG ACCACTGCTC ACCGGACTC CCTCTTCTTC TCCCCCTCCC	600
35	ATCTCGAACC TGAAACCCGG ACAGGTACTA TGATGTCGGG CGACTATGCA TTCAGACCCG	660
	AGCAGCAAGG CACATATGGC GAATCCCAGC ATCAACAGCA CCAGTTCCAG CAACAGCAAC	720
40	AGCCACAGCA GCAACAGCAG TACGATGGGC AGCAGTATGA TGGACGAAC ACAACGCTTC	780
	TCGATTGCA AGGATACTTT TCGGATTGG CGGGACAGCA GCACTATGAT CAGACTCAA	840
	CCGTTGAGTA TGTGGGACCT CAGCAGCGGT ATTCTCCAG CGATGCGATTC TCTCCAACCG	900
45	CCGCAATGGC ACCTCCAATG CTTACAACCA ACGACCTCCC ACCGCCGGAA GCGCTTGAGT	960
	ACCAAGCTGCC CCTTGACCCCT CGCGAGGTAC CATTCGCTAT TCAAGATCCC CATGATGATT	1020
	CTACGCCAAT GTCAAAGTTC GATAACATCG CAGCTGTACT CAGACATAGA GGCGAACGA	1080
50	TTGCTAAGAA GCCGGCATAC TGGGTGTTGG ATAGTAAGGG CAAGGAGATT GCATCGATTA	1140
	CGTGGGATAA GCTGGCATCT AGAGCCGAAA AGGTTGCGCA AGTCATCCGC GACAAAAGCT	1200
55	CTCTGTACCG GGGTGATCGG GTTGCTCTCA TCTACCGCGA TTCAGAGGTT ATTGATTTCG	1260
	CCATTGCCTT GCTGGGATGC TTCATTGCTG GAGTTGTTGC CGTTCCCATC AATGATCTGC	1320
60	AGGACTACCA ACGCTTGAAAC CACATTCTTA CTACAACGCA GGCCCATCTA GCGCTGACCA	1380

- 34 -

	CCGATAACAA CCTCAAAGCC TTTCAACGAG ACATTACTAC ACAAAAGTTG ACATGGCAA	1440
	AGGGTGTGA ATGGTGAAG ACAAACGAGT TTGGCACTTA TCACCCCAAG AAGAAGGAGG	1500
5	ATGTCGGCGC TTGGTTGTT CCCGATCTGG CATATATCGA GTTTTCGCGG GCCCCAAC	1560
	GAGACTTGAG AGGTGTTGTT CTGAGCCACC GAACCAATTAT GCACCAAATG GCTTGTCTTA	1620
10	GTGCATTAT TTCTACTATC CCGGTAATG GACCTGGCGA CACTTCAAC CCGTCTCTTC	1680
	GCGACAAGAA TGGTCGACTT ATTGGTGGCG GCGCAAGCAG CGAAATTTG GTGTCGTACC	1740
	TCGATCCCCG TCAGGGCATT GGATGATTG TGAGCGTGT ACTGACCGTC TACGGCGGCC	1800
15	ACACCACTGT TTGGTTCGAC AACAAAGCTG TTGATGTTCC TGGACTGTAC GCCCACCTCC	1860
	TTACCAAGTA CAAATCGACC ATCATGATTG CCGACTACCC AGGATTGAAG CGAGCCGCCT	1920
	ACAACATCCA GCAAGAGCCA ATGGTGACCC GAAATTTAA GAAGGGAATG GAGCCAAACT	1980
20	TTCAATGAT CAAGCTTGC TTGATTGACA CCTTGACTGT AGACAGCGGG TCCCACGAAG	2040
	TTTGGCTGA CCGATGGCTA CGACCGTTGA GAAACCTCG TGCCCGTGAG GTTGTGCAC	2100
25	CTATGTTTG TCTACCTGAA CACGGAGGCA TGGTGATTAG TGTGCGTGAC TGGCTAGGAG	2160
	GAGAAGAGCG CATGGGATGC CCATTAAGC TTGAACTTGG GGAGGATACA GAGTCTGACG	2220
30	AAGAGAAAGA GGAAACAGAG AAGCCAGCAG TTTCAATGG CTTGGTAGT CTCTTGTCA	2280
	GTGGTGGCAC AGCAACAACC GAAGAGAGGG CAAAGAATGA GCTTGGCGAA GTCCCTTTGG	2340
	ATCGTGAGGC TCTAAAGACC AACGAAGTTG TGGTGGTGGC CATAGGTAAC GATGCCCGTA	2400
35	AAAGGGTGC GGATGACCCA GGCTGGTAC GGGTCGGTTC TTTGGATAC CCCATACCG	2460
	ATGCCACACT CTCCGTCGTC GATCCAGAAA CGGGTTACT GGCGTCACCA CATTCCGTGG	2520
	GTGAAATCTG GGTCGACTCC CCTTCTCTTT CAGGTGGTT CTGGCGCG CCAAAGAATA	2580
40	CTGAGCTGAT TTTCCATGCT CGTCCTTACA AGTTGACCC AGGTGATCCT ACACCGCAGC	2640
	CCGTCGAGCC CGAACCTCG CGAACAGGCT TGCTGGCAC CGTCATCGAG GGTAAAATCT	2700
45	TTGTTCTGGG CCTTTACGAA GACCGAATTG GACAAAGGT TGAGTGGTT GAGCATGGAC	2760
	ACGAACATAGC AGAGTACCGC TACTTCTTG TTCAGCACAT CGTTGTGAGC ATTGTCAAGA	2820
	ACGTTCCAAA GATATACGAT TGTTCAGCCT TTGACGTCTT TGTCAATGAC GAACACCTGC	2880
50	CAGTCGTGGT GCTGGAGTCA GCAGCTGCGT CAACGGCACC ATTGACATCT GGAGGACCTC	2940
	CTCGACAACC GGATACAGCT CTGCTAGAGT CATTGGCTGA GCGCTGCATG GAGGTTCTCA	3000
55	TGTCAGAGCA TCATCTGAGA CTGTAACGCG TTATGATCAC AGCACCCGAC ACTTTGCCTC	3060
	GAGTTGTTAA GAACGGACGA CGCGAAATTG GTAACATGCT TTGCCGTGG GAGTTTGATC	3120
	TCGGCAACCT TCCATGTGTG CACGTCAAGT TTGGCGTGGA GCATGCAGTA CTTAACCTCC	3180
60	CTATTGGTGT AGACCCCTATA GGTGGTATCT GGTCAACCGTT GGCGTCCGAT TCTCGTGC	3240
	AATTCTTATT GCCAGCTGAC AAGCAATACT CTGGTGTGCA CAGGCGCGAA GTCGTTATCG	3300
65	ATGACCGTAC TTCAACGCCCTAAACATT TCTCTTGAT TTGGATCTT ATCCAATGGC	3360

- 35 -

	GCGTGGCCCG TCAACCAGAA GAGCTAGCGT ACTGCACAAT CGATGGCAAAG AGCCGAGAAG	3420
	GTAAGGGTGT AACATGGAAG AAATTGACCA CCAAGGTCGC TTCCGTTGCC ATGTACCTGA	3480
5	AGAACAAAGGT CAAGGTGAGC CGGGGAGACC ACATCATCCT CATGTACACA CATTAGAGG	3540
	AGTTTGCTT TGCCATCCAT GCCTGCATT CCTTGGCGC AATGTCAATT CCCATCGCAC	3600
10	CCCTCGACCA GAACCGATTG AACGAAGATG TCCCAGCTT CCTGCATATT GTATCTGATT	3660
	ACAAACGTCAA GGCTGTGCTG GTCAACGCTG AGGTGCGATCA TCTAATCAAG GTAAAGCCTG	3720
	TGGCTAGCCA TATCAAACAG TCAGCCCAGG TTCTCAAGAT CACGAGCCCT GCCATCTACA	3780
15	ACACAACTAA GCCGCCAAAG CAAAGTAGTG GATTGAGGGA TTTGAGATT ACCATTGACC	3840
	CTGCCTGGAT TCGGCCTGGC TACCCCGTCA TTGTTGGAC TTATTGGACC CCCGATCAAC	3900
	GACGAATTTC AGTTCAGCTT GGACATGACA CCATTATGGG CATGTGCAAG GTTCAAAAGG	3960
20	AAACTTGCCA AATGACAAGT TCAAGACCTG TGCTTGGATG TGACGAAGC ACGACTGCC	4020
	TAGGCTTAT TCATACGGCT CTGATGGAA TTTATATCGG AACACCAACC TACCTCCTAT	4080
25	CACCTGTCGA GTTTGCAGCC AACCCCATGT CTCTATTGCGT CACCTGTCG AGATACAAGA	4140
	TTAAGGATAC TTATGCGACA CCACAGATGC TTGATCATGC CATGAACCTCC ATGCAGGCCA	4200
	AGGGCTTTAC ACTTCATGAA CTTAAGAACAA TGATGATCAC TGCCGAGAGC CGACCAAGAG	4260
30	TTGATGTTTT CAAAAGGTC AGACTTCACT TTGCTGGGC TGGGCTCGAT AGAACTGCTA	4320
	TTAACACGGT CTATTGCGAT GTCCTCAACC CCATGGTAGC GTCGGATCT TATATGTGCA	4380
35	TCGAGCCTAT TGAGCTTGG TTGGACACGC AAGCGCTTCG ACGTGGCTG GTTATTCCIG	4440
	TGGACCCCTGA ATCAGATCCT CTGGCCCTAC TGGTACAGGA CAGCGGTATG GTTCCAGTT	4500
	CAACCCAAAT AGCCATCATC AACCCCTGAAA GCAGAACATAA CTGCCTCGAT GGTGAGTATG	4560
40	GTGAAATTG GGTGACTCT GAAGCCTGCG TCAAGTCATT CTATGGCTCC AAAGACGCTT	4620
	TTGACGCTGA GCGCTTGAT GGCGAGCTC TTGACGGCGA TCCCAACATT CAGTATATCC	4680
45	GTACCGGAGA CTTGGGTTTC CTTCTATAATG TTAGTCGACC TATTGGCCCT AATGGTGCC	4740
	AGGTGGACAT GCAAGTGTG TTTGTTCTCG GCAACATTGG CGAGACTTTT GAGATCAACG	4800
	GATTGAGCCA TTTCCAATG GATATTGAGA ACTCGGTGGA AAAATGCCAC AGAAACATTG	4860
50	TGGCGAATGG CTGGTAAGTA TAAAATCTCT ATTTGAAGCG AATATGCTAA CAAAGTCAGT	4920
	CGGGTGTCC AAGCTGGTGG CTTGGTGGTT GTTCTGGTTG AAGTCAACCG CAAGCCATAC	4980
55	CTGGCATCGA TTGTTCCCGT CATTGTCAC GCTATCCTCA ATGAACACCA AATCATTGTA	5040
	GATATCGTCG CATTGTCAC CAAGGGAGAC TTCCACGGT CTCGTCTAGG AGAGAACAG	5100
	CGTGGCAAGA TTCTTGGTGG CTGGTTAGT AGAAAGCTGA GGACTCTTCG CCAGTTCTCG	5160
60	ATTCGCGATA TGGACGCCGA ATCCACAGCT GGTGATATGA TGGATCCTTC TAGAGCATCA	5220
	ATGGTCAGCG TACGAAGCGG AGGCGGTGCT GCTCCGGAT CTTCTAGTTT GAGGAATGTC	5280
65	GAACCTGCGC CTCAAATCTT GGAGGAGGAA CATGACCAGA TGACTCCTCG TCACGAATAC	5340

- 36 -

	GAAGCAGCCC CTACCATGAT TTCTGAACCTT CCCGACGGCC AAGAGACACC GACAGGGTTT	5400
	CAGCACTCGC AATACTAACCA CCCACCACAA TCAGCCGTT CTCAAGCACC AGCCCAGCTG	5460
5	AACCTTCTC ACCAGCCCGA TCAAGGATTG GATATGGACT TTTCACGATA TAGTTCAGCA	5520
	GAGCCCGATC ACGGCCCTGT CCACAGACGT CCAGTCCAGGCCA ACAACCCGAG	5580
10	CCTATGCAAG GGTACGGTCA AGCGCCGCC CAGATCCGGC TACCAGGTGT TGATGGACGA	5640
	GAGGAGGGAG GGTTCTGGTC ACAGCAGGAA AAGAACGAGA AGAGTGAAGA AGACTGGACA	5700
	ACTGATGCCA TGATGCATAT GAATCTGGCA GGTGATATGA AACCGCCACG ATGATAATAC	5760
15	ACAAACATAAG AGCGAACTGA CGAACGGAG TCGGAGTTGG GAAGCATTAA GAAACGAATA	5820
	ACAAACAATT GGACTTGTCG GTCTGATGGC CTATTTACTT CATTCTAGA TGAGGATTGG	5880
20	ATAGTGAATA TGTGATTGGA TAAAGCCTGG GTTTGTGAGT TTGTGAATGC AGTGGGTGCT	5940
	TGCTATAAGC TGTTTATTG AGGTCTTGG AGGAGTGTCT AACAAAGATG CAAAGTTACT	6000
	AGT	6003

25 Preferably, the CPS1 nucleic acid molecule which hybridizes to a nucleic acid molecule having a sequence as set forth in SEQ ID No. 43 is from the plant pathogen *Fusarium graminearum*. In another preferred embodiment, the CPS1 gene is from a plant pathogen such as *Fusarium avenaceum*, *F. carpineum*, *F. chlamydosporum*, *F. coccophilum*, *F. culmorum*, *F. episphaeria*, *F. equiseti*, *F. flocciferum*, *F. moniliforme*, *F. oxysporum*, *F. redolens*, *F. sambucinum*, *F. solani*, *F. subglutinans*, *F. trichothecioides*, *F. udum*, or *F. ventricosum*.

The peptide synthetase product of SEQ ID NO:43 has a deduced amino acid sequence as follows (SEQ ID No. 44):

35	MMSGDYAFRP EQQGTYGESQ HQQHQFQQQQ QPQQQQQQYDG QQYDGRRTTL LDSQGYFSDF	60
	AGQQHYDQTO TVEYVGPOQR YSSSDAFSPT AAMAPPMLTT NDLPPEALE YQLPLDPREV	120
	PFAIQDPHDD STPMSKFDNI AAVLRHRGRT IAKKPAYWVL DSKGKEIASI TWDKLASRAE	180
40	KVAQVIRDKS SLYRGDRVAL IYRDSEVIDF AIALLGCFIA GVVAVPINL QDYQRLNHIL	240
	TTTQAHLALT TDNNLKAFQR DITTQKLTWP KGVEWWKTNE FGSYHPKKKE DVPALVVPDL	300
	AYIEFSRAPT GDLRGVVLSH RTIMHQMACL SAIISTIPGN GPGDTFNPSL RDKNGRLIGG	360
45	GASSEILVSY LDPRQGIGMI LSVLLTVYGG HTTVWFDNKA VDVPGLYAHL LTKYKSTIMI	420
	ADYPGLKRAA YNYQQEPMVT RNFKKGMEPN FQMIKLCID TLTVDSGSHE VLADRWLRLP	480
50	RNPRAREVVA PMLCLPEHGG MVISVRDWLG GEERMGCPNK LELGEDTESD EEEKEETEKPA	540
	VSNFGSLLS GGGTATTEER AKNELGEVLL DREALKTNEV VVVAIGNDAR KRVTDPPGLV	600
	RVGSFGYPIP DATLSVVDPE TGLLASPHSV GEIWVDSPL SGGFWAQPKN TELIFHARPY	660

- 37 -

	KFDPGDPTPQ PVEPEFRLTG LLGTVIEGKI FVLGLYEDRI RQKVEWVEHG HELAEYRYFF	720
5	VQHIVVSIVK NVPKIYDCSA FDVFVNDEHL PVVVLSESAA STAPLTSGGP PRQPDTALLE	780
	SLAERCMEV L MSEHHRLRYC VMITAPDTLP RVVKNGRREI GNMLCRREFD LGNLPCVHVK	840
	FGVEHAVLNL PIGVDPIGGI WSPLASDSRA EFLLPADKQY SGVDRREVVI DDRTSTPLNN	900
10	FSCISDLIQW RVARQPEELA YCTIDGKSRE GKGVTWKKFD TKVASVAMYL KNKVKVRPGD	960
	HIILMYTHSE EFVFAIHACI SLGAIVIPIA PLDNQNRLNED VPAFLHIVSD YNVKAVLVNA	1020
	EVDHLIKVLP VASHIKQSAQ VLKITSPAII NTTKPPKQSS GLRDLRFTID PAWIRPGYPV	1080
15	IWWTYWTPDQ RRISVQLGHD TIMGMCKVQK ETCQMTSSRP VLGCVRSTTG LGFIHTALMG	1140
	IYIGTPYLL SPVEFAANPM SLFVTLSRYK IKDTYATPQM LDHAMNSMQA KGFTLHELKN	1200
20	MMITAESRPR VDVFQKVRLH FAGAGLDRTA INTVYSHVLN PMVASRSYMC IEPIELWLDT	1260
	QALRRGLVIP VDPESDPLAL LVQDSGMVPV STQIAIIINPE SRIHCLDGEY GEIWDSEAC	1320
	VKSFYGSKDA FDAERFDGRA LDGDPNIQYI RTGDLGFLHN VSRPIGPNGA QVDMQVLFVL	1380
25	GNIGETFEIN GLSHFPMDIE NSVEKCHRNI VANGCAVFQA GGLVVVLVEV NRKPYLASIV	1440
	PVIVNAILNE HQIIVDIVAF VNKGDFPRSR LGEKQRGKIL GGWVSRKLRT LAQFSIRDMD	1500
30	AESTAGDMMPSRASMVSVR SGGGAAPGSS SLRNVEPAPQ ILEEEHDQMT PRHEYEAAPT	1560
	MISELPDGQE TPTGFQHSQY EHPPQSAGSQ APAQLNLSHQ PDQGFDMDFS RYSSAEPDHG	1620
35	PVHRRPVPGQ AQQPEPMQGY GQAPPQIRLP GVDGREGGGF WSQQEKNEKS EEDWTTDAMM	1680
	HMNLAGDMKP PR	1692

In another embodiment of the invention, there is provided an isolated nucleic acid molecule encoding a CPS1 peptide synthetase homolog and which hybridizes to a nucleic acid molecule having a sequence corresponding to SEQ ID No. 45 as follows :

	AAAAAGAAGG GGCCTACCGGA GTTGACCGAG ATATTGCTAG ATAAGGAAGC GCTCAAGATG	60
45	AACGATGTTG TGGTCCTTGC AATAGGAGAA GAGGCCAGTA AACGTGCGAA TGAGCCTGGC	120
	ACAATGCGAG TTGGCGCTTT TGGATACCCA ATACCAGATG CGACGCTAGC CGTCGTAGAT	180
	CCAGAGACGA ATCTCTTGTG TTCACCTAC TCGATAGGAG AGATTTGGGT AGACTCACCT	240
50	TCATTGTCTG GTGGTTCTG GCAATTGCAG AAGCACACTG AAACATATATT TCACGCCCGC	300
	CCATACCGCT TTGTGGAGGG CAGTCCTACCC CCGCAGTTGC TTGAGCTTGA GTTCTCCGG	360
	ACAGGCTTAC TCGGATTCTGT CGTAGAGGGC AAGGTCTTTA TCCTTGGTCT CTATGAAGAT	420
55	CGCATCAGGC AGCGCGTTGA ATGGGTAGAA CATGGTCAGC TGGAAGCTGA ACACAGATAAC	480
	TTCTTCGTGC AGCACCTCGT CACCAAGTATC ATGAAGGCTG TTCCCAAGAT CTACGACTGG	540
60	TAAGTCTTCT CATGTTTAG ATGAGCGTTC TAACACTATG CAGCTCATCT TTCGACTCGT	600

- 38 -

	ACGTCAATGG CGAATACTG CCTATCATCC TCATCGAGAC ACAGGCTGCA TCGACAGCCC	660
5	CTACGAACCC TGGTGGACCG CCACAGCAAC TCGACATCCC CTTCTAGAC TCACTGTCTG	720
	AGCGATGCAT GGAAGTGTG TATCAAGAAC ACCATCTGCG AGTATACTGC GTCATGATCA	780
	CAGCGCCAAA CACATTACCA CGAGTTGTA AGAATGGTCG ACGAGAAATT GGCAACATGC	840
10	TCTGTCGAAG AGAATTGAT AATGGCTCAT TACCTTGTGT CCACGTCAAG TTTGGTGTG	900
	AGAGGTCAGT TCTCAACATC GCGTTGGTG ATGACCCCTC CGGAGGCATG TGGTCATATG	960
15	AAGCCTCGAT GGCGCGTCAG CAGTTCTGA TGCTCCAAGA CAAGCAGTAT TCTGGAGTAG	1020
	ATCACCGCGA AGTCGTCATG GATGACAGAA CATCGACACC TCTCAACCAA TTCTCCAACA	1080
	TTCACGACCT CATGCAATGG CGCGTATCAC GGCAGGCTGA AGAGCTCGCA TATTGCACAG	1140
20	TCGACGGTCG AGGCAAAGAA GGCAAGGGCG TCAACTGGAA GAAGTTCGAC CAGAAAGTTG	1200
	CGGGTGTGCG AATGTACCTG AAGAACAAAGG TCAAAGTGCA AACCGGCGAT CATCTGCTTC	1260
	TGATGTATAC GCACTCGAA GACTTTGTAT ATGCGGTACA TGCATGCTT GTGCTTGGCG	1320
25	CTGTATGCAT ACCAATGGCA CCAATCGACC AGAACCGATT GAATGAGGAT GCACCTGCAT	1380
	TGCTGCACAT CCTTGCAGAC TTCAAGGTCA AGGCCATCCT CGTCAATGCC GATGTGGATC	1440
30	ATCTCATGAA GGTCAAGCAA GTATCGCAGC ACATCAAACA ATCAGCAGCC ATCTTCAGA	1500
	TCAACGTGCC GCACACTTAC AACACAACCA AGCCACCTAA GCAGTCGAGT GGTTGTGGGG	1560
35	ATCTCAAGCT CACAATACGG CCTGCCTGGG TACAGCCTGG TTTCCCAGTT CTTGTATGGA	1620
	CATACTGGAC TCCAGATCAA CGCCGTATAG CCGTACAAC AGGTCAATGC CAAATCATGG	1680
	CACTAGGCAA GGTCCAGAAG GAGACTTGTC AAATGACAAG TACAAGGCCA GTCCTAGGAT	1740
40	GTGTACGGAG TACCATCGGA CTTGGCTTCA TTCATACCTG CATCATGGC ATCTTCCTTG	1800
	CCGCACCCAC TTACCTCGTG TCGCCTGTG ACTTTGCACA AAATCCAAAC ATACTCTTCC	1860
45	AGACGTTATC AAGATAACAAG ATCAAGAATG CGTAGCAAC CAGTCAAATG TTGGATCAGG	1920
	CTATTGCCCG TGGGGCTGGA AAGAACATGG CCCTGCACGA ACTCAAGAAT CTCATGATTG	1980
	CGACTGATGG TAGGCCCGT GTTGATGTTT ACCAGAGAGT GCGCGTACAC TTTTCACCAAG	2040
50	CAAGCTTGGGA CGGGACAGCG ATTAACACAG TCTACTCTCA CGTGCTCAAC CCAATGGTAG	2100
	CATCGCGATC ATACATGTGC ATCGAGCCAA TAGAACTGCA TCTCGACGTC AACGCTCTTC	2160
55	GAAGAGGTCT GATCATGCCG GTCGACCCAG ATACCGAGCC TGGCGCTCTA ATGGTCCAGG	2220
	ACTCTGGTAT GGTGCCAGTC TCCACACAAA TAGCAATTGT GAACCCAGAG ACAAAACCAGC	2280
	TTTGCTTGGT TGGCGAATAT GGGCAAATCT GGGTCAATC	2320

60 Preferably, the CPS1 nucleic acid molecule which hybridizes to a nucleic acid molecule having a sequence as set forth in SEQ ID No. 45 is from the plant pathogen *Pyrenophora teres*. In another preferred embodiment, the CPS1

gene is from a plant pathogen such as *Pyrenophora avenae*, *P. bromi*, *P. leuceienes*, *P. phaeocomes*, *P. schroeteri*, *P. trichostoma*, or *P. tritici-repentis*.

The peptide synthetase product of SEQ ID No:45 has a deduced amino acid sequence as follows (SEQ ID No. 46):

5	KKKGPTELTE ILLDKEALKM NDVVVLAIGE EASKRANE PG TMRVGAF GYP IP DATLAVVD	60
	PETNLLCSPY SIGEIWVDSP SLSGGFWQLQ KHTETIFHAR PYRFVEGSPT PQLLELEFLR	120
10	TGLLGFFVVEG KV FILGLYED RIRQRVEWVE HGQLEAEHRY FFVQHLVTSI MKAVPKIYDC	180
	SSFDSYVNNGE YLP II LIETQ AASTAPTNPG GPPQQLDIPF LDSLSERCME VLYQEHHLRV	240
	YCVMITAPNT LPRVVVKNGRR EIGNMLCRRE FDNGSLPCVH VKFGVERSVL NIALGDDPSG	300
15	GMWSYEASMA RQQFLMLQDK QYSGVVDHREV VMDDRTSTPL NQFSNIHDLM QWRVSRQAEE	360
	LAYCTVDGRG KEGKGVNWKK FDQKVAGVAM YLKNKVKVQT GDHLLL MYTH SEDFVYAVHA	420
20	CFVLGAVCIP MAPIDQNRLN EDAPALLHIL ADFKVKA ILV NADVDHLMKV KQVSQHIKQS	480
	AAIFKINVPH TYNTTKPPKQ SSGCRDLKL T IRPAWVQPGF PVLWWTYWT P DQRRIAVQLG	540
	HSQIMALGKV QKETCQMTST RPVLGCVRST IGLGF IHTCI MGIFLAAPTY LVSPV DFAQN	600
25	PNILFQTLSR YKIKNAYATS QMLDHAIARG AGKNMALHEL KNLMIA TDGR PRVDVYQRVR	660
	VHFSPASLDR TAINTVYSHV LNPMVASRSY MCIEPIELHL DVNALRRGLI MPVDPDTEPG	720
	ALMVQDSGMV PVSTQIAIVN PETNQLCLVG EYGEIWVQ	758
30		

As used herein, the term "nucleic acid" refers to deoxyribonucleic acid (DNA) or ribonucleic acid and polymers thereof in either a single or double stranded form. As used herein, "nucleic acid" also encompasses nucleic acids containing known analogs of naturally occurring nucleotides. Unless otherwise indicated, a particular nucleic acid sequence also encompasses conservatively modified variants thereof such as degenerate codon substitutions and complementary sequences. As used herein, the term "nucleotide sequence" refers to a polymer of DNA or RNA which may be single or double stranded and may contain synthetic, non-natural, or altered nucleotide bases capable of incorporation into DNA or RNA polymers. The terms "nucleic acid", "nucleic acid molecule", "nucleic acid fragment", or nucleic acid sequence or segment", may also be used interchangeably with the terms "gene", "cDNA", "DNA" and "RNA".

Other DNA molecules of the present invention include DNA molecules that have a nucleic acid sequence which is more than 70% identical to

- 40 -

the nucleotide sequence of SEQ. ID. Nos. 2, 41, 43, or 45. Nucleotide sequence similarity may be determined by the BLAST program with the default parameters (Altschul et al., "Basic Local Alignment Search Tool," *J. Mol. Biol.*, 215:403-410 (1990), which is hereby incorporated by reference).

5 Preferred sequences include those DNA molecules which will hybridize to a nucleic acid molecule having the sequence of SEQ. ID No. 2, 41, 43, 45 or their compliments. Generally, stringent conditions are selected to be about 50°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. The T_m is dependent upon the solution conditions and the base composition of the probe, and may be calculated using the following equation:

10

$$\begin{aligned} T_m = 79.8^{\circ}\text{C} &+ (18.5 \times \log[\text{Na}^+]) \\ &+ (58.4^{\circ}\text{C} \times \%[\text{G+C}]) \\ 15 &- (820 / \# \text{bp in duplex}) \\ &- (0.5 \times \% \text{ formamide}) \end{aligned}$$

More preferred stringent conditions are when the temperature is 20°C below T_m , and the most preferred stringent conditions are when the temperature is 10°C below T_m . Nonspecific binding may also be controlled using 20 any one of a number of known techniques such as, for example, blocking the membrane with protein-containing solutions, addition of heterologous RNA, DNA, and SDS to the hybridization buffer, and treatment with RNase.

Wash conditions are typically performed at or below stringency. Generally, suitable stringent conditions for nucleic acid hybridization assays or 25 gene amplification detection procedures are asas set forth above. More or less stringent conditions may also be selected.

For the purposes of defining the level of stringency, reference can conveniently be made to Sambrook, J., E.F. Fritsch, et al. 1989 "Molecular Cloning: a Laboratory Manual, 2nd Edition, Cold Spring Harbor, NY, Cold Spring 30 Harbor Laboratory Press, at 11.45. An example of low stringency conditions is

- 41 -

4-6X SSC/0.1-0.5% w/v SDS at 37°-45° C for 2-3 hours. Depending on the source and concentration of the nucleic acid involved in the hybridization, alternative conditions of stringency may be employed such as medium stringent conditions. Examples of medium stringent conditions include 1-4X SSC/0.25%
5 w/v SDS at ≥ 45° C for 2-3 hours. An example of high stringency conditions includes 0.1-1X SSC/0.1% w/v SDS at 60 C for 1-3 hours. The skilled artisan is aware of various parameters which may be altered during hybridization and washing and which will either maintain or change the stringency conditions. For example, another stringent hibridization condition is hybridization at 4X SSC at
10 65° C, followed by a washing in 0.1X SSC at 65° C for about one hour. Alternatively, an exemplary stringent hybridization condition is in 50% formamide, 4XSSC, at 42° C. Still another example of stringent conditions include hybridization at 62° C in 6X SSC, .05X BLOTO, and washing at 2X SSC, 0.1% SDS at 62° C.

15 Other proteins or polypeptides of the present invention include polypeptides that have an amino acid sequence having at least 75 % similarity to the amino acid sequence of at least one of SEQ. ID. No. 3, SEQ ID No. 42, SEQ ID No. 44, or SEQ ID No. 46. In a preferred embodiment of the invention, the protein or polypeptide will have at least 90% similarity with at least one of SEQ.
20 ID No. 3, SEQ ID No. 42, SEQ ID No. 44, or SEQ ID No. 46. Protein sequence similarity may be determined by the BLAST program with the default parameters (Altschul et al., "Basic Local Alignment Search Tool," *J. Mol. Biol.*, 215:403-410 (1990), which is hereby incorporated by reference). The CPS1 protein of SEQ. ID. No. 3 has a molecular weight of about 190-200 kDa, preferably 193.2 kDa.
25 The CPS1 protein contains two structurally similar modules, both of which are similar to SafB1, the first module of saframycin synthetase B (overall 25% identity; 50% similarity) and have apparent amino-acid-activating and thiolation domains with core sequences conserved in known peptide synthetases.

The DNA molecule encoding a CPS1 protein or polypeptide of the
30 present invention can be incorporated in cells using conventional recombinant DNA technology. Generally, this involves inserting the DNA molecule into an expression system to which the DNA molecule is heterologous (i.e., not normally

- 42 -

present). The heterologous DNA molecule is inserted into the expression system or vector in proper sense orientation and correct reading frame. The vector contains the necessary elements for the transcription and translation of the inserted protein-coding sequences. U.S. Patent No. 4,237,224 to Cohen and Boyer, which
5 is hereby incorporated by reference, describes the production of expression systems in the form of recombinant plasmids using restriction enzyme cleavage and ligation with DNA ligase. These recombinant plasmids are then introduced by means of transformation and replicated in unicellular cultures including procaryotic organisms and eukaryotic cells grown in culture. Recombinant genes
10 may also be introduced into viruses, such as vaccinia virus. Recombinant viruses can be generated by transfection of plasmids into cells infected with virus.

Suitable vectors include, but are not limited to, the following viral vectors such as lambda vector system gt11, gt WES.tB, Charon 4, and plasmid vectors such as pBR322, pBR325, pACYC177, pACYC184, pUC8, pUC9,
15 pUC18, pUC19, pLG339, pR290, pKC37, pKC101, SV 40, pBluescript II SK +/- or KS +/- (see "Stratagene Cloning Systems" Catalog (1993) from Stratagene, La Jolla, Calif, which is hereby incorporated by reference), pQE, pIH821, pGEX, pET series (see Studier et. al., "Use of T7 RNA Polymerase to Direct Expression of Cloned Genes," Gene Expression Technology, vol. 185 (1990), which is hereby
20 incorporated by reference), and any derivatives thereof. Suitable vectors are continually being developed and identified. Recombinant molecules can be introduced into cells via transformation, transduction, conjugation, mobilization, or electroporation using methods well known in the art. The DNA sequences are cloned into the vector using standard cloning procedures in the art, as described by
25 Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Springs Laboratory, Cold Springs Harbor, New York (1982), which is hereby incorporated by reference.

A variety of host-vector systems may be utilized to express the protein-encoding sequence(s). Primarily, the vector system must be compatible
30 with the host cell used. Host-vector systems include but are not limited to the following: bacteria transformed with bacteriophage DNA, plasmid DNA, or cosmid DNA; microorganisms such as yeast containing yeast vectors; mammalian

- 43 -

cell systems infected with virus (e.g., vaccinia virus, adenovirus, etc.); insect cell systems infected with virus (e.g., baculovirus); and plant cells infected by bacteria or transformed via particle bombardment (i.e. biolistics). The expression elements of these vectors vary in their strength and specificities. Depending upon the host-vector system utilized, any one of a number of suitable transcription and translation elements can be used.

Different genetic signals and processing events control many levels of gene expression (e.g., DNA transcription and messenger RNA ("mRNA") translation). Transcription of DNA is dependent upon the presence of a promoter which is a DNA sequence that directs the binding of RNA polymerase and thereby promotes mRNA synthesis. The DNA sequences of eukaryotic promoters differ from those of prokaryotic promoters. Furthermore, eukaryotic promoters and accompanying genetic signals may not be recognized in or may not function in a prokaryotic system, and, further, prokaryotic promoters are not recognized and do not function in eukaryotic cells. Similarly, translation of mRNA in prokaryotes depends upon the presence of the proper prokaryotic signals which differ from those of eukaryotes. Efficient translation of mRNA in prokaryotes requires a ribosome binding site called the Shine-Dalgarno ("SD") sequence on the mRNA. This sequence is a short nucleotide sequence of mRNA that is located before the start codon, usually AUG, which encodes the amino-terminal methionine of the protein. The SD sequences are complementary to the 3'-end of the 16S rRNA (ribosomal RNA) and probably promote binding of mRNA to ribosomes by duplexing with the rRNA to allow correct positioning of the ribosome. For a review on maximizing gene expression, see Roberts and Lauer, Methods in Enzymology, 68:473 (1979), which is hereby incorporated by reference.

Promoters vary in their "strength" (i.e., their ability to promote transcription). For the purposes of expressing a cloned gene, it is desirable to use strong promoters in order to obtain a high level of transcription and, hence, expression of the gene. Depending upon the host cell system utilized, any one of a number of suitable promoters may be used. For instance, when cloning in *E. coli*, its bacteriophages, or plasmids, promoters such as the T7 phage promoter, *lac* promoter, *trp* promoter, *recA* promoter, ribosomal RNA promoter, the P_R and

- 44 -

P_L promoters of coliphage lambda and others, including but not limited, to *lacUV5*, *ompF*, *bla*, *lpp*, and the like, may be used to direct high levels of transcription of adjacent DNA segments. Additionally, a hybrid *trp-lacUV5* (*tac*) promoter or other *E. coli* promoters produced by recombinant DNA or other synthetic DNA techniques may be used to provide for transcription of the inserted gene. Bacterial host cell strains and expression vectors may be chosen which inhibit the action of the promoter unless specifically induced. In certain operons, the addition of specific inducers is necessary for efficient transcription of the inserted DNA. For example, the *lac* operon is induced by the addition of lactose or IPTG (isopropylthio-beta-D-galactoside). A variety of other operons, such as *trp*, *pro*, etc., are under different controls.

Specific initiation signals are also required for efficient gene transcription and translation in prokaryotic cells. These transcription and translation initiation signals may vary in "strength" as measured by the quantity of gene specific messenger RNA and protein synthesized, respectively. The DNA expression vector, which contains a promoter, may also contain any combination of various "strong" transcription and/or translation initiation signals. For instance, efficient translation in *E. coli* requires a Shine-Dalgarno ("SD") sequence about 7-9 bases 5' to the initiation codon ("ATG") to provide a ribosome binding site. Thus, any SD-ATG combination that can be utilized by host cell ribosomes may be employed. Such combinations include but are not limited to the SD-ATG combination from the *cro* gene or the *N* gene of coliphage lambda, or from the *E. coli* tryptophan E, D, C, B or A genes. Additionally, any SD-ATG combination produced by recombinant DNA or other techniques involving incorporation of synthetic nucleotides may be used.

The present invention also relates to anti-sense nucleic acid for essential cell proteins, such as replication proteins, which serve to render the host cells incapable of further cell growth and division. Anti-sense regulation has been described by Rosenberg et al., "Production of Phenocopies by Kruppel Antisense RNA Injection Into Drosophila Embryos," *Nature*, 313:703-706 (1985); Preiss et al., "Molecular Genetics of Kruppel, A Gene Required for Segmentation of the Drosophila Embryo," *Nature*, 313:27-32 (1985); Melton, "Injected Anti-sense

- 45 -

RNAs Specifically Block Messenger RNA Translation In vivo," Proc. Natl. Acad. Sci. USA, 82:144-148 (1985); Izant et al., "Constitutive and Conditional Suppression of Exogenous and Endogenous Genes by Anti-sense RNA," Science, 229:345-352 (1985); Kim et al., "Stable Reduction of Thymidine Kinase Activity in Cells Expressing High Levels of Anti-sense RNA," Cell, 42:129-138 (1985); Pestka et al., "Anti-mRNA: Specific Inhibition of Translation of Single mRNA Molecules," Proc. Natl. Acad. Sci. USA, 81:7525-7528 (1984); Coleman et al., "The Use of RNAs Complementary to Specific mRNAs to Regulate the Expression of Individual Bacterial Genes," Cell, 37:429-436 (1984); and McGarry et al., "Inhibition of Heat Shock Protein Synthesis by Heat-Inducible Antisense RNA," Proc. Natl. Acad. Sci. USA, 83:399-403 (1986), which are hereby incorporated by reference.

Once the isolated DNA molecules encoding the CPS1 protein or polypeptide, as described above, has been cloned into an expression system, they 15 are ready to be incorporated into a host cell. Such incorporation can be carried out by the various forms of transformation noted above, depending upon the vector/host cell system. Suitable host cells include, but are not limited to, bacteria, virus, yeast, mammalian cells, insect, plant, and the like. In the present invention, the host cells may be from plants such as corn, oat, grass, weed, 20 bamboo, and sugarcane.

One aspect of the present invention involves using an inhibitor of the CPS1 protein to interfere with the plant infection process in order to impart disease resistance to plants. In one mechanism, the peptide synthetase CPS1 could be inhibited by an appropriate drug, thereby causing the plants to be 25 resistant to fungal attack. Alternatively, the CPS1 nonribosomal product could be degraded by an enzyme for which it is a suitable substrate, and, when the gene encoding this enzyme is genetically engineered into plants, the plants will become resistant to fungal attack.

In this aspect of the present invention, large numbers of 30 compounds can be screened for their activity as inhibitors of CPS1 protein by a high-throughput screening assay as described in U.S. Patent No. 5,876,946 to

- 46 -

Burbaum et al., which is hereby incorporated by reference. Generally, a library of compounds is assayed for inhibition of an enzyme catalyzed reaction and the amounts of fluorescence bound to individual suspendable solid supports measured to determine the degree of inhibition. For example, the amount of fluorescence 5 bound to a microbead in the presence of inhibitory compounds is greater than for non-inhibitory compounds. The amounts of fluorescence bound to individual beads are determined by confocal microscopy. Using this type of assay, inhibition can be determined of a peptide synthetase such as CPS1. For CPS1, the substrate 10 can be amino acids (or hydroxy acids), linked at one end to the microbead and at the other end to a fluorescent label. The enzyme inhibitors can be utilized to impart fungal resistance to a variety of plants including oats, grasses, weeds, sugarcane, and corn in particular.

Thus, the present invention provides a method for identifying 15 inhibitors of a CPS1 protein , wherein said CPS1 protein is a peptide synthetase of a plant pathogen. The method comprises: providing a CPS1 protein or polypeptide, contacting the protein or polypeptide with potential inhibitor compounds; determining peptide synthetase activity, and selecting compounds which decrease the peptide synthetase activity. The method is especially useful in identifying inhibitors of a CPS1 protein from plant pathogens of the genera 20 *Cochliobolus*, *Alternaria*, *Fusarium*, and *Pyrenophora* such as those described hereinabove. Preferably, the method may be used to identify inhibitors of a CPS1 protein from *Alternaria solani*, *Fusarium graminearum*, and *Pyrenophora teres*.

Another aspect of the present invention involves using one or more 25 of the above DNA molecules encoding a CPS1 protein or polypeptide or a gene encoding an enzyme that degrades the CPS1 N.R.P. product to transform plants in order to impart fungal resistance to the plants. This concept of pathogen-derived resistance, according to U.S. Patent No. 5,840,481 to Johnston and Sanford, which is hereby incorporated by reference, is that host resistance to a particular parasite can effectively be engineered by introducing a gene, gene fragment, or modified 30 gene or gene fragment of the pathogen into the host. This approach is based on the fact that in any parasite-host interaction, there are certain parasite-encoded cellular functions (activitics) that are essential to the parasite but not to the host

- 47 -

and that when one of the essential functions of the parasite such as survival or reproduction is disrupted, the parasitic process will be stopped. "Disruption" refers to any change that diminishes the survival, reproduction, or infectivity of the parasite. Such essential functions, which are under the control of the 5 parasite's genes, can be disrupted by the presence of a corresponding gene product in the host which is (1) dysfunctional, (2) in excess, or (3) appears in the wrong context or at the wrong developmental stage in the parasite's life cycle. If such faulty signals are designed specifically for parasitic cell functions, they will have little effect on the host. Therefore, the procedure for making plants, for example, 10 resistant to infection by one or more fungus involves isolating DNA coding for a gene such as CPS1 of a fungus, operably linking the DNA within an expression vector, transforming the plant cell or plant tissue with the expression vector, and growing the transformed plant cells or plant tissue in the presence of the fungus such as e.g., *Cochliobolus heterostrophus*, *Alternaria solani*, *Fusarium* 15 *graminearum*, or *Pyrenophora teres*, where the CPS1 DNA is expressed as a gene product and the CPS protein disrupts the essential activity of the fungi.

Thus, the present invention provides a method of imparting disease resistance to a plant by over-expressing a CPS1 polypeptide in the plant, wherein the polypeptide has protein synthetase activity. In practicing this aspect of the 20 invention, the plant may be any plant in which it is desired to impart disease resistance. Thus, the plant may be an agricultural crop or ornamental plant. The plant may be herbaceous or woody. The plant may be a monocot or dicot. Examples of plants which may be used in practicing the present invention, include 25 but are not limited to, corn, oats, grasses, weeds, sugarcane, barley, wheat, rice, tomato, potato, citrus, malus, rye, cotton, brassica, cabbage, and carrot. Many other plants may also be used in the practice of the present invention. As a guideline, plants which serve as hosts for *Cochliobolus sp.*, *Fusarium sp.*, *Alternaria sp.*, and *Pyrenophera sp.*, may be used. Reference to host plants may be conveniently made to *Fungi on Plants and Plant Products in the United States*, 30 David, F. Farr et al. editors, American Phytopathological Society Press, St. Paul, Minnesota, 1989.

- 48 -

In another preferred embodiment, the CPS1 peptide is from the genera *Cochliobolus*, *Alternaria*, *Fusarium*, or *Pyrenophora*. In an even more preferred embodiment, the CPS1 gene is from *Cochliobolus heterostrophus*, *Alternaria solani*, *Fusarium graminearum* or *Pyrenophora teres*.

5 Promoters and other regulatory regions which function in plants are well known and include e.g., constitutive promoters, inducible promoters, temporally regulated and tissue specific promoters. Examples of constitutive promoters include e.g., actin, CAMV 35S, MAS, ubiquitin, rice cyclophilin, maize H3 histone, and actin 2. Examples of tissue specific promoters include e.g., leaf 10 specific promoters such as the RuBisCo ssu, Cab (chlorophyll a/b/binding) protein, and the AldP gene promoter from rice (Kagaya et al., 1995 *Molecular and General Genetics* 248:668-674. Examples of root specific promoters include e.g., beta tubulin (Oppenheimer et al. *Gene* 63:87, 1988), and SbPRP1 (Suzuki et al., 15 *Plant Mol. Biol.* 21:109-119, 1993. Thus, using well known methods and widely available regulatory sequences, the skilled artisan is direct expression of a subject CPS nucleic acid molecule in a plant.

To provide regulated expression of a CPS gene of the present invention, plants are transformed with a vector which replicates in a plant cell and which have a promoter which directs expression of the CPS gene product in the 20 plant. Methods of plant transformation are well known in the art. A vector comprising a subject nucleic acid molecule coding for a CPS gene or fragment thereof may be introduced into a plant by leaf disk transformation-regeneration procedure as described by Horsh et al. (1985) *Science* 227:1229-1231. Other methods of transformation such as protoplast culture (Horsh et al. 1984 *Science*, 25 223:496; DeBlock et al. (1984) *Embo J.* 2:2143; Barton et al. (1983) *Cell*, 32:1033) may also be used and are within the scope of this invention.

In transforming dicot plant species, plants may be transformed 30 *Agrobacterium*-derived vectors such as those described in Klett et al. (1987) *Annu. Rev. Plant Physiol.*, 38:467. Other well known methods are available to insert the subject CPS genes into plant cells. Such alternative methods include biolistic approaches (Klein et al. 1987, *Nature*:327:70), electroporation,

- 49 -

microinjection (Potrykus and Spangenberg eds., *Gene Transfer to Plants*, Springer Verlag, Berlin , 1995), chemically-induced DNA uptake, the use of viruses or pollen as vectors, liposome mediated transformation, transformation using wounded or enzyme-degraded immature embryos, or wounded or enzyme
5 degraged embryonic callus.

For transformation of monocot plant species, a number of well known methods may be used such as biolistic methods (Tang, K. W. et al. 2000, "Acta Biotechnologica. 20(2): 175-183; de-Villiers, S. M. et al., 2000, South African Journal of Plant and Soil. 17(1): 50- 53) protoplast transformation,
10 electroporation of partially permeabilized cells, and introduction of DNA using glass fibers, etc. *Agrobacterium* may also be used (Raineri, D. M., et al., 1990 Bio/Technology Vol. 8 :33-38).

As used herein, "transformation" refers to the transfer of an exogenous nucleic acid molecule into a host cell. The nucleic acid molecule may
15 be stably or transiently introduced into the host cell and may be maintained non-integrated for example, as a plasmid, or alternatively, may be integrated into the host genome. The resulting transformed plant cell(s) may then be used to regenerate a transformed plant via standard methods.

The invention is further illustrated by the following examples
20 which are not intended in any way to limit the scope of the invention.

EXAMPLES

EXAMPLE 1 -- Materials and Methods for Examples 2-7

Strains, Media, Crosses and Transformation. C4 (*ToxI*⁺; *MAT-*
2) and C5 (*ToxI*⁻; *MAT-I*) are member of near-isogenic *C. heterostrophus* strains
25 (Leach et al., 1982, which is hereby incorporated by reference). R.C4.2696
(*Tox*⁺; *MAT-2*; *hygB*^R) is a C4-derived mutant generated using the REMI mutagenesis procedure (Lu et al., "Tagged Mutations at the *ToxI* Locus of *Cochliobolus heterostrophus* Using Restriction Enzyme-Mediated Integration," Proc. Natl. Acad. Sci. USA, 91:12649-12653 (1994), which is hereby incorporated
30 by reference). Strains 1301R33 (*Tox*⁻; *MAT-2*; *hygB*^R), 1301R45 (*Tox*⁻; *MAT-I*;

- 50 -

hygB^R), 1301R26 (*Tox⁺*; *MAT-2*; *hygB^R*) are progeny of the cross C5 X R.C4.2696. Culture media, including CM (complete medium), CMX (complete medium with xylose instead of glucose), CMNS (CM with salts omitted), and MM (minimal medium) have been described, as have mating procedures (Leach et al., 5 1982; Turgeon et al., "Transformation of the Fungal Maize Pathogen *Cochliobolus heterostrophus* Using the *Aspergillus nidulans* *amdS* Gene," Mol. Gen. Genet., 201:450-453 (1985), which are hereby incorporated by reference). All strains were grown at 24°C under the warm white light or black light (F40/350BL) (Sylvania Inc., Danvers, MA). Ascospore germination was done at 10 32°C in the dark for 3 days. REMI transformants were purified by transferring the transformants from the original REMI plates to fresh CMNS medium containing hygromycin B (Calbiochem^R) at 80 ug/ml. For conidiation, stable transformants were transferred to CMX containing the same drug but at a higher concentration (120 ug/ml) to compensate for reduced drug activity due to the inhibition by the 15 salts in the medium. Single conidia were picked up under a dissecting microscope and grown on CMNS hygromycin B plates; stable colonies were then transferred to individual CMX/ hygromycin B plates. All purified transformants were stored at -70°C in CM liquid medium containing 25% of glycerol in 96-well microtiter dishes.

20 **Bioassays.** Fungal strains were grown on CMX plates (100 X 15mm) for 7-10 days at 24°C under the light for maximum conidiation. To verify normal T-toxin production by a race T isolate, 1.0 ml of T-toxin-sensitive *E. coli* (DH5a) cells were evenly spreaded on LB medium containing ampicillin (100 ug/ml) and the plates were allowed to air dry for 30 min in a laminar hood. Agar 25 plugs bearing fungal mycelia were inoculated (upside down) onto the *E. coli* cell lawn and the plates were incubated at 32°C. Wild type race T and race O were used as controls for each assay plate. T-toxin-producing strains of the fungus will inhibit growth of the *E. coli* cells and produce halos. *Tox-* mutants can be distinguished from wild type by failure to produce a halo (tight) or by production 30 of halos smaller (leaky) or larger than wild type (overproducing). All *Tox-* mutants were transferred to Fries medium (Pringle et al., "The Isolation of the Toxin of *Helminthosporium victoriae*," Phytopathology, 47:369-371 (1957),

- 51 -

which is hereby incorporated by reference), which optimizes toxin production, and retested. T-cytoplasm corn plants (inbred W64A) are used to verify the *Tox*⁻ mutants identified from the *E. coli* assay using the procedure described below. Mutants defective in T-toxin production fail to produce typical race T symptoms 5 on T-corn. Pathogenicity phenotype on N-cytoplasm corn and virulence of *Tox*⁺ strains to T-cytoplasm corn were determined by a plant assay where about 3,000 transformants generated using the REMI mutagenesis procedure (Lu et al., "Tagged Mutations at the *Tox1* Locus of *Cochliobolus heterostrophus* Using Restriction Enzyme-Mediated Integration," Proc. Natl. Acad. Sci. USA, 10 91:12649-12653 (1994), which is hereby incorporated by reference) were screened for mutants defective in ability to cause disease on corn plants. Two week old N-cytoplasm corn plants (inbred W64A) grown in the green house (5-6 15 plants in one 4" X 6" pot) were inoculated with 5 ml conidial suspensions (10^5 conidia/ml) using a pressurized Preval Spray Gun Power Unit thin layer chromatography sprayer (Alltech Associates, Deerfield, IL), incubated in the mist chamber for 24 hours (23°C) and then taken to the growth chamber (23°C, 80% humidity, 14 hours of light). The mutant phenotypes were determined by occurrence of apparent variations in disease symptom development, mainly by 20 lesion size comparison. Mutants producing lesions smaller than wild type were retested and lengths of typical lesions from each mutant were compared with wild type 7 days after inoculation and measurements were taken for statistical evaluation.

DNA manipulations and sequencing. Genomic and plasmid DNA preparation, restriction enzyme digestions, gel electrophoresis and gel blot 25 analysis were done using standard protocols (Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor, New York:Cold Spring Harbor Laboratory Press (1989), which is hereby incorporated by reference). DNA was sequenced at the Cornell DNA Sequencing Facility using TaqCycle automated sequencing with DyeDeoxy terminators (Applied 30 Biosystems, Foster City, CA). pUCATPH was used for subcloning (Table 1). Primers used for sequencing (Table 2) were designed using Primer Select (DNASTAR Inc., LaserGene system) and synthesized by the Cornell

- 52 -

Oligonucleotide Synthesis Facility. Sequencing of each plasmid clone was initiated with vector-specific primers or primers designed to previously determined sequences. Sequences obtained were analyzed using the same system and nucleotide or protein database searches were performed with the BLAST 5 program (Altschul et al., "Basic Local Alignment Search Tool," J. Mol. Biol., 215:403-419 (1990), which is hereby incorporated by reference).

Table 1. Transformation vectors and clones used

Plasmid	Length (kb) ^a	Characteristics	Reference
pUCATPH	<u>5.1</u>	See Figure 29.	
pUCATPH N	<u>4.6</u>	Cloning vector, same as pUCATPH but lacking a 420 bp <i>NarI</i> fragment containing the <i>HindIII</i> site	Figure 8
p214B7	<u>9.2</u>	A clone containing pUCATPH recovered from the tagged site in mutant R.C4.2696 by religation of <i>BglII</i> -digested genomic DNA	Figure 2
p214M1	<u>6.3</u>	As above but with <i>MscI</i> -digested genomic DNA	Figure 2
p214S1	<u>9.3</u>	As above but with <i>SacI</i> -digested genomic DNA	Figure 2
p214S1N	<u>3.3</u>	<i>NarI</i> fragment derived from 214S1 containing a 0.8 kb <i>NarI-SacI</i> fragment of genomic DNA ligated to pUC18	Figure 4
p214SNP	<u>8.4</u>	Vector for targeted integration constructed by ligating <i>HindIII</i> -digested pUCATPH into the <i>HindIII</i> site of p214S1N	Figure 4
p118BSP	<u>7.3</u>	Vector for targeted integration constructed by ligation of a 2.2 kb <i>SacI</i> fragment of p118B14 into the <i>SacI</i> site of pUCATPH	Figure 6
p118BCS	<u>5.4</u>	Vector for targeted integration constructed by ligation of a 0.8 kb <i>SspI</i> fragment of p118BC4 into the <i>SspI</i> site of pUCATPHN	Figure 9
p118B14	<u>10.4</u>	A clone recovered from the p214SNP integration site in transformant #118 by ligation of a <i>BglII</i> -digested genomic DNA fragment containing the entire vector	Figure 5
p118BC4	<u>6.7</u>	A clone recovered from same site as above but by ligation of a <i>BclI</i> -digested genomic DNA fragment containing part of vector (214SNP) sequence	Figure 5
p9P2	<u>7.3</u>	A clone recovered from the p118BSP integration site in transformant #9 by ligation of a <i>PstI</i> -digested genomic DNA fragment containing pUC18	Figure 7
p12H6	<u>8.0</u>	A clone recovered from the p118BCS integration site in transformant #12 by ligation of a <i>HindIII</i> -digested genomic DNA fragment containing the entire vector.	Figure 9

5 a. An underlined kb number indicates that the plasmid carries genomic DNA sequences (see related figures for details).

Table 2. Primers used for sequencing recovered genomic DNA flanking the REMI insertion site at the R.C4 2696 mutation.

Name ^a	Position ^b	Sequence ^c	Plasmid ^d	Origin ^e
M13RMT		GCGGATAACAATTCACACAGGA SEQ. ID. No. 4	A	pUC18
1. RP1b	775	AGGCCAGCTGCTCTCTTG SEQ. ID. No. 5	A	214B7TrpC
2. RP2	604	ACTCGGACGGACCGAACAA SEQ. ID. No. 6	A	214B7RP1b
3. RP3	119	CGGAAGGAGTGCACCAA SEQ. ID. No. 7	A	214B7RP2
4. RP4	-232	GCTGCTTGCATCTGGTCTTG SEQ. ID. No. 8	A	214B7RP3
5. RP5	-812	AGACCCAGCTGTTGCCATTG SEQ. ID. No. 9	A	214B7RP4
6. RP5b	-1215	CGGAGACGAAAGCCTGAGA SEQ. ID. No. 10	A	214B7RP4
7. RP6	-1392	TGCCAGCTGCGTCCAAGAAG SEQ. ID. No. 11	A	214B7RP5
8. RP7	-1839	GCTAGCATGGCCCTCACAC SEQ. ID. No. 12	A	214B7RP6
TrpC		TGTGTTGACCTCCACTAGCTC SEQ. ID. No. 13	A	PUCATPH
9. FP1	1885	CTACGGGATGCAGAGGGAAAGT SEQ. ID. No. 14	A	214B7TrpC
10. FP1b	1828	GCCATGATTAGCACGATAACCC SEQ. ID. No. 15	B	214B7TrpC
11. FP2	2028	CGCGCTGCATACAACCTACCAA SEQ. ID. No. 16	B	214M1FP1b
12. FP3	2490	TGGTGGCACTACAAAGAACAA SEQ. ID. No. 17	C	214M1FP2
13. FP4	2949	CAGCGTGTGAATGGTAGAA SEQ. ID. No. 18	C	214S1FP3
14. FP4B	2745	CTGGGTAGATTGCCCTTCAC SEQ. ID. No. 19	C	214S1FP4
15. FP5	3421	GAGCGATCAGTCAGAACATT SEQ. ID. No. 20	C	214S1FP4
16. FP6	3948	CGCTGACGTTGACCATCTGA SEQ. ID. No. 21	C	214S1FP5
17. FP7	4411	GCATATGCAACGAGTCAAA SEQ. ID. No. 22	C, D	214S1FP6
18. FP8	5035	ACGGTGACCTGTTGATA SEQ. ID. No. 23	D	118B14FP7
19. FP9	5457	ATGCGCACAAATAGCCCAGTA SEQ. ID. No. 24		118BC4FP8
20. RP48	2865	TTCAAGCAACTGTGGCGTAGG SEQ. ID. No. 25	D	214S1FP6
21. FP10	5790	GATCCTAGCGACCGCACACCAAC SEQ. ID. No. 26	F	9P2FP9
22. FP11	6327	CCTGCTGCTGGTGCTTCT SEQ. ID. No. 27	F	9P2FP10

- 55 -

23. FP11b	6211	GAGTTGCAAATCGTGACAGC SEQ. ID. No. 28	F	9P2FP10
24. FP12	6457	TATCAGCTGTTCAATGTTCTA SEQ. ID. No. 29	F	9P2FP11
25. FP13	6854	TGTTATCCCATTGCCATTG SEQ. ID. No. 30	F	9P2FP12
26. FP14	7400	AAGGACGGAGATTGGTGGAG SEQ. ID. No. 31	F	9P2FP13
27. FP15	7771	GGAGATGGCGGTGACGA SEQ. ID. No. 32	F	9P2FP14
28. FP16	8145	GCATGGCTTGAGGAC SEQ. ID. No. 33	F	9P2FP15
29. FP17	8492	AGATTGGCTAGTATGGAGGTAA SEQ. ID. No. 34	F	9P2FP16
M13F40		GTTTTCCCAAGTCACGAC SEQ. ID. No. 35	G	pUC18
30. RP1	8953	TACTACTAGCATACCAGCATACCT SEQ. ID. No. 36	G	9P5M13F4
31. RP2	8559	TCAACCTCGGAATACCAAGTC SEQ. ID. No. 37	G	9P5RP1

- a. "RP" indicates reverse primer; "FP" indicates forward primer. Primers designed to genomic DNA sequences are numbered in order. For stock tube and the notebook, Primers 1-17 have a leading number "214"; 18-20 with "118"; 21-29 with "9P2" and 30-31 with "9P5". M13RMT(a M13R mutant version; there is a mutation in the polylinker of pUC18) and M13F-40 are provided by Cornell DNA Sequenceing Facility. TrpC primer site is in the pUCATPH *TrpC* promoter region 38 bp from *SalI* site with sequencing direction from *SalI* to *KpnI*.
- b. The position of the first base of each primer corresponds to the assembled sequence (*CPS1* + *TES1*, total 11.3 kb found in Lu's folder in the computer Yoder lab1)
- c. Each primer sequence is given in the 5' to 3' direction
- d. Plasmids used as templates for each sequencing reaction. A = p214B7; B = p214M1; C = p214S1; D = p118B14; E = p118BC4; F = p9P2. G = p9P5 (=9P2)
- e. Original sequences that were used for primer design can be found in the CPS1 sequence notebook or in Shunwen Lu's folder (CPS1 sequence) in the computer Yoder lab1 under the same names as listed.
- Recovery of tagged DNA from the REM1 insertion site and targeted gene disruption.** Genomic DNA of mutant R.C4.2696 was digested with *Bgl*II, *Msc*I (no sites in pUCATPH) or *Sac*I (which cuts the vector once) and purified by phenol extraction and ethanol precipitation, then dissolved in TE (pH 8.0). Ligation was performed in 50 ul reaction mixture, containing 1 x T4 DNA ligase buffer with 10 mM ATP, 60 units T4 DNA ligase (New England Biolabs, Beverly, MA) and 3 ug of *Bgl*II-digested genomic DNA, at 14°C overnight. 10 ul of ligation mixture was used to transform 200 ul of competent DH5a cells,

- 56 -

prepared using the calcium chloride treatment (Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor, New York: Cold Spring Harbor Laboratory Press (1989), which is hereby incorporated by reference), to ampicillin resistance. Ampicillin resistant clones were analyzed by
5 digestion of plasmid DNA with several diagnostic restriction enzymes and clones containing the REMI vector plus flanking genomic DNA were sequenced using the vector-specific primers (M13R or *TrpC*). Three plasmids, p214B7, p214M1 and p214S1 (Figure 2) were recovered and used for sequencing. For targeted gene disruption in wild type, p214B7 was amplified and plasmid DNA purified by
10 equilibrium centrifugation in CsCl-ethidium bromide gradients (Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor, New York: Cold Spring Harbor Laboratory Press (1989), which is hereby incorporated by reference). 30 ug of plasmid DNA (linearized with *Bgl*II for double crossover integration as illustrated in Figure 3) were used to transform wild type and the
15 transformants were purified by isolation of single conidia, assayed for pathogenicity and characterized by gel blot analysis.

Sequence extension by targeted integration and plasmid rescue.

Two overlapping cosmid clones were isolated by probing a genomic DNA library of C4 constructed on a cosmid vector, but both extended into the left region only
20 of p214B7. To extend to the right, a chromosome walking strategy was employed (Figure 3). Three targeted gene disruption experiments (each followed by plasmid rescue) were done successively. In the first experiment, a vector was constructed as follows (see Figure 4 for details): p214S1 was digested with *Nar*I and religated to create p214S1N, which was then digested with *Hind*III and ligated into the
25 *Hind*III site of pUCATPH to create p214SNP for transformation of race O (C5). One transformant (Tx118) resulting from homologous integration (confirmed by gel blot analysis) was used for plasmid rescue as described above. Two new plasmids p118B14 and p118BC4 were recovered, both of which carry sequence at the 3' end but only 172 and 680 bp more than p214S1, respectively (Figure 5). To
30 continue the walk, p118B14 was digested with *Sac*I and ligated into the *Sac*I site of pUCATPH to create p118BSP (Figure 6). This vector was transformed into wild type and one plasmid, p9P2 was recovered (from transformant Tx9), which extends 4.4 kb into the region 3' of p118BC4 and contains the 3' end of *CPS1*.

(Figure 7). A third experiment was done in an attempt to recover a 15 kb *XhoI* fragment at the 3' end of that tagged gene. p118BCS (Figure 8) was constructed by subcloning a 0.8 kb *SspI* fragment into the same site of pUCATPHN. Plasmid rescue using *XhoI* digested-genomic DNA of a transformant (TX12) failed to 5 recover the 15 kb *XhoI* fragment, but p12H6 was recovered using *HindIII*-digested genomic DNA of the same transformant; the genomic DNA matched that already cloned on p9P2 (Figure 9).

EXAMPLE 2 -- Characterization of the REM1 mutant.

- 10 1) In all culture conditions used in the lab, mutant R.C4.2696 grows just like wild type with no variations in growth rate, color and morphological features (Figure 10A). It produces normal conidia that germinate and form infection structures like wild type when induced on artificial surfaces (Figure 10B) and shows normal mating ability when crossed to wild type testers.
- 15 15) No pleiotropic phenotypes associated with the mutation have been detected so far.
- 20 2) The mutant differs from wild type in the ability to cause disease on corn plants. When tested on T-cytoplasm corn, the mutant produces race T type symptoms but the disease develops more slowly than with wild type although it produces wild type levels of T-toxin as detected in a microbial assay (Figure 11), suggesting that the reduced virulence is not related to a deficiency in the ability to produce T-toxin. This is clearer on N-cytoplasm corn where the mutant produces lesions significantly smaller than those produced by wild type (Figure 12). When the mutant was crossed to a wild type race O tester, the small lesion phenotype and ability to produce T-toxin segregated independently, indicating that 25 mutant phenotype is not associated with the reduced fitness trait tightly linked with the *Tox1* locus (Klittich et al., "Reduced Fitness Associated With *Tox1* of *Cochliobolus heterostrophus*," *Phytopathology*, 76:1294-1298 (1986), which is hereby incorporated by reference). The statistical evaluation of lesion size in the race O genetic background indicates that the mutation causes 60% reduction in the 30 fungal virulence to corn plants (Figure 13).
- 30 3) The mutant phenotype is caused by a tagged, single site mutation. In crosses between the mutant and wild type testers, progeny segregated 1 : 1 for parental types only and all hygromycin B-resistant progeny

produced lesions similar to the mutant parent; all hygromycin B-sensitive progeny produced wild type lesions (Figure 14), indicating that a tagged mutation is responsible for the reduced pathogenicity of the mutant.

5 **EXAMPLE 3 -- Cloning and sequencing of DNA flanking the REMI vector insertion site.**

A total of 11.3 kb of genomic DNA surrounding the insertion site was cloned and completely sequenced (Figure 15). The sequence was derived from seven plasmid clones. The first three (p214B7, p214M1 and p214S1) were 10 recovered from the tagged site in mutant R.C4.2696 and cover about 60 % (6.6 kb) of the entire region. The rest (p118B14, p118BC4, p9P2 and p12H6) were recovered from transformants generated using the chromosome walking strategy. DNA to the left of the insertion site (3.4 kb) was cloned on p214B7; DNA on the right (7.9 kb) was cloned on different overlapping plasmids. p9P2 carries the 15 largest amount (4.6 kb) including genomic DNA on p12H6 (Figure 15).

EXAMPLE 4 -- Identification of CPS1 and TES1 at the sequenced region.

Analysis of the combined sequences revealed two open reading frames (ORFs). ORF1(5.4 kb) starts 576 bp upstream of the REMI vector 20 insertion site and ends with an in-frame stop codon (TAG) 3029 bp from the end of the sequenced region in the right flank (Figure 15). No "TATA" box-like element is found in the expected position, but five putative "CAAT" boxes are located upstream of the start codon (ATG), three of them are in the range found in most filamentous fungal promoters (60-200 bp) (Gurr et al., 1987, which is hereby 25 incorporated by reference). Sequence around ATG of ORF1 (CACCATGCT (SEQ. ID. No. 38)) is similar to the fungal consensus (CACCATGGC (SEQ. ID. No. 39)). Although there are several ATGs found upstream, they are less likely to be used as a start codon because the surrounding sequences lack similarity to the consensus. Three putative introns are identified by their conserved 5' and 3' 30 border sequences and potential branch sites (Table 3). Splicing these introns eliminated stop codons which would otherwise interrupt the 5.4 kb open reading frame. Three introns have similar size (45-53 bp respectively) which is in the range of intron size determined from most fungal genes. A putative

polyadenylation signal (ATAA) is found 223 bp downstream of the translation termination site. The G+C content of ORF1 is 51.5%, which is similar to most *Cochliobolus* genes (Turgeon et al., "Cloning and Analysis of the Mating Type Genes from *Cochliobolus heterostrophus*," *Mol. Gen. Gene.*, 238(1-2):270-284 (1993); VanWert et al., "Structure of the *Cochliobolus-heterostrophus* Glyceraldehyde-3-Phosphate Dehydrogenase Gene," *Curr. Genet.*, 22(1):29-35 (1992); Yang et al., "A Polyketide Synthase is Required for Fungal Virulence and Production of the Polyketide T-Toxin," *Plant Cell*, 8(11):2139-2150 (1996); Rose et al., "A Decarboxylase Required for Polyketide Toxin Production and High Virulence by *Cochliobolus heterostrophus*," *8th Int. Symp. Mol. Plant-Microbe Int.*, Knoxville, p. J-49 (1996), which are hereby incorporated by reference). Interestingly, ORF1 is flanked by two regions of G+C rich DNA. The first (1.4 kb, 60.7% G+C) is found between ORF1 and ORF2; the second (1.2 kb, 60.3% G+C) is found 1.8 kb downstream of the stop codon of ORF1 (Figure 16).

15 Database searches using the translated protein sequence of ORF1 revealed high similarity to SafB, one of the multifunctional enzymes catalyzing the biosynthesis of the cyclic peptide antibiotic saframycin Mx1 produced by the bacterium *Myxococcus xanthus* (Pospiech et al., "Two Multifunctional Peptide Synthetases and an O-methyltransferase are Involved in the Biosynthesis of the DNA-Binding

20 Antibiotic and Antitumour Agent Saframycin Mx1 from *Myxococcus xanthus*," *Microbiology*, 142(4):741-746 (1996), which is hereby incorporated by reference). The entire nucleotide sequence of ORF1 (designated *CPSI*) is given in Figure 17.

Table 3. Characteristics of putative introns in *CPSI* and *TESI*

25

Gene	Intron	Size(bp)	Location	5'Border	3'Border	Branch Site
<i>CPSI</i>	I	45	3060-3105	GTAAGT	TAG	GTCTAAC
	II	51	4532-4582	GTAAGT	CAG	TGCTAAC
	III	53	5187-5239	GTACGT	CAG	TACTAAC
<i>TESI</i>	I	49	528-566	GTAAGT	TAG	CCTTAAG
Consensus				GTA ^A /CGT	T/CAG	YNCTAAC*

- 60 -

* Y = Pyrimidine (T or C); N = purine or pyrimidine.

ORF2 starts about 1.6 kb upstream of the start codon of *CPS1* and
5 is transcribed in the opposite direction (Figure 15). No "TATA" box-like element
and CAAT box are found; instead, an AT-rich sequence "AAACTAT" (SEQ.
ID. No. 40) is located 11 bp upstream of the start codon ATG and a CT motif is
found in the -30 region, which is characteristic of a number of fungal genes that
lack a CAAT box in their promoter region (Gurr et al., "The Structure and
10 Organization of Nuclear Genes of Filamentous Fungi," in Kinghorn, ed., Gene
Structure in Eukaryotic Microbes, Vol. 22, published by the Society for General
Microbiology, Oxford, England:IRL Press, pp. 93-140 (1987), which is hereby
incorporated by reference). The sequence around ATG matches perfectly fungal
gene consensus. A putative intron (50 bp) is found in the middle of ORF2 with
15 conserved 5' and 3' border sequences and a potential branch site (Table 3). A
putative polyadenylation signal (AAATA) is found 189 bp downstream of the
translation stop codon TGA. The G+C content of ORF2 is 55.5%, which is
slightly higher than the normal range because the 5' end of ORF2 is located in the
region of G+C rich DNA upstream of ORF1 (Figure 16). Database search
20 revealed that ORF2 encodes a protein with high similarity to *Homo sapiens*
thioesterase II (hTE, Liu et al., "Binding of HIV-1 Nef to a Novel Thioesterase
Enzyme Correlates with Nef-Mediated CD4 Down-Regulation," J. Biol. Chem.,
272(21):13779-13785 (1997), which is hereby incorporated by reference) and *E.*
coli thioesterase II encoded by the *tesB* gene (Naggert et al., "Cloning,
25 Sequencing and Characterization of *Escherichia-coli* Thioesterase II," J. Biol.
Chem., 266(17):11044-11050 (1991), which is hereby incorporated by reference).
The nucleotide sequence of ORF2 (designated *TES1*) is given in Figure 18.

EXAMPLE 5 -- Modular structure of CPS1.

30 Predicted CPS1 protein (1743 amino acids, *M_r* 193235) contains
two structurally similar modules, both of which are similar to SafB1, the first
module of saframycin synthetase B (overall 25% identity; 50% similarity) and

have apparent amino-acid-activating and thiolation domains but lack methyltransferase activity, thus appearing to be typical type I modules (Figure 19). The number of amino acids in each module is different: the first module (CPS1A) consists of 574 amino acids (from the first residue of core 1 to the last 5 residue of core 6), which is larger than most type I modules; the second module (CPS1B) has 530 amino acids, which is average. The distance between the two modules is 193 amino acids, much shorter than most peptide synthetases (500-600 aa), but this distance is not highly conserved, i.e., an opposite variation is found in HC-toxin synthetase and cyclosporine synthetase, both of which have about 1,000 10 aa between the first and second amino-acid-activating module (Figure 20F).

Amino acid alignment of the two modules of CPS1 to SafB1 indicated that these modules are highly similar to each other in both overall amino acid composition and conserved motif sequences as defined by Stachelhaus and Marahiel (Stachelhaus et al., "Modular Structure of Peptide Synthetases Revealed 15 by Dissection of the Multifunctional Enzyme GrsA," *J. Biol. Chem.*, 270(11):6163-6169 (1995); Marahiel, "Protein Templates for the Biosynthesis of Peptide Antibiotics," *Chem. Biol.*, 4(8):561-567 (1997), which are hereby incorporated by reference). When aligned to other bacterial or fungal peptide synthetases, CPS1 only showed local similarity to cyclosporine synthetase (Weber et al., "The Peptide Synthetase Catalyzing Cyclosporine Production in 20 *Tolyphocladium niveum* is Encoded by a Giant 45.8-Kilobase Open Reading Frame," *Current Genetics*, 26(2):120-125 (1994), which is hereby incorporated by reference) and tyrocidine synthetase A (Mootz et al., "The Tyrocidine Biosynthesis Operon of *Acillus brevis*: Complete Nucleotide Sequence and 25 Biochemical Characterization of Functional Internal Adenylation Domains," *J. Bacteriol.*, 179(21):6843-6850 (1997), which is hereby incorporated by reference), but when the amino acids in motif regions were aligned, a overall conservation was observed. Both CPS1A and CPS1B have all five core sequences in the amino-acid-activating domain (Figure 20A-E). Cores 3 and 4 are well conserved 30 except for the replacement of an aspartic acid residue of core 4 by a leucine in CPS1A. Cores 1, 2 and 5 show weak conservation, but similar variations are also seen in SafB1. A thiolation domain is found in both modules, which contains a highly conserved motif (core 6, Figure 20F). The serine residue in this motif has

- 62 -

been shown to be the active site for 4'-phosphopantetheine attachment (Schlumbohm et al, "An Active Serine is Involved in Covalent Substrate Amino Acid Binding at Each Reaction Center of Gramicidin S Synthetase," *J. Biol. Chem.*, 266(34):23135-23141 (1991); Stein et al., "Detection of 4'-
5 Phosphopantetheine at the Thioester Binding Site for L-Valine of GramicidinS
Synthetase 2," *FEBS Lett.*, 340(1-2):39-44 (1994), which are hereby incorporated
by reference). The distances between the six core sequences in the two modules
are also largely conserved. Two exceptions are found in the first module, which
has 312 aa between cores 2 and 3, larger than normal (150-200); 61 between cores
10 5 and 6, only half of that of most peptide synthetases. SafB1 also shows distance
variations at these two interval regions (Figure 20B and E). In addition to amino-
acid-activating and thiolation domains, CPS1 also has an integrated thioesterase
domain (TE) in the carboxy-terminal end of CPS1B (Figure 19). A signature
sequence GXSXG, which is highly conserved in animal fatty acid thioesterase
15 type II enzymes and several peptide synthetases, is found in this domain (Figure
21).

EXAMPLE 6 -- Sequence homology analysis of TES1 protein.

The predicted TES1 protein consists of 367 amino acids (M_r
20 41013). Amino acid alignment of TES1 to hTE, TESB and *Mycobacterium*
tuberculosis TESB homolog (Philipp et al., "An Integrated Map of the Genome of
the Tuberle bacillus, *Mycobacterium tuberculosis* H37Rv, and Comparison with
Mycobacterium leprae," *Proc. Natl. Acad. Sci. USA*, 93(7):3132-3137 (1996),
which is hereby incorporated by reference) showed that these proteins have an
25 overall 40% identity and 60% similarity. A highly conserved VHS motif (putative
active site) is found in the C-terminal region of TES1 at a conserved position
(Figure 22). All these thioesterases have no sequence similarity with the
previously identified animal type I or type II thioesterases known to be involved
in the chain termination of fatty acid synthesis (Naggert et al., "Cloning,
30 Sequencing and Characterization of *Escherichia-coli* Thioesterase II," *J. Biol.
Chem.*, 266(17):11044-11050 (1991), which is hereby incorporated by reference).
Interestingly, TES1 has more homology to hTE than to two bacterial genes,
suggesting that both proteins belong to a new family of eucaryotic thioesterases.

EXAMPLE 7 -- Targeted disruption of CPS1.

Disruption of either CPS1A or CPS1B restored the original mutant phenotype. Ten transformants from each of four individual disruption experiments using different constructs, including the plasmid recovered from the REMI insertion site in the mutant (p214B7) and three vectors for chromosome walking (p214SNP, p118BSP and p118BCS) were purified and assayed on N-cytoplasm corn. All transformants showed the same small lesion phenotype as that of the original REMI mutant. Gel blot analysis confirmed that all 10 transformants showing the mutant phenotype resulted from homologous integration of the transforming vector that disrupted the wild type *CPS1* (Figures 24-26). No transformants showing the wild type phenotype were obtained, presumably because of the large genomic DNA fragments (over 800 bp in all disruption experiments) on the transforming vector that resulted from high 15 efficiency of homologous recombination and the low chance to recover transformants with ectopic integration.

EXAMPLE 8 -- Methods and Materials for Examples 9-10

Strains, growth conditions and transformation. Strains of 20 *Cochliobolus* species and relatives used for genomic DNA hybridization are listed in Table 4. The strain HvW, a victorin-producing isolate of *C. victoriae* was recovered from storage and grown on CMX medium (Turgeon et al., “Transformation of the Fungal Maize Pathogen *Cochliobolus heterostrophus* Using the *Aspergillus nidulans amdS* Gene,” *Mol. Gen. Genet.*, 201:450-453 25 (1985), which is hereby incorporated by reference) for conidiation or on oat meal agar medium (Churchill et al., “Victorin-Deficient REMI Mutants of *Cochliobolus victoriae* Demonstrate a Requirement for Victorin in Pathogenesis,” *Fungal Genet. News*, 42A:41 (1995), which is hereby incorporated by reference) for victorin detection at 24°C under warm white lights (Sylvania Inc., Danvers, MA). 30 Transformation was done using the *C. heterostrophus* procedure (Turgeon et al., “Cloning and Analysis of the Mating Type Genes from *Cochliobolus heterostrophus*,” *Mol. Gen. Gene.*, 238(1-2):270-284 (1993), which is hereby incorporated by reference).

- 64 -

Table 4. Detection of *CPSI* homologs in *Cochliobolus* spp and relatives

Strain ^a	Host ^b	<i>Eco</i> RI digest ^c	Hybridization		<i>Bgl</i> II digest ^e
			<i>Hind</i> III digest ^d		
<i>C. heterostrophus</i> race T (C4)	Corn (<i>Turf-13</i>)	+	5.2	3.2	4.2
race O (C5)		+	5.2	3.2	4.2
<i>C. carbonum</i> race 1 (26R13)	Corn ¹ (<i>hm1hm1</i>)	+	6.6		5.0
race 2 (YugY)		N	6.6		5.0
race 3 (BZ1703)*		N	6.6		5.0
<i>C. victoriae</i> (<i>HvW</i>)	Oats (<i>Vb</i>)	+	N		5.0
<i>C. sativus</i> (A20)	Grasses ²	+	3.0		N
<i>C. specifer</i> (D5-7)	Grasses ²	+	N		N
<i>C. homomorphus</i> (ATCC 13409)	Unknown	N	5.8		N
<i>C. dactyloctenii</i> (7938-9)	Unknown	N	5.9		N
<i>S. turcica</i> (NK2)	Sorghum and maize ³	+	N		N
<i>S. rostrata</i> (32197)	Weeds and bamboo ⁴	+	2.8		N
<i>B. sacchari</i> (764-1) (1249-10)	Sugarcane ⁵		5.4	2.5	N
		+	5.4	2.5	N
		N			

- a. *C.* = *Cochliobolus*. *S.* = *Setosphaeria*. *B.* = *Bioplaris*. The name of isolates (or lab strains) of each species are given in parentheses and those known to produce host-specific toxins are underlined. * Provided by Tsukiboshi Takao (Japan) and the isolate could be either BZ1209 or BZ1703.
- b. Genotype susceptible to the host-specific toxin-producing isolate is given in parentheses. References for hosts of those species not mentioned in the previous chapters are as follows: 1: Welz et al., "Phenotypic Variation and Parasitic Fitness of Races of *Cochliobolus-carbonum* on Corn in North Carolina," *Phytopathology*, 83(6):593-601 (1993); Leonard et al., "Genetic Diversity in Field Populations of *Cochliobolus-carbonum* on Corn in North Carolina USA," *Phytopathology*, 80(11):1154-1159 (1990) (for races 2 and 3 only). 2: Domsch et al., "Compendium of Soil Fungi, Vol. 1," New York, New York:Academic Press, pp. 216-222 (1980). 3: David et al., "Fungi on Plants and Plant Products," St. Paul, Minnesota:APS Press, p. 635 (1989); Thakur et al., "Characterization of a New Race of *Exserohilum-turcicum* Virulent on Corn With Resistance Gene HTN," *Plant Dis.*, 73(2):151-155 (1989). 4: Rao et al., "New Fungal Diseases on Some Weeds," *Indian Bot. Rep.*, 6(1):38 (1987); Bhat et al., "Unrecorded

- 65 -

- Pathogen on Bamboo Causing Blight in India," Curr. SCI. (BANGALORE), 58(20):1148-1149 (1989). 5: Yoder, "Toxins in Pathogenesis," Ann. Rev. Phytopathol., 18:103-129 (1980).
- 5 c. Genomic DNAs (from a previously prepared gel blot filter, Rose et al., "A Decarboxylase Required for Polyketide Toxin Production and High Virulence by *Cochliobolus heterostrophus*," 8th Int. Symp. Mol. Plant-Microbe Int., Knoxville, p. J-49 (1996) were probed with the 3.4 kb *CPSI* fragment cloned on p214B7 (Figure 2). "+" indicates a strong hybridization signal. All species hybridized to a large fragment (~23 kb).
- 10 d. Genomic DNAs selected from a lab collection were probed with the *CPSI* 3.2 kb fragment cloned on p214S1 (Figure 2). The size of fragments that hybridized to the probe is given in kb. The intensities of hybridization signals were similar to each other. N = not done.
- 15 e. Genomic DNAs were probed with the same *CPSI* fragment as in c. Gel blot is shown in Figure 26.

DNA manipulations and targeted disruption of the CPSI homolog in C. victoriae.

- Genomic DNAs for probing were prepared according to Yoder ("Cochliobolus heterostrophus, Cause of Southern Corn Leaf Blight," in 20 Sidhu, ed., Genetics of Plant Pathogenic Fungi, Vol. 6, San Diego, California: Academic Press, pp. 93-112 (1988), which is hereby incorporated by reference), or selected from a lab DNA collection (stored at 4°C). A gel blot filter bearing known genomic DNAs was also probed. Plasmid DNA preparation, restriction enzyme digestions, gel electrophoresis, gel blot analysis were done 25 using standard protocols (Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor, New York: Cold Spring Harbor Laboratory Press (1989), which is hereby incorporated by reference). For probing, *CPSI* fragments of *C. heterostrophus* cloned on p214B7 (3.4 kb left flank) and p214S1 (3.2 kb right flank) (Figure 2) were prepared by restriction enzyme digestion of 30 the plasmid DNAs followed by purification using the QIAquick Gel Extraction Kit (QIAGEN Inc., Chatsworth, CA). The plasmid p118B14, which carries the 2.2 kb *Bgl* II fragment of *CPSI* interrupted by the *hygB* cassette (see Figure 5) was linearized with *Bgl* II and introduced into HvW genome. Transformants were purified by isolation of single conidia and genomic DNAs were digested with 35 *Bgl* II and probed with the *CPSI* 3.2 kb fragment.

Bioassays. Pathogenicity was determined by an oat plant assay. Fungal strains were grown on individual oat meal agar medium plates (60 X 15 mm) containing hygromycin B (60 ug/ml) for 10 days at 24°C under lights.

- 66 -

Conidia were scraped from the plates and suspended in 6 ml of sterilized distilled water. One ml of conidial suspension of each strain was mixed with 60 seeds of susceptible or resistant oats. Inoculated seeds were planted in 4" X 6" pots and seedlings were allowed to grow for two weeks. Seed germination rate and 5 symptom development were recorded at different stages (4, 6, 8 and 24 days after inoculation). Detection of victorin production using HPLC analysis was done by Alice Churchill in Dr. Vladimir Macko's lab at Boyce Thompson Institute for Plant Research.

10 **EXAMPLE 9 -- Detection of CPS1 homologs.**

Genomic DNAs of 12 isolates (or lab strains) of 9 fungal species hybridized to *CPS1* (Table 4). All 6 *Cochliobolus* species, including 4 known plant pathogens (*C. carbonum*, *C. victoriae*, *C. sativus* and *C. spicifer*) and 2 species with unknown hosts (*C. homomorphus* and *C. dactyloctenii*) gave 15 hybridization signals of the same intensity as that of *C. heterostrophus CPS1* fragments (Figure 26, only *C. carbonum* and *C. victoriae* are shown). Two phytopathogenic *Setosphaeria* species and *Bioplaris sacchari*, a sugarcane pathogen gave a similar hybridization intensity.

CPS1 homologs appear to be polymorphic among different species, 20 i.e., all species gave one or two unique bands when *BglII* or *HindIII* digested genomic DNAs were probed (except for *C. victoriae*, which showed the same hybridization pattern as *C. carbonum*) (Table 4 and Figure 26). Interestingly, *EcoRI* digested genomic DNAs of the same species did not show polymorphisms; all species hybridized to a large fragment (~23 kb, Table 4), indicating the absence 25 of an *EcoRI* site in all *CPS1* homologs as in the *C. heterostrophus* gene. In *C. heterostrophus*, a > 12 kb of genomic region which includes *CPS1* (5.4 kb), *TES1* (1.1 kb) and sequence downstream of the 3' end of *CPS1* has no *EcoRI* sites. In contrast to species-dependent polymorphisms, *CPS1* homologs appear to be highly conserved among different isolates of the same species. Both *C. heterostrophus* race T and race O hybridized to the same 4.2 kb *BglII* fragment (or 30 5.2 and 3.2 kb *HindIII* fragments); all three *C. carbonum* races hybridized to the same 5.0 kb *BglII* fragment (or 6.6 kb *HindIII* fragment) (Table 4, Figure 26) and

- 67 -

B. sacchari isolates 764-1 and 1249-10 hybridized to the same *Hind*III fragments (5.4 and 2.5 kb) (Table 4).

EXAMPLE 10 -- Targeted disruption of *CPSI* homolog in *C. victoriae*.

5 Twenty transformants were obtained from transformation of the victorin-producing isolate HvW with *Bgl*II-linearized plasmid p118B14 (Figure 5). Six transformants were purified and assayed for both victorin production and pathogenicity to susceptible oat plants. All transformants produced wild type levels of victorin as determined by HPLC analysis (Figure 27), but four of them
 10 (Tx7, Tx2, Tx5 and Tx8) showed dramatically reduced virulence in the plant assay. The seed germination rate on the eighth day after inoculation is only 13-25% for wild type and two transformants (Tx9 and Tx4), but 45-63% for the other four transformants. On day 24 after inoculation, all plants emerged from the seeds inoculated with wild type, Tx9 or Tx4 were killed but most (29-63%) from the
 15 seeds inoculated with Tx2, Tx7, Tx5 or Tx8 still survived (Table 5, Figure 28A). Gel blot analysis confirmed that transformants showing the reduced virulence phenotype resulted from homologous integration of the transforming vector that disrupted the wild type *CPSI* homolog in *C. victoriae* genome; transformants showing the wild type phenotype resulted from ectopic integration events that left
 20 the native gene intact (Figure 28B). All transformants remained nonpathogenic to resistant oats, indicating that disruption of the *CPSI* homolog does not affect host specificity of the fungus.

25 Table 5. Disease development of oat plants inoculated with *C. victoriae* transformants (Tx).

Strain ^a	<u>No. germinated^b</u>			Germination Rate (%) ^c	<u>No. survivors^d</u>	
	4	6	8		24	%
Control-1	28	41	45	75	75	100
Control-2	40	50	50	83	50	100
Control-3	1	7	12	20	0	0
Tx2	8	26	27	45	16	59
Tx4	5	15	15	25	0	0

- 68 -

Tx5	2	24	28	47	8	29
Tx7	14	36	38	63	24	63
Tx8	7	29	29	47	13	47
Tx9	0	3	8	13	0	0

- a. Control-1 = uninoculated susceptible oat seeds. Control-2 and Control-3 = resistant and susceptible oat seeds inoculated with wild type *C. victoriae* (isolate HvW), respectively. Six transformants were tested on both resistant and susceptible seeds, but only data for the later are shown (all transformants gave the same results as Control-2 when tested on resistant seeds). Repeat experiments gave similar results (data not shown).
- 5 b. Sixty oat seeds were used for each strain. Emerged oat plants were counted 4, 6 and 8 days after inoculation.
- c. Calculation based on the data collected on the day 8.
- 10 d. Recorded on day 24 after inoculation. The percentage of survivors is based on the number of plants recorded on days 8 and 24.

EXAMPLE 11 -- Isolation of CPSI genes from other plant pathogens

15 As disclosed in the previous examples, the *Cochliobolus heterostrophus* gene *CPSI* encodes a peptide synthetase that appears to be a general factor for fungal virulence to their hosts. Thus, *CPSI* has been found to be highly conserved among at least 9 fungal species belonging to 3 genera including the genus *Cochliobolus* and closely related genera *Bioplaris* and *Setosphaeria*; it has been demonstrated to be required for pathogenesis of three different plant pathogens i.e., *C. heterostrophus* race O , race T to corn and *C. victoriae* to oats (Lu, 1998, "Molecular-genetic analysis of general and specific pathogenesis factors in *Cochliobolus heterostrophus*," Ph.D thesis, Cornell University).

20 To further explore the role of *CPSI* in fungal pathogenesis and its conservation in other fungi, genomic DNAs of additional species in *Cochliobolus* and other closely or distantly related genera were probed with *ChCPSI* by DNA-DNA hybridization (Lu, S.-W., B. G. Turgeon and O. C. Yoder. 1999. "A gene cluster from the corn pathogen *Cochliobolus heterostrophus* required for nonribosomal peptide biosynthesis and general virulence of fungi." Fungal Genetics Conference, March 1999, Pacific Grove, California). Genomic DNAs of 40 filed isolates (or lab strains) representing 34 fungal species belonging to 16 genera hybridized when probed with *ChCPSI* (Figs. 30A-30C).

- 69 -

Fungal genomic DNAs were prepared according to a previously described procedure (Yoder, 1988. "Cochliobolus heterostrophus, cause of Southern Corn Leaf Blight". Genetics of Plant Pathogenic Fungi. G. S. Sidhu. San Diego, Academic Press. 6: 93-112). Plasmid DNA preparations, restriction enzyme digestions and preparation of DNA gel blots were performed following standard protocols (Sambrook, J., E. F. Fritsch, et al. 1989 "Molecular Cloning: A Laboratory Manual, 2nd Edition". Cold Spring Harbor, NY, Cold Spring Harbor Laboratory Press). A 3.2 kb *ChCPS1* fragment (corresponding to ChCPS1 amino acids 173-1208) was obtained by restriction enzyme digestion of a plasmid clone p214S1 (Lu, 1998, Ph.D thesis), (see Figure 2), followed by purification using the QIAquick Gel Extraction Kit (QIAGEN Inc., Chatsworth, CA). The purified *ChCPS1*/fragment was labeled with α -[³²P]dCTP (Turgeon, B. G., H. Bohlmann, et al, 1993. "Cloning and analysis of the mating type genes from *Cochliobolus heterostrophus*." Mol. Gen. Genet. 238: 270-284.). DNA-DNA hybridization was carried out at 62° C in 6 X SSC, 0.05 X BLOTO (Sambrook et al., 1989). Filters were washed in 2 X SSC, 0.1% SDS at 62° C for 60 minutes.

All 16 *Cochliobolus* species, including the known plant pathogens *C. carbonum*, *C. victoriae*, *C. miyabeanus*, *C. sativus* and *C. spicifer*, and five genera closely related to *Cochliobolus*, i.e., *Pyrenophora*, *Setosphaeria*, *Bipolaris*, *Stemphyllium* and *Alternaria* showed hybridization intensities comparable to that of *C. heterostrophus* itself (Fig. 30A).

DNAs of species from nine distantly related genera, including several of economic importance (e. g., *Magnaporthe grisea*, *Fusarium graminearum*, *Gaeumannomyces graminis*) or of medical importance (e. g., *Candida albicans*) hybridized weakly to *CPS1* (FigS. 30B, 30C) whereas no signal was detected in DNA of the basidiomycete *Ustilago maydis*.

Three *CPS1* homolog genes were cloned and characterized. Three of them were cloned from phytopathogenic fungi, including the wheat head scab fungus *Fusarium graminearum* (*FgCPS1*, 6003 bp, SEQ. No. 43), the potato early blight fungus *Alternaria solani*, (*AsCPS1*, 2369bp, SEQ. No. 41) and the barley net blotch fungus *Pyrenophora teres* (*PtCPS1*, 2306 bp, SEQ. No. 45). *FgCPS1* was cloned as a full length gene using both PCR amplification and the plasmid rescue procedure that was preceded by targeted

- 70 -

gene disruption in the genome. *AcCPSI* and *PiCPSI* homologs were partially cloned by direct PCR amplification.

The Polymerase Chain Reaction (PCR) was carried out using degenerate primers designed to conserved regions of *C. heterostrophus CPSI* (*ChCPSI*). Two sets of degenerate primers were designed to amino acids at or close to conserved core sequences of *C. heterostrophus CPSI* (*ChCPSI*). The first pair of primers: 5'TGYTTYATHGCNGGN GTNGTNGCNGTNCC3' (CHFP6, corresponding to positions 493-521 of *ChCPSI*) and 5'YTGYTGNGGNGGNCCNCCNGGRTT3' (CHRP4, 2197-2220 of *ChCPSI*), was used to amplify *CPSI* from *Fusarium graminearum*. The second pair of primers: 5'-AARAARAARGGNCCNACNGAG-3' (FP4CB, corresponding to positions 1531-1550 of *ChCPSI*) and 5'SRYTGNA CCCADATYTCNCC3' (RP2DB, corresponding to positions 3883-3902 of *ChCPSI*), was used to amplify *CPSI* from *A. solani* and *Pyrenophora teres*. PCR was carried out in a Perkin Elmer Cetus 9600-thermocycler with fungal (*Fusarium graminearum*, *Alternaria solani*, and *Pyrenophora teres*) genomic DNA as a template.

Reaction mixtures contained about 500 ng of genomic DNA in 100 ul of reaction buffer [1 x *Ex Taq* buffer, 0.2 mM dNTPs, 0.2 uM of each primer and 0.05 U/ml *Takara Ex Taq* (Pan Vera Corporation)]. An initial denaturing step of 95 °C for 3 min. was followed by 30 cycles of 94 °C for 1 min, 47 °C (for *G. zae*) or 55 °C (for *A. solni* and *Pyrenophora teres*) for 3 min, and 72°C for 3 min. Reactions were cooled to 4 °C after a final extension of 72 °C for 10 min. PCR products were examined (10 ul of each reaction) by agarose-gel (0.75%) electrophoresis.

DNA from isolated clones of the three different fungi was sequenced at the Cornell DNA Sequencing Facility using TaqCycle automated sequencing with DyeDeoxy terminators (Applied Biosystems, Foster City, CA). Primers used for sequencing were designed using Primer Select (DNASTAR) and synthesized by the Cornell Oligonucleotide Synthesis Facility. Sequencing of each plasmid clone was initiated with vector-specific primers or primers designed to previously determined sequences. Sequences were analyzed using MapDraw and MegAlign (DNASTAR) and nucleotide or

- 71 -

protein database searches were performed with the BLAST program (Altschul et al., 1990, 1997).

The *FgCPS1* open reading frame (5125 bp, SEQ. No. 43) has 50% nucleotide identity to *ChCPS1* (SEQ ID No.2) in about 4.4 kb of overlap (Fig. 2). No "TATA" box-like element was found in the 5' untranslated region, but other promoter sequences including two putative "CAAT" boxes and a "CT" motif were located upstream of the start codon (ATG) in *FgCPS1* (Fig. 32). Only one putative intron was found 1508 bp upstream the stop codon (TGA) in contrast to three in *ChCPS1* (Figs. 31A, 31B and 32). A putative polyadenylation signal "AATAA" was located 62 bp downstream of the stop codon (Fig. 32). The predicted *FgCPS1* protein (1692 amino acids, M_r 187983 Da, SEQ ID No. 44) has 68% identity, 73% similarity to *ChCPS1* (SEQ ID No. 3) in a about 1,500 amino acid overlap (Figs. 31A and 31B) that contains two structurally similar modules highly similar to those of *ChCPS1*. *FgCPS1* has no significant similarity to *ChCPS1* at the C-terminus, which is relatively shorter and lacks the thioesterase domain as seen in *ChCPS1* (Figs. 31A and 31B). The annotated *FgCPS1* sequence is given in Figure 32.

AsCPS1 (2369 bp, SEQ. No. 41) has 76% nucleotide identity to *ChCPS1* (SEQ ID No.2) in the entire cloned region which contains two conserved introns (Figs. 31A and B). The translated *AsCPS1* protein (partial) includes 758 amino acids corresponding to amino acids 511-1269 in *ChCPS1* and has up to 93% identity, 95% similarity to *ChCPS1* (Fig. 2). The annotated *AsCPS1* sequence is given in Figure 33.

PtCPS1 (2306 bp, SEQ. No. 45) has 78% nucleotide identity to *ChCPS1* (SEQ ID No.2) in the entire cloned region which contains only one intron (Fig. 2). The translated *PtCPS1* protein (partial) includes 758 amino acids corresponding to amino acids 511-1269 in *ChCPS1* and has 93% identity, 96% similarity to *ChCPS1* (Fig. 2). The annotated *PtCPS1* sequence is given in Figure 34.

30

EXAMPLE 12 -- Targeted disruption of CPS1 homolog in *F. graminearum*.

A 2.2 kb XbaI fragment from pUCATPH (Lu, et al, 1994) containing the bacterial hygmycin resistance gene (hygB) driven by the

- 72 -

Aspergillus nidulans trpC promoter was inserted into the *Xba*I site of a PCR clone pFgC8, which carries the 1.0 kb internal fragment of *FgCPSI*, to create pFgC8-hygB. This construct was transformed (in a circular form or linearized with *Hind*III or *Bgl*II) into an isolate (GZ3639) of wild type *F. graminearum*. Twenty 5 transformants were obtained by transformation of a wild type isolate (GZ3696) with circular plasmid pFgC8-hygB. DNA gel blot analysis of eight such transformants confirmed that the *CPSI* homolog in *F. graminearum* was disrupted by a single cross over recombination (Fig. 35A). Two transformants were obtained when the *Bgl*II-digested pFgC8-hygB was used and only one recovered 10 when the plasmid was digested with *Hind*III. Gel blot analysis indicated that all three transformants obtained using linearized plasmid integrated into the genome at a ectopic location that left the wild type *FgCPSI* gene intact (Fig. 35A). For virulence assays, *F. graminearum* strains were grown on PDA (or PDA plus hygromycin B for transformants) plates (100 X 15 mm) for 7-15 days at 24 C 15 under black lights (Sylvania Inc., Danvers, MA) for maximum conidiation. A susceptible spring wheat cultivar, Norm Hard Red (kindly provided by G. Bergstrom, Cornell university) was used. Two months old wheat plants grown in the greenhouse at anthesis (10 plants in one 4" X 6" pot) were sprayed with 10 ml conidial suspensions (10^5 or 10^4 conidia/ml) using a pressurized Preval Spray 20 Gun Power Unit thin layer chromatography sprayer (Alltech Associates, Deerfield, IL) or "injected" into the spikelets (5-10 μ l/ per spikelet) using a Benchmate pipettor (NICHIRYO, Japan). The inoculated plants were incubated in a mist chamber for 48 hours (23° C) and then transferred to a growth chamber (23° C, 14 hours of light). Mutant phenotypes were identified by the comparison of the 25 number of infected or "bleached" spikelets on each head of wheat plants.

Symptoms were recorded 7-10 days after inoculation. All *cpsI*⁻ disruptants had reduced virulence on wheat plants in the assays while ectopic transformants caused disease symptoms indistinguishable from those of wild type (Fig. 35B).

Although the invention has been described in detail for the 30 purposes of illustration, it is understood that such detail is solely for that purpose, and variations can be made therein by those skilled in the art without departing

- 73 -

from the spirit and scope of the invention which is defined by the following claims.

WHAT IS CLAIMED IS:

1. An isolated nucleic acid molecule from a plant pathogen encoding a CPS1 synthetase, wherein the nucleic acid molecule comprises a nucleotide sequence which hybridizes under stringent conditions to a nucleic acid molecule having a sequence as set forth in SEQ ID No. 41 or a complement thereof; or wherein the nucleic acid molecule comprises a nucleotide sequence as set forth in SEQ ID No. 41.
- 10 2. The isolated nucleic acid molecule according to claim 1 wherein the plant pathogen is selected from the group consisting of *Alternaria solani*, *Alternaria Alternaia alternatherae*, *A. alternata*, *A. amaranthi*, *A. araliae*, *A. brassicae*, *A. brassicicola*, *A. camelliae*, *A. cassiae*, *A. cheiranthi*, *A. cinerariæ*, *A. gossypii*, *A. helianthi*, *A. helianthinficiens*, *A. mali*, and *A. raphani*.
- 15 3. The isolated nucleic acid molecule according to claim 2 wherein the plant pathogen is *Alternaria solani*.
- 20 4. An isolated nucleic acid molecule coding for a polypeptide having the amino acid sequence as set forth in SEQ ID No. 42.
5. An isolated nucleic acid molecule comprising the nucleotide sequence as set forth in SEQ ID No. 41.
- 25 6. An isolated polypeptide encoded by the nucleic acid molecule of claim 1.
- 30 7. The polypeptide according to claim 6, wherein the polypeptide has an amino acid sequence which has at least 75 % similarity to the amino acid sequence of SEQ. ID. No. 42 as determined by a BLAST program with the default parameters.

WHAT IS CLAIMED IS:

1. An isolated nucleic acid molecule from a plant pathogen encoding a CPS1 synthetase, wherein the nucleic acid molecule comprises a nucleotide sequence which hybridizes under stringent conditions to a nucleic acid molecule having a sequence as set forth in SEQ ID No. 41 or a complement thereof; or wherein the nucleic acid molecule comprises a nucleotide sequence as set forth in SEQ ID No. 41.
- 10 2. The isolated nucleic acid molecule according to claim 1 wherein the plant pathogen is selected from the group consisting of *Alternaria solani*, *Alternaria Alternaia alternatherae*, *A. alternata*, *A. amaranthi*, *A. araliae*, *A. brassicae*, *A. brassicicola*, *A. camelliae*, *A. cassiae*, *A. cheiranthi*, *A. cinerariae*, *A. gossypii*, *A. helianthi*, *A. helianthinficiens*, *A. mali*, and *A. raphani*.
- 15 3. The isolated nucleic acid molecule according to claim 2 wherein the plant pathogen is *Alternaria solani*.
- 20 4. An isolated nucleic acid molecule coding for a polypeptide having the amino acid sequence as set forth in SEQ ID No. 42.
5. An isolated nucleic acid molecule comprising the nucleotide sequence as set forth in SEQ ID No. 41.
- 25 6. An isolated polypeptide encoded by the nucleic acid molecule of claim 1.
- 30 7. The polypeptide according to claim 6, wherein the polypeptide has an amino acid sequence which has at least 75 % similarity to the amino acid sequence of SEQ. ID. No. 42 as determined by a BLAST program with the default parameters.

- 75 -

8. The polypeptide according to claim 7 wherein the polypeptide comprises an amino acid sequence as set forth in SEQ ID No. 42.

9. An isolated nucleic acid molecule from a plant pathogen encoding
5 a CPS1 synthetase, wherein the nucleic acid molecule comprises a nucleotide sequence which hybridizes under stringent conditions to a nucleic acid molecule having a sequence as set forth in SEQ ID No. 43 or a complement thereof; or wherein the nucleic acid molecule comprises a nucleotide sequence as set forth in SEQ ID No. 43.

10

10. The isolated nucleic acid molecule according to claim 1 wherein the plant pathogen is selected from the group consisting of *Fusarium graminearum*, *Fusarium avenaceum*, *F. carpineum*, *F. chlamydosporum*, *F. coccophilum*, *F. culmorum*, *F. episphaeria*, *F. equiseti*, *F. flocciferum*, *F. moniliforme*, *F. oxysporum*, *F. redolens*, *F. sambucinum*, *F. solani*, *F. subglutinans*, *F. trichothecioides*, *F. udum*, or *F. ventricosum*.

11. The isolated nucleic acid molecule according to claim 10 wherein the plant pathogen is *Fusarium graminearum*.

20

12. An isolated nucleic acid molecule coding for a polypeptide having the amino acid sequence as set forth in SEQ ID No. 44.

13. An isolated nucleic acid molecule comprising the nucleotide sequence as set forth in SEQ ID No. 43.

14. An isolated polypeptide encoded by the nucleic acid of claim 9.

15. The polypeptide according to claim 14, wherein the polypeptide has an amino acid sequence which has at least 75 % similarity to the amino acid sequence of SEQ. ID. No. 44 as determined by a BLAST program with the default parameters.

16. The polypeptide of claim 15 wherein the polypeptide comprises the amino acid sequence as set forth in SEQ ID No. 44.

17. An isolated nucleic acid molecule from a plant pathogen encoding
5 a CPS1 synthetase, wherein the nucleic acid molecule comprises a nucleotide sequence which hybridizes under stringent conditions to a nucleic acid molecule having a sequence as set forth in SEQ ID No. 45 or a complement thereof; or wherein the nucleic acid molecule comprises a nucleotide sequence as set forth in SEQ ID No. 45.

10

18. The isolated nucleic acid molecule according to claim 1 wherein the plant pathogen is selected from the group consisting of *Pyrenophora teres*, *Pyrenophora avenae*, *P. bromi*, *P. leuceienes*, *P. phaeocomes*, *P. schroeteri*, *P. trichostoma*, or *P. tritici-repentis*.

15

19. The isolated nucleic acid molecule according to claim 18 wherein the plant pathogen is *Pyrenophora teres*.

20

20. An isolated nucleic acid molecule coding for a polypeptide having the amino acid sequence as set forth in SEQ ID No. 46.

21. An isolated nucleic acid molecule comprising the nucleotide sequence as set forth in SEQ ID No. 45.

25

22. An isolated polypeptide encoded by the nucleic acid of claim 17.

30

23. The polypeptide according to claim 22, wherein the polypeptide has an amino acid sequence which has at least 75 % similarity to the amino acid sequence of SEQ. ID. No. 46 as determined by a BLAST program with the default parameters.

24. The polypeptide of claim 23 wherein the polypeptide comprises the amino acid sequence as set forth in SEQ ID No. 46.

- 77 -

25. A vector comprising the nucleic acid molecule of any one of claims
1-5, 9-13, or 17-21.

26. A vector according to claim 25, wherein the nucleic acid molecule
5 is operably linked to a promoter.

27. A vector according to claim 26, wherein the nucleic acid molecule
is in a sense orientation.

10 28. A vector according to claim 26, wherein the nucleic acid molecule
is in an antisense orientation.

29. The vector according to claim 26, wherein the vector is capable of
maintaining and expressing the nucleic acid molecule in bacterial cells.

15 30. The vector according to claim 26, wherein the vector is capable of
maintaining and expressing the nucleic acid molecule in plant cells.

31. A host cell transformed with the vector according to claim 25.
20 32. A host cell according to claim 31, wherein the host is a plant.

33. A host cell according to claim 31, wherein the host is selected from
the group consisting of corn, oats, grasses, weeds, bamboo, and sugarcane.

25 34. A host cell according to claim 31, wherein the host is corn.

35. A plant transformed with the nucleic acid molecule according of
any one of claims 1-5, 9-13, or 17-21.

30 36. An isolated DNA molecule according to claim 1, wherein the
nucleic acid molecule has a nucleotide sequence which is 70% identical to the

- 78 -

nucleotide sequence of SEQ. ID. No. 41 as determined by a BLAST program with default parameters.

37. An isolated DNA molecule according to claim 1, wherein the
5 nucleic acid molecule is 80% identical to the nucleotide sequence of SEQ. ID.
No. 41 as determined by a BLAST program with default parameters.

38. An isolated DNA molecule according to claim 1, wherein the
nucleic acid molecule is 90% identical to the nucleotide sequence of SEQ. ID.
10 No. 41 as determined by a BLAST program with default parameters.

39. An isolated DNA molecule according to claim 1, wherein the
nucleic acid molecule has a nucleotide sequence which is 70% identical to the
nucleotide sequence of SEQ. ID. No. 43 as determined by a BLAST program
15 with default parameters.

40. An isolated DNA molecule according to claim 1, wherein the
nucleic acid molecule is 80% identical to the nucleotide sequence of SEQ. ID.
No. 43 as determined by a BLAST program with default parameters.

20 41. An isolated DNA molecule according to claim 21, wherein the
nucleic acid molecule is 90% identical to the nucleotide sequence of SEQ. ID.
No. 43 as determined by a BLAST program with default parameters.

25 42. An isolated DNA molecule according to claim 1, wherein the
nucleic acid molecule has a nucleotide sequence which is 70% identical to the
nucleotide sequence of SEQ. ID. No. 45 as determined by a BLAST program
with default parameters.

30 43. An isolated DNA molecule according to claim 1, wherein the
nucleic acid molecule is 80% identical to the nucleotide sequence of SEQ. ID.
No. 45 as determined by a BLAST program with default parameters.

- 79 -

44. An isolated DNA molecule according to claim 21, wherein the nucleic acid molecule is 90% identical to the nucleotide sequence of SEQ. ID. No. 45 as determined by a BLAST program with default parameters.

5 45. A method for identifying inhibitors of a CPS1 protein, wherein
said CPS1 protein is a peptide synthetase of a plant pathogen, said method
comprising:
 providing a CPS1 protein or polypeptide;
 contacting the protein or polypeptide with potential inhibitor
10 compounds;
 determining peptide synthetase activity; and
 selecting compounds which decrease the peptide synthetase
activity.

15 46. The method of Claim 45 wherein the CPS1 protein is from
Alternaria solani.

47. The method of Claim 45 wherein the CPS1 protein is from *Fusarium graminearum*.

48. The method of Claim 45 wherein the CPS1 protein is from *Pyrenoponera teres*.

49. A method of imparting disease resistance to a plant, said method comprising over-expressing a CPS1 polypeptide in the plant, wherein the polypeptide has protein synthetase activity.

50. A method according to claim 49, wherein the plant is selected from
a group consisting of corn, oats, grasses, weeds, sugarcane, barley, wheat, rice,
30 tomato, potato, citrus, malus, rye, cotton, brassica, cabbage, and carrot.

51. The method of claim 49 wherein the CPSI polypeptide is from *Alternaria solani*.

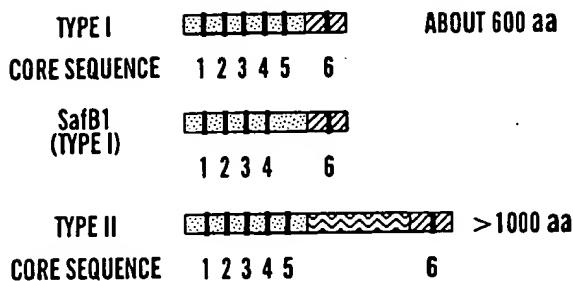
- 80 -

52. The method of Claim 49 wherein the CPS1 polypeptide is from
Fusarium graminearum.

53. The method of Claim 45 wherein the CPS1 polypeptide is from
5 *Pyrenophora teres*.

1/51

AMINO-ACID ACTIVATING MODULES

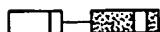


DOMAINS

-  AMINO ACYLADENYLATE FORMATION: CORES 1-5 (CORES 2, 3, AND 4: ATP BINDING; CORE 4: ATPase; CORE 1: UNKNOWN FUNCTION)
-  THIOESTER FORMATION (4' PHOSPHOPANTETHIN BINDING), CORE SEQUENCE 6 ONLY.
-  AMINO ACID N-METHYLATION (> 400 aa)

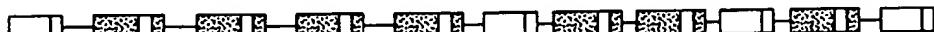
FIG. 1A

ENNIAVIN B



DHIV = D-2-HYDROXYISOVALERIC ACID

CYCLOSPORIN A

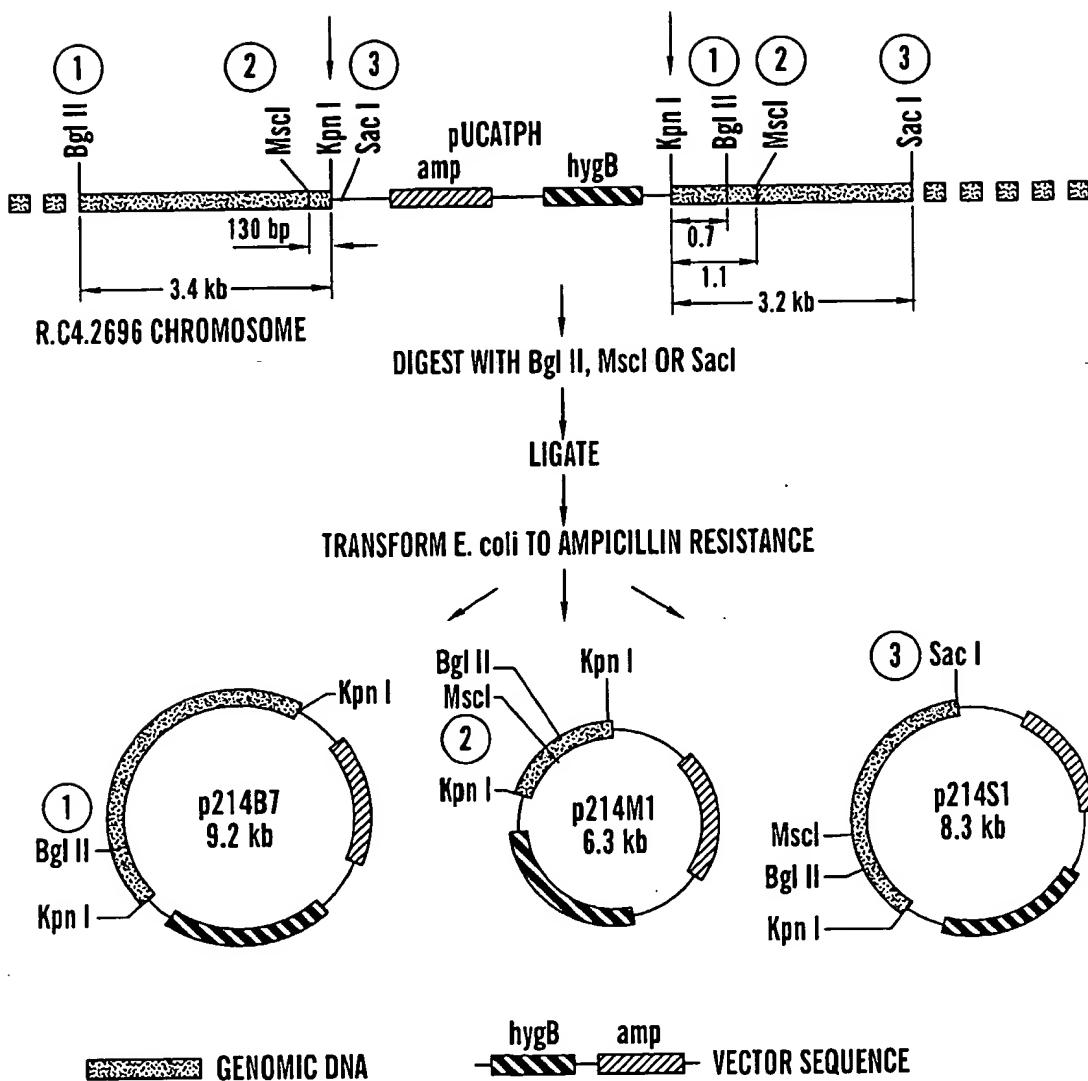


DAla—MeLeu—MeLeu—MeVal—MeBmt—Abu—Sar—MeLeu—Val—MeLeu—Ala

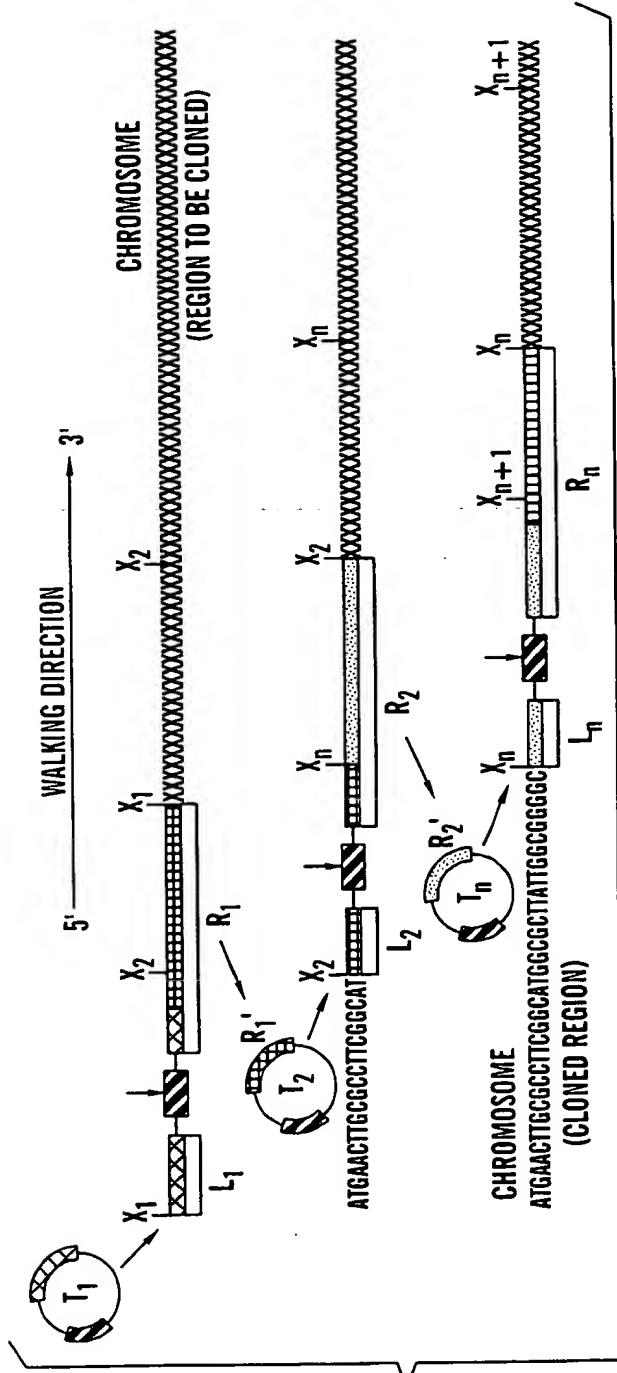
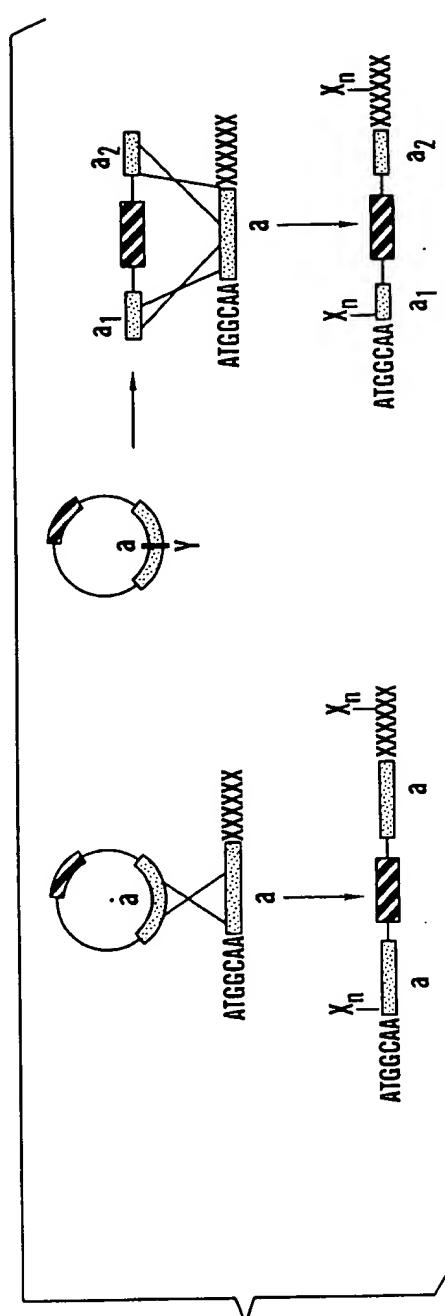
MeBmt = (4R)-4-[(E)-2-BUTENYL]-L-THREONINE

Abu = α -AMINO BUTYRIC ACID; Sar = SARCOSINE**FIG. 1B**

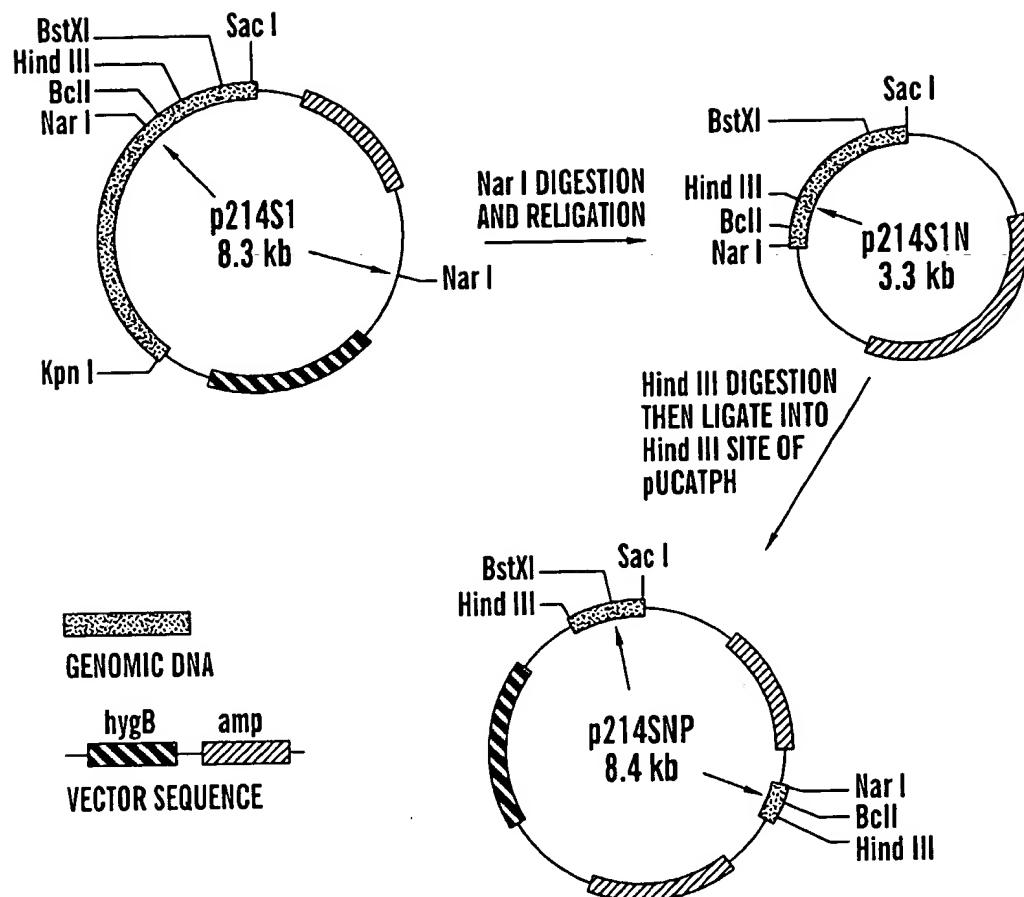
2/51

**FIG. 2**

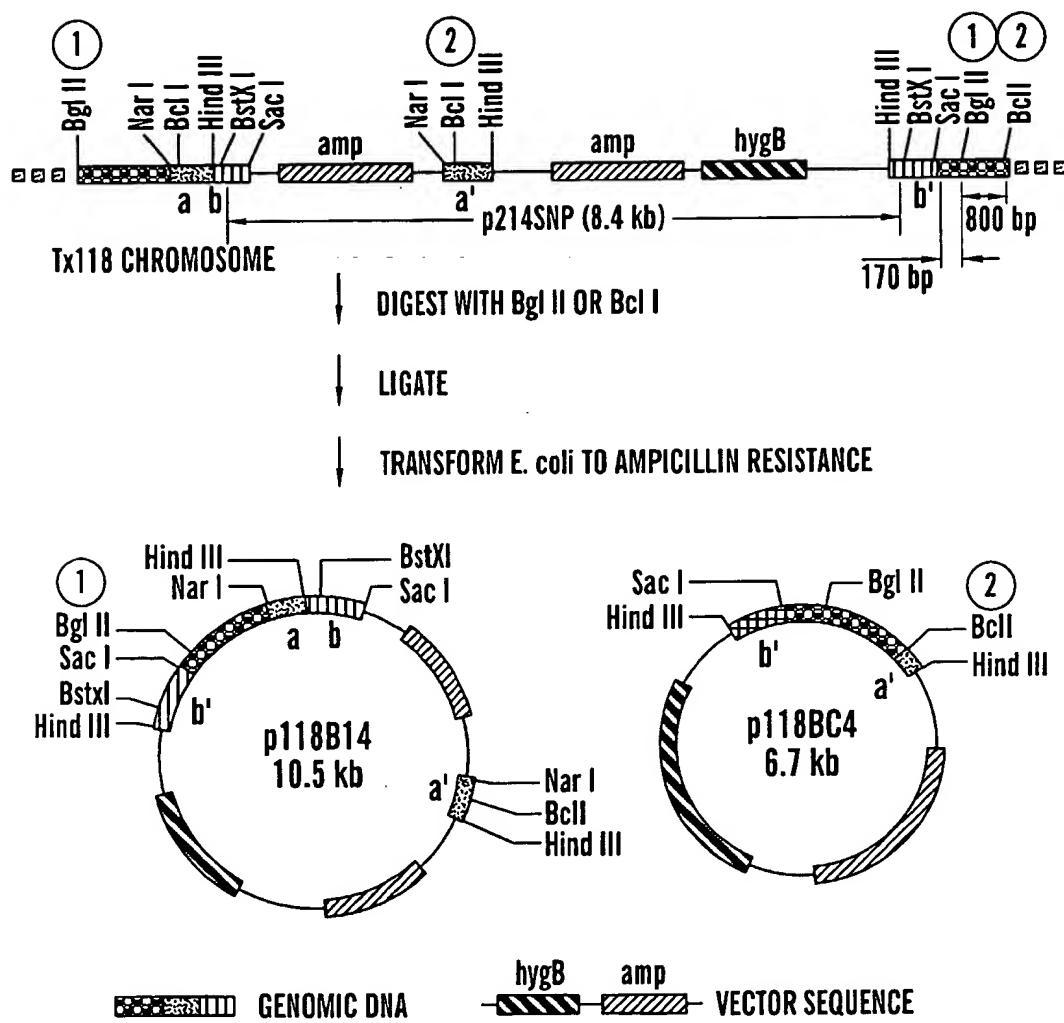
3/51

**FIG. 3A**

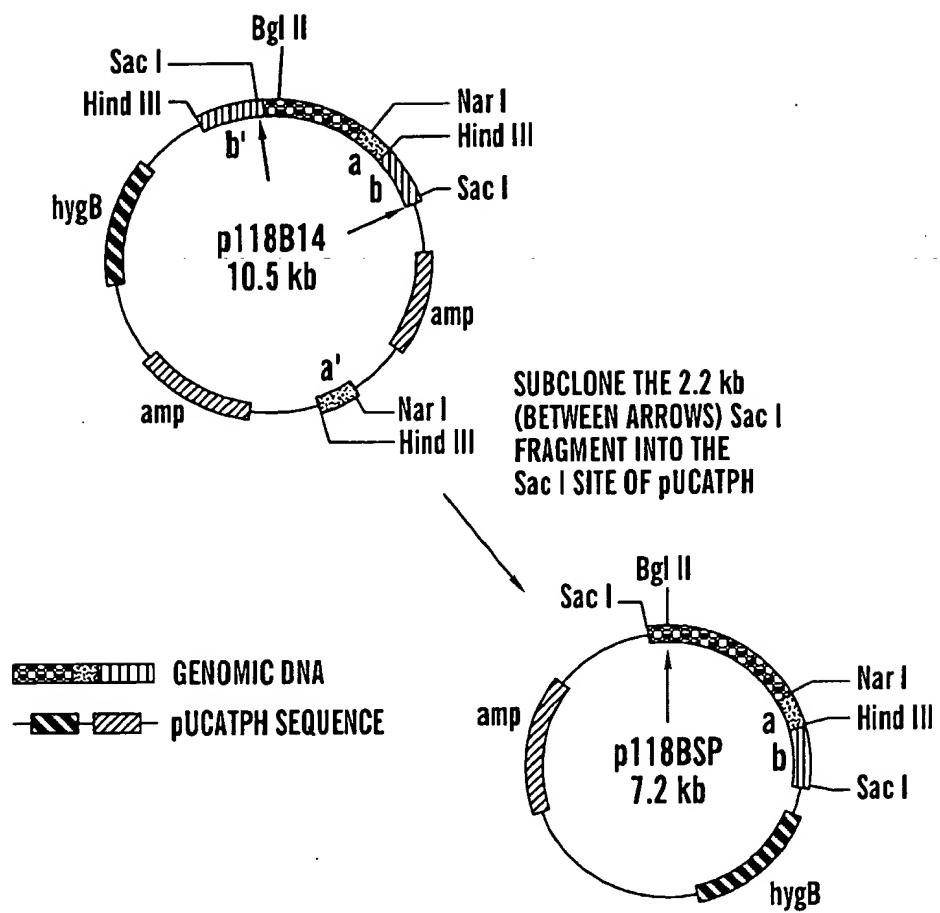
4/51

**FIG. 4**

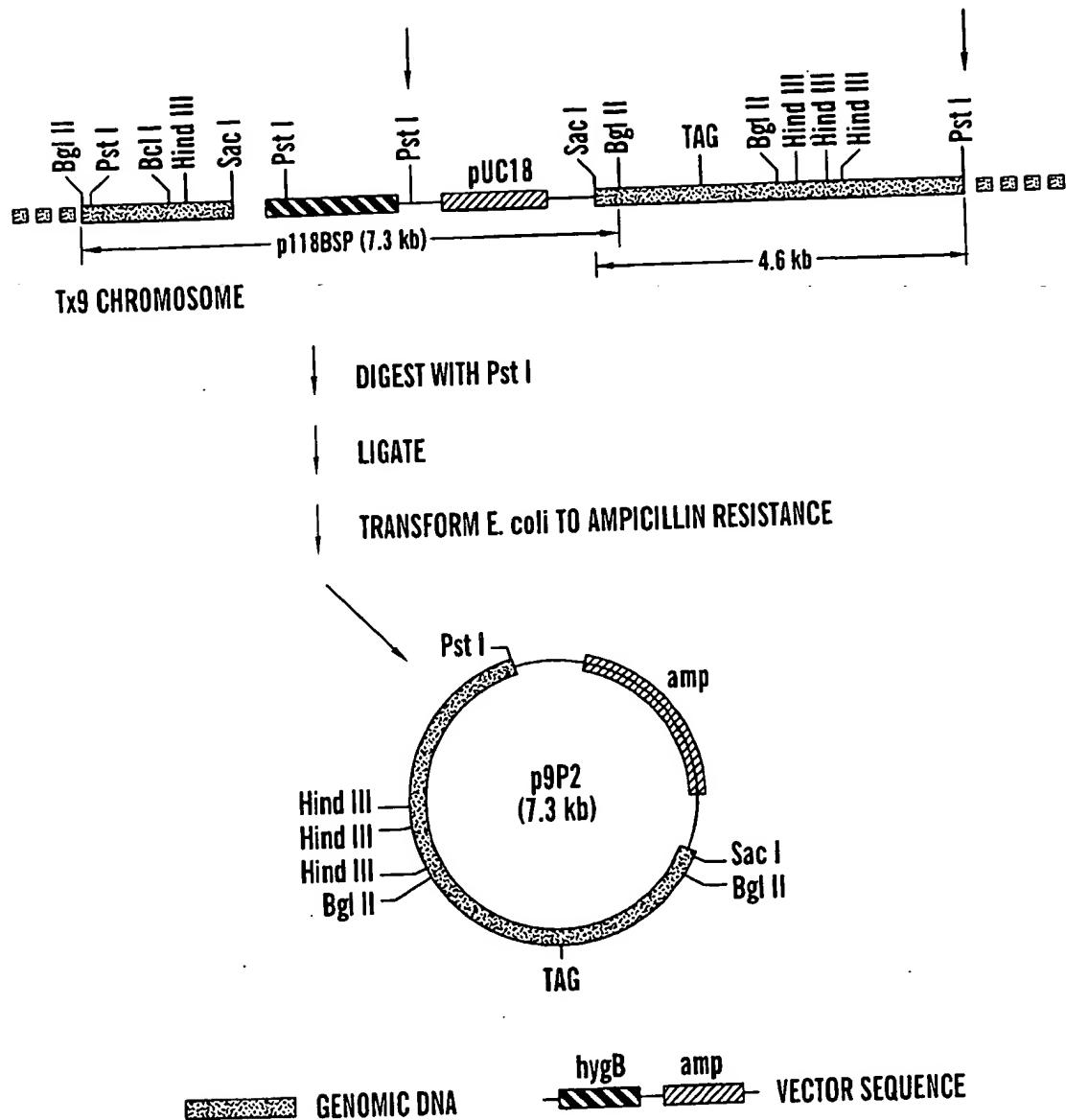
5/51



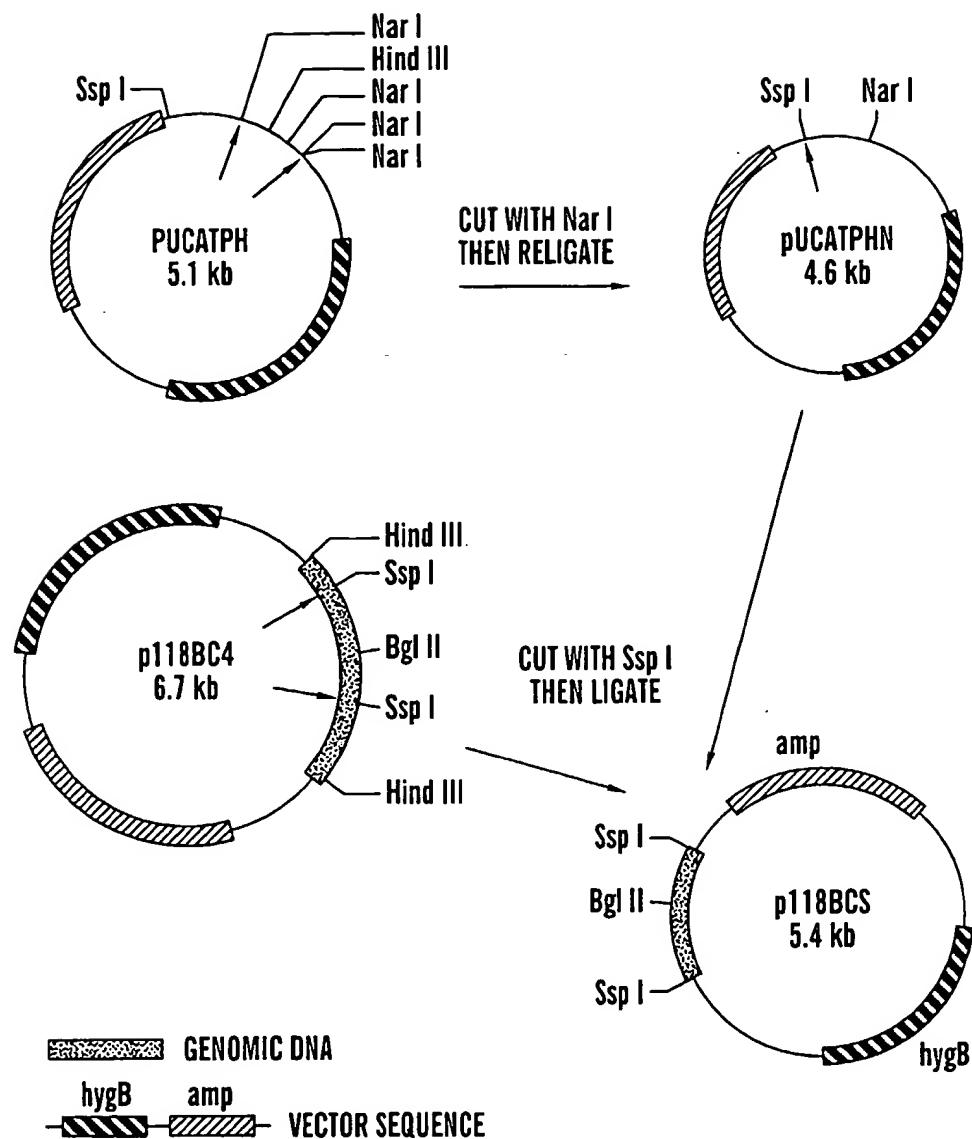
6/51

**FIG. 6**

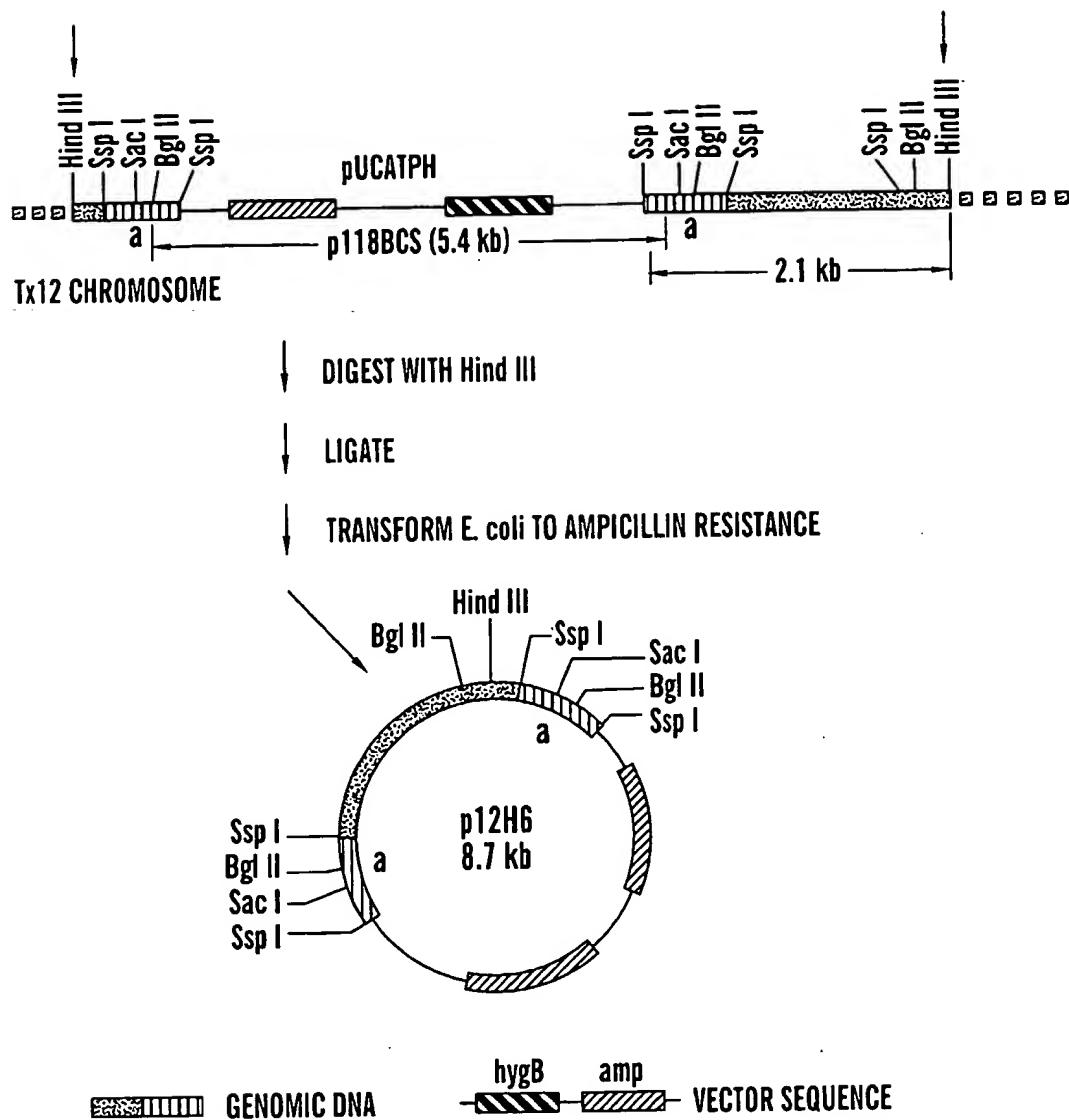
7/51

**FIG. 7**

8/51

**FIG. 8**

9/51

**FIG. 9**

10/51

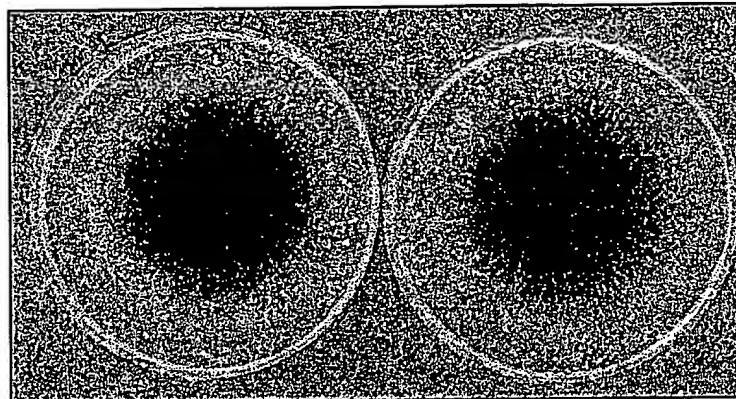


FIG. 10A

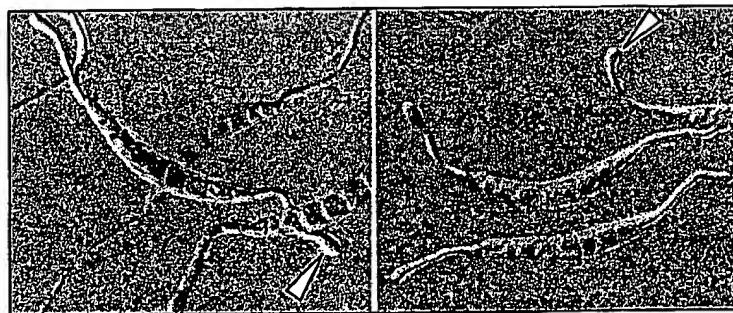


FIG. 10B

11/51

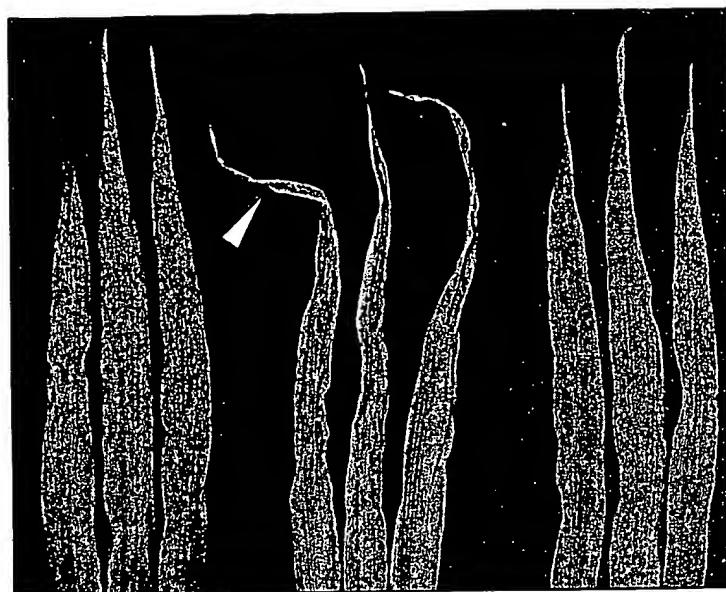


FIG. 11A

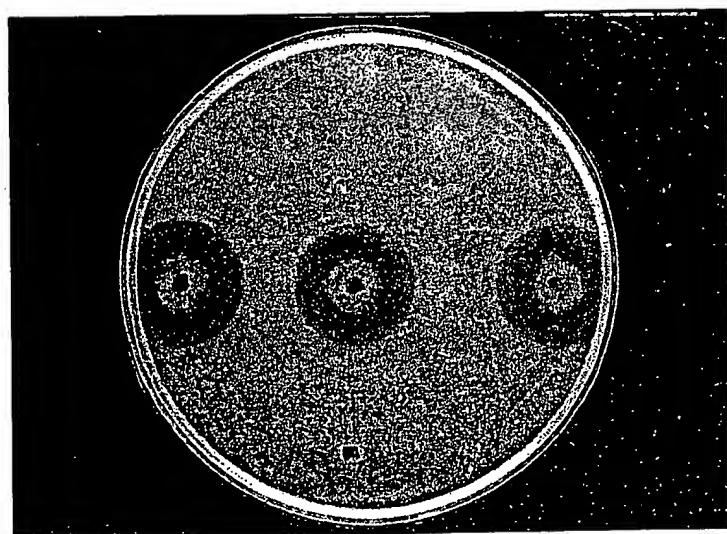


FIG. 11B

12/51

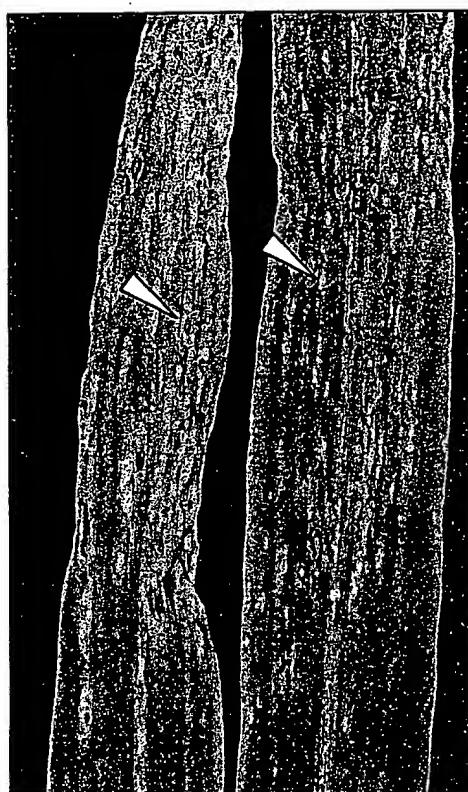
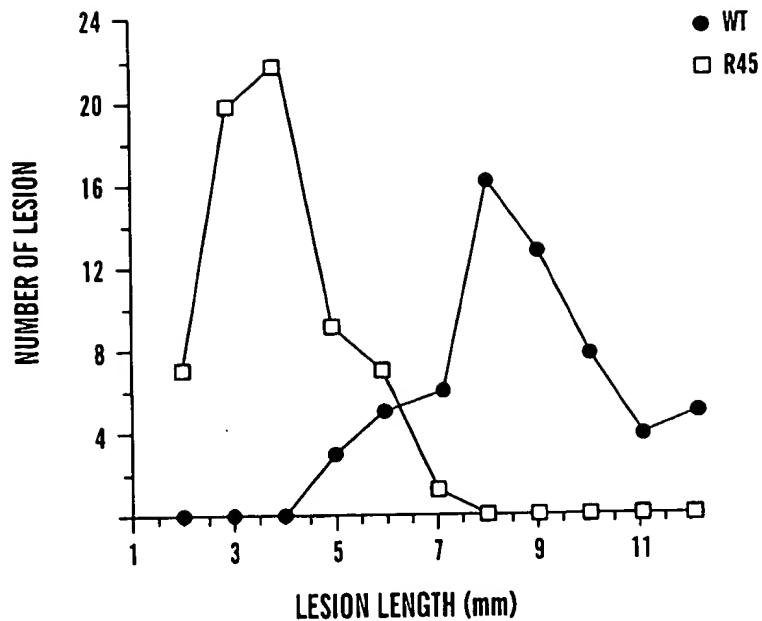


FIG. 12

13/51

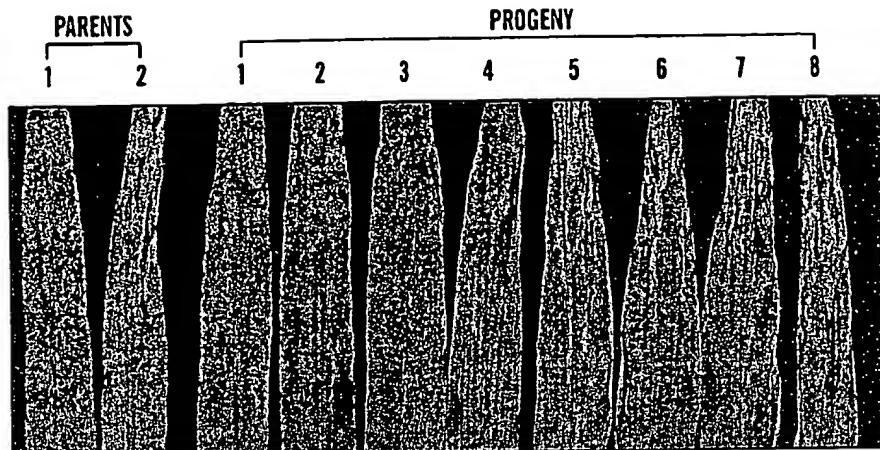
**FIG. 13A**

STRAIN	LESION SIZE (mm)			MEAN	SD	A*			
	1-4	5-8	9-12						
WT	0	52	48	8.5	1.0	A*			
R45	86	14	0	3.5	0.9	B			

* SIGNIFICANT DIFFERENCE AT P<0.01.

FIG. 13B

14/51

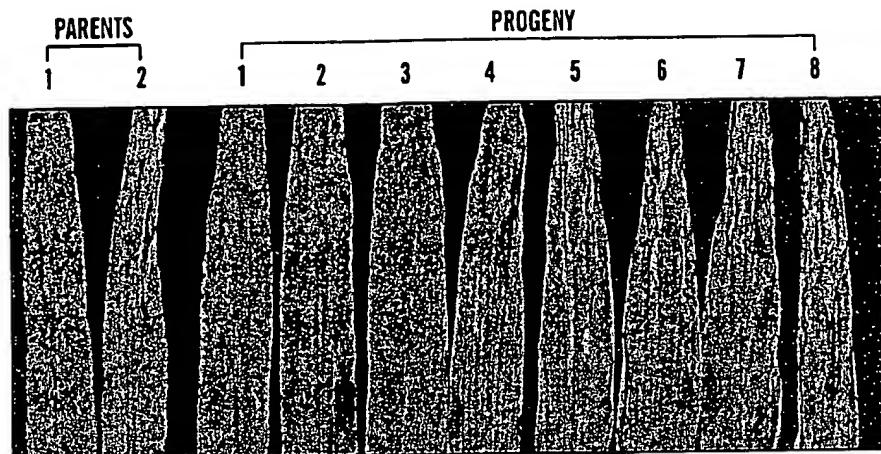
***FIG. 14A***

CROSS	PROGENY	PARENTAL TYPE		NONPARENTAL TYPE	
		path hyg ^R	PATH hyg ^S	path hyg ^R	PATH hyg ^S
R.C4.2696 X C5	RANDOM SPORES	24	22	0	0
1301-R33* X C5	tetrad1	4	4	0	0
	tetrad2	4	4	0	0
	tetrad3	4	4	0	0
	RANDOM SPORES	21	22	0	0

*13012-R33 (path, hyg^R, Tox, MAT-2) IS A PROGENY FROM THE FIRST CROSS, CARRYING THE R.C42696 MUTATION

FIG. 14B

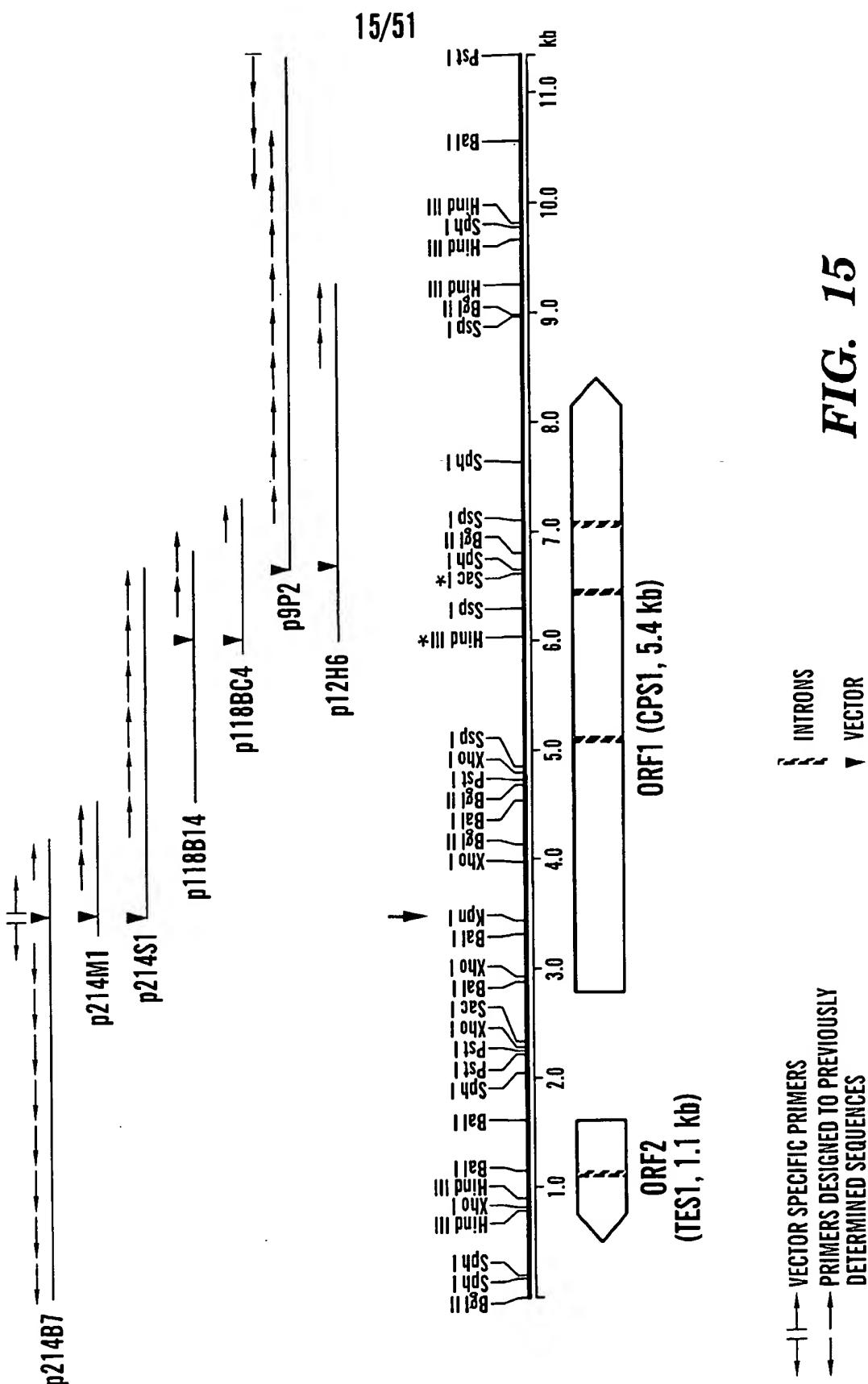
14/51

***FIG. 14A***

CROSS	PROGENY	PARENTAL TYPE		NONPARENTAL TYPE	
		path hygB ^R	PATH hygB ^S	path hygB ^R	PATH hygB ^S
R.C4.2696 X C5	RANDOM SPORES	24	22	0	0
1301-R33* X C5	tetrad1	4	4	0	0
	tetrad2	4	4	0	0
	tetrad3	4	4	0	0
	RANDOM SPORES	21	22	0	0

*13012-R33 (path, hygB^R, Tox, MAT-2) IS A PROGENY FROM THE FIRST CROSS, CARRYING THE R.C42696 MUTATION

FIG. 14B

**FIG. 15**

16/51

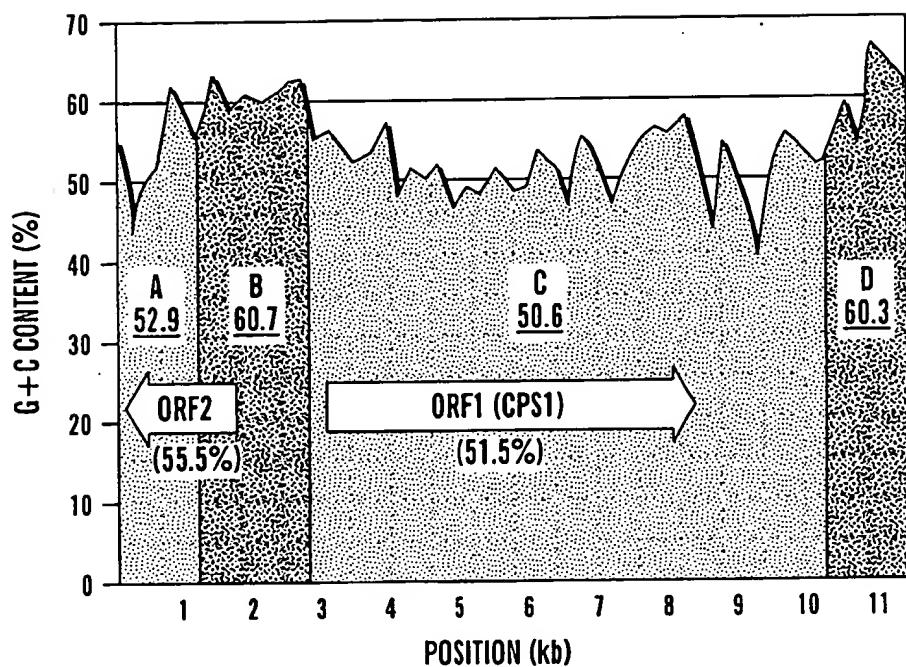


FIG. 16

17/51

TGCCTGCGCCTGTGCTTGTGCCGTGGAATGTCGGCCCGCTGCTGCAT	- 776
AGCCTATCTGTACATACAACACCACATCCCACATCCGCTCACCTGCCTTGCC	- 726
TCCCTCCTCGTGCCACACATCCGCCGCCACAACACCACATGGCTGCCACCA	- 676
ACCCCGAGCTGCAGGCCAAACTGCAGGAGCTGGACCACGAGCTCGAGGAG	- 626
GGCGATATTACACAAAAAGGGTCCGTACTGCTGCACCACCGCCATCC	- 576
GCCTCTCTGCGTGCCTAATCAGTCGATAGCTATGAAAAACGTCGCACC	- 526
GTGCTGCTGTCGCAGTATCTAGGGCCTGACTTTGCTGCCAGTTGCAGGC	- 476
CGACCTGAACCAGCAGAACCCACCCACCATCCAGTGAGGGCTTCGCT	- 426
CCCGCACCGCATCCTTGCTATTCCGTCCGGTCCGAGTCCATCACGGCGA	- 376
CCACAACCCCCACATATCCAGCTCCCCGCCCGACTCATACCATGACGC	- 326
TTCCGCACAGGGCCAATTGGGGCGACCCATGCCATATGCGAACGCCCTCG	- 276
CCGCTGCCTCGGGGGCTCGCAGTACATGGCATACCGCCAGCCAAGTC	- 226
GGCCGTTTCAAGAGAACGAGCTGGGCCTGCGTACAAATTGCTCCAGCG	- 176
****CAATTCTCACAGCTGTCGAAGGAAGCGAGACGTTATTCCACGGCCTC	- 126
AAACGCCCTGAATACAACCACTCGCGCAGGCCACCATGATGGCAACTAC	- 76
*****GCCTTCAATCCAGACAATCAGCAAAGTTATGATGGCCAATTGGCTCTCC	- 26
GGGAGAGGCCAGTCGAAGGAGCACCATGCTCGAGGTAACCAGGGTTATT	25
M L E V N Q G Y	
TTTCCGACTTCACAGGCCAGCAGATGCAAGACAATCGCGACTCGTATGGG	75
F S D F T G Q Q M Q D N R D S Y G	
GGACCCAACCGCTACTCGTCGGAGATGCCCTTCTCCTACCGCCCGAT	125
G P N R Y S S G D A F S P T A A I	
TCCACCTCCCATGATGAACCCAAACGATCTCCCTTGGCGCTGCTGAAA	175
P P P M M N P N D L P L G A A E	
CCATGATGCCGCTAGAGCCCCGCGATCTGCCTTTGACGTTACGACCCCT	225
T M M P L E P R D L P F D V Y D P	

FIG. 17A

18/51

CACAACCCCAATGTCAAAATGTCAAAGTTGACAACATTGGCGCTGTCTT	275
H N P N V K M S K F D N I G A V L	
GCGTCACCGAACGTCGACACAGCCAAGGACGACTGCCTCTGGTCCTTG	325
R H R S R T Q P R T T A F W V L	
ACGCAAAAGGCCAAGAGACGGCGTCCATCACCTGGGAAAAGGTGGCTAGT	375
D A K G K E T A S I T W E K V A S	
CGCGCGAAAAGGTGGCCAAAGTGAATTGGGACAAGAGCAACCTCTATCG	425
R A E K V A K V I R D K S N L Y R	
AGGCGACCGTGTGGCATTAGTGTACAGGGATACAGAAATCATTGATTTG	475
G D R V A L V Y R D T E I I D F	
TCGTGGCGTTGATGGGCTGCTTCATTGCGGGCGTTGTAGCGGTACCCATC	525
V V A L M G [REDACTED] [REDACTED]	
Core 1	
AATAGCGTCGACGACTACCAGAAACTCATTCTCTCCTAACGACAACCTCA	575
[REDACTED] D Y Q K L I L L L T T T Q	
AGCTCATCTCGCATTGACCACAGACAACATCTCAAGGCCTTCATCGTG	625
A H L A L T T D N N L K A F H R	
ACATTAGTCAGAACCGTCTGAAATGGCCGAGTGGGTAGAGTGGTGGAAAG	675
D I S Q N R L K W P S G V E W W K	
ACGAAACGAGTTGGCAGCCACCACCCCAAGAAACATGACGATACTCCAGC	725
T N E F G S H H P K K H D D T P A	
TTTGCAGTACCAAGAGGTTGCCTATATTGAGTTCTCGCGTGCACCTACTG	775
L Q V P E V A Y I E [REDACTED] [REDACTED]	
Core 2	
GTGACCTTCGCGGTGTGGTGCTTAGTCACCGGACTATTATGCACCAAATG	825
[REDACTED] [REDACTED] [REDACTED] M H Q M	
GCCTGCATCAGTGCATGATTAGCACGATAACCCACCAACGCTCAGAGCCA	875
A C I S A M I S T I P T N A Q S Q	
AGACACGTTCACTAGCCTACGGGATGCAGAGGGAAAGTTCGTTGCTC	925
D T F S T S L R D A E G K F V A	
CAGCACCGTCCAGAAACCCACAGAAGTGAATCCTCACGTACCTCGACCCG	975
P A P S R N P T E V I L T Y L D P	
CGCGAAAGCGCTGGTCTCATTCTCAGTGTCTTGTGAGTTATGGAGG	1025
R E S A G L I L S V L F A V Y G G	

FIG. 17B

19/51

CCACACCACCGTATGGCTCGAGACAGCGACCATGGAAACCCGGGTCTAT	1075
H T T V W L E T A T M E T P G L	
ATGCACATCTCATCACCAAATAACAAGTCCAACATACTGCTAGCGGATTAC	1125
Y A H L I T K Y K S N I L L A D Y	
CCAGGCCTCAAGCGCGCTGCATACAACATACCAACAGGATCCAATGGCTAC	1175
P G L K R A A Y N Y Q Q D P M A T	
AAGAAACTTCAAGAAAAACACAGAACCCAACTTCGCCCTCCGTGAAGATCT	1225
R N F K K N T E P N F A S V K I	
GTCTGATTGACACGCTTACCGTCGACTGTGAATTCACGAAATTCTCGGA	1275
C L I D T L T V D C E F H E I L G	
GATCGATATTCAGGCCACTCGCAAACCCCTAGAGCGCGAGAACTGATCGC	1325
D R Y F R P L R N P R A R E L I A	
GCCAATGCTCTGCTTGCCAGAACATGGTGGAAATGATAATATCTGTACGCG	1375
P M L C L P E H G G M I I S V R	
ACTGGCTAGGTGGAGAGGAGCGCATGGGCTGCCGCTAAGCATAAGCAGTA	1425
D W L G G E E R M G C P L S I A V	
GAAGAGTCAGATAATGATGAAGATGATACAGAGGATAAGTATGCAGCGC	1475
E E S D N D E D D T E D K Y A A A	
AAATGGCTACTCCAGTCTTATTGGTGGTGGCACTACAAAGAACAAAAAGG	1525
N G Y S S L I G G G T T K N K K	
AGAAGAAGAAGAAGGCCGACAGAGCTTACAGAAATCTTGCTGGACAAG	1575
E K K K K G P T E L T E I L L D K	
GAAGCTCTGAAGATGAACGAAGTCATTGTTCTGCCATTGGAGAAGAAC	1625
E A L K M N E V I V L A I G E E A	
AAGCAAGCGGGCAAACGAGCCGGCACCATGCGAGTCGGTGCCTTGGAT	1675
S K R A N E P G T M R V G A F G	
ACCCCATACCGGATGCACACTAGCTATTGTAGACCTGAGACAAGTCTT	1725
Y P I P D A T L A I V D P E T S L	
CTATGTTACCATACTCGATAGGCGAGATCTGGTAGATTGCCCTCACT	1775
L C S P Y S I [REDACTED] V D S P S L	
Core 3	
CTCTGGTGGCTTCTGGCAGCTGCAGAACATACAGAGACCATTTCATG	1825
[REDACTED] S G G E W Q L Q K H T E T I F H	

FIG. 17C

20/51

CTCGACCATAACCGTTCGTTGAGGGTAGCCCTACGCCACAGTTGCTTGAA	1875
A R P Y R F V G G S P T P Q L L E	
CTCGAGTTTCTGCGTACTGGACTCCTCGGCTTGTGTTAGAGGGAAAAAT	1925
L E F L [REDACTED] V V E G K I	
Core 4	
ATTTCCTTGGACTGTACGAAGATCGCATCAGACAGCGTGTGAATGGG	1975
F V [REDACTED] R D R D R V	
Core 5	
TAGAAAATGGTCAGCTTGAAGCCGAGCATCGATACTTTTGTGCAGCAC	2025
[REDACTED] A E H R Y F F V Q H	
CTGGTCACAAGCATTATGAAGGCCGTGCCAAAAATTACGACTGgttaagt	2075
L V T S I M K A V P K I Y D C	
gagctgccaaacagagcaaggact <u>gtctaacgtgtcatag</u> CTCGTCGTTG	2125
S S F	
ATTCTTATGTAATGGTGAATACCTGCCAATCATTCTCATCGAGACGCAG	2175
D S Y V N G E Y L P I I L I E T Q	
GCCGCATCGACTGCCAACAAACCCAGGTGGACCACCACAACAATTGGA	2225
A A S T A P T N P G G G P P Q Q L D	
TATACCATTTGGATTCACTATCTGAGAGGTGCATGGAGGTCTTACCA	2275
I P F L [REDACTED] S E R C M E V L Y	
Core 6	
AAGAGCATCTTACGGGTATACTGCGTGTGATTACAGCACCTAATACA	2325
Q E H H L R V Y C V M I T A P N T	
CTTCCACGAGTCATCAAGAACGGACGGCGAGAAATTGCAATATGCTGTG	2375
L P R V I K N G R R E I G N M L C	
TAGGAGAGAGTTGACAATGGCTCTGCCCTGTGTNCACGTNAAGTTG	2425
R R E F D N G S L P C V H V K F	
GCATTGAGCGATCAGTGCAGAACATTGCCTCGGTGACGATCCGCTGGC	2475
G I E R S V Q N I A L G D D P A G	
GGCATGTGGTCATTTGAGGCATCAATGGCACGTCAAGCAATTCTGATGCT	2525
G M W S F E A S M A R Q Q F L M L	
CCAAGACAAGCAATACTCTGGTGTGATCATCGCGAAGTCGTATTGACG	2575
Q D K Q Y S G V D H R E V V I D	
ACAGGACATCGACTCCACTCAATCAGTTCTGAATATCCACGACCTGATG	2625
D R T S T P L N Q F S N I H D L M	

FIG. 17D

21/51

CAATGGCGTGTATCTGGCAGGCCAGGAACCTGCTTACTGCACTGTCGA	2675
Q W R V S R Q A E E L A Y C T V D	
CGGTCGAGGAAAAGAGGGCAAAGGGCTCAATTGGAAGAAGTTGATCAA	2725
G R G K E G K G V N W K K F D Q	
AGGTTGCAGGCCGTAGCAATGTACCTCAAGAACAGGTCAAGGTCCAGGCC	2775
K V A G V A M Y L K N K V K V Q A	
GGCGATCATCTCCTTCTGATGTACACGCATTCAAGAACAGGTCAAGGTCCAGGCC	2825
G D H L L L M Y T H S E E F V Y A	
TGTCATGCATGTTTGCTGGAGCTGTTGCATACCAATGGCGCCAA	2875
V H A [REDACTED] P [REDACTED] M [REDACTED] P [REDACTED]	
Core 1	
TTGATCAGAACCGGTTGAATGAGGATGCAGCCGGCCTTGCTGCATATCCTT	2925
[REDACTED] Q N R L N E D A P A L L H I L	
GCAGATTCAGGTCAAAGCCATTCTGTCAACGCTGACGTTGACCATCT	2975
A D F K V K A I L V N A D V D H L	
GATGAAGATCAAGCAAGTATCGCAGCACATCAAACAATCGGCCGCTATCC	3025
M K I K Q V S Q H I K Q S A A I	
TCAAGATCAGTGTGCCAAACACATACAGCACAACAAAGCCGCCAAAGCAA	3075
L K I S V P N T Y S T T K P P K Q	
TCCAGTGGCTGCCCGACCTCAAGCTTACAATTGACCCGGATGGATTCA	3125
S S G C R D L K L T I R P A W I Q	
GGCGGGTTCCAGTGTAGTCTGGACATACTGGACGCCGATCAACGTC	3175
A G F P V L V [REDACTED] Y W [REDACTED] D [REDACTED] R	
Core 2	
GTATCGCAGTTCAAGCTGGGCCATAGCAAATCATGGCACTGTGCAAGGTC	3225
[REDACTED] R [REDACTED] M A L C K V	
CAAAAAGAAACATGCCAAATGACAAGTACACGACCAGTCCTGGTTGTGT	3275
Q K E T C Q M T S T R P V L G C V	
CCGGAGCACGATAGGACTTGGTTCTTACACTGTCTCATGGGAATCT	3325
R S T I G L G F L H T C L M G I	
TCCTGCCGCACCCACATACCTGGGTACCTGTTGACTTGCACAAAC	3375
F L A A P T Y L V S P V D F A Q N	
CCTAATATTCTGTTCCAAACGCTTGCAGTACAAGATCAAGGATGCATA	3425
P N I L F Q T L S R Y K I K D A Y	

FIG. 17E

22/51

TGCAACGAGTCAAATGTTGGACCACGCCATCGCACGGAGCTGGTAAGA	3475
A T S Q M L D H A I A R G A G K	
GTATGGCTCTGCACGAGCTGAAGAACATCTCATGATTGCGACTGATGGAAGA	3525
S M A L H E L K N L M I A T D G R	
CCACCGCGTTGATGTTgttaagtgaacattgtatgagaggactttcatga	3575
P R V D V	
<u>ttgc</u> ttaactcaatgcagACCAAAGAGTGCCTGTGCACTTGCGCCAGCCA	3625
Y Q R V R V H F A P A	
ACTTAGACCCAACCGCAATCAACACTGTCTACTCACATGTATTGAACCCA	3675
N L D P T A I N T V Y S H V L N P	
ATGGTAGCATCACGATCATAATGTGTATTGAGCCAGTCGAGCTCCATCT	3725
M V A S R S Y M C I E P V E L H L	
CGATGTGCATGCTCTGCACGCCCTCGTCATGCCGTTGACCGTGACA	3775
D V H A L R R G L V M P V D P D	
CAGAGCCCCAACGCTTGCTCGTCCAAGACTCGGGCATGGTGCCAGTGAGC	3825
T E P N A L L V Q D S G M V P V S	
ACGCAAATATCCATTGTCAACCCAGAGACCAACCAACTGTGCTTGAAACGG	3875
T Q I S I V N P E T N Q L C L N G	
CGAGTACGGCGAGATCTGGGTGCAGTCCGAGGGCAATGCTTATAGCTTCT	3925
E Y [REDACTED] S E A N A Y S E	
Core 3	
ACATGTCGAAAGAGCGCTTGGATGCAGAACGCTTCAATGGGAGGACGATT	3975
Y M S K E R L D A E R F N G R T I	
GACGGAGACCCAAATGTGCATATGTTCGTACAGGCATTTAGGATTTT	4025
D G D P N V R [REDACTED] C D [REDACTED] P E L	
Core 4	
GCACAGCGTGACACGCCATTGGACCCAACGGTGCACCTGTTGATATGC	4075
H S V T R P I G P N G A P V D M	
AGGTGCTTTCTGCTTGGAAAGCATAGGTGACACTTTGAAGTCAACGGA	4125
Q V L F V L G S I G D T F E V N G	
CTGAACCATTCTCTATGGACATTGAGCAGTCTGTTAACGTTGTCACCG	4175
L N H F S M D I E Q S V E R C H R	

FIG. 17F

23/51

GAATATTGTCCCTGGAGGCTGgtacgttcttcgattcgctgttatttag	4225
N I V P G G C	
<u>taaatacttaactaacactcacat</u> agTGCTGTTCCAGGCAGGTGGGCTG	4275
A V F Q A G G L	
TTGTTGTCGTTGTGGAAATCTTCCGACGCAACTCCTCGCAAGCATGGTG	4325
V V V V V E I F R R N F L A S M V	
CCTGTGATTGTCAATGCAATTTGAACGAGCATCAGCTGGTCATTGACAT	4375
P V I V N A I L N E H Q L V I D I	
TGTCTCGTTGTGAAAAGGGCGACTTCCACCGGTCTCGTCTGGCGAGA	4425
V S F V Q K G D F H R S R L G E	
AGCAACCGCGAAAGATTCTTGCAGGATGGGTACACGGAAGATGCGCACA	4475
K Q R G K I L A G W V T R K M R T	
ATAGCCCAGTACAGTATAACGGGATCCTAACGGATTCCCAGATGAT	4525
I A Q Y S I R D P N [REDACTED] M I Core 6	
CACGGAAAGAGCCTGGTCCACGGGCTAGCATGACTGGAAGTATGCTTGGC	4575
T E E P G P R A S M T G S M L G	
GAATGGGCGGCCAGCCAGTATCAAGGCCGGTCGACAAGAGCACCGAGT	4625
R M G G P A S I K A G S T R A P S	
CTAATGGGCATGACAGCAGTATGAATAATCTATCCCTAACACAGCAGCA	4675
L M G M T A T M N N L S L T Q Q Q	
ACAGCAGCAATACCAACAGCCGGTATGTATGCTAACAGCAAGGCATGC	4725
Q Q Q Y Q Q P G M Y A Q Q Q G M	
ACCCCCAGCAACAACACCAATTAGCATGTCCAACACGCCACCACAGT	4775
H P Q Q Q H Q F S M S N T P P Q G	
CCACCCCAAGGCCTAGAACTACATGATCCTAGCGACCGCACACCAACAGA	4825
P P Q G V E L H D P S D R T P T D	
CAACCCGCACTTTCTTGCCGACCCCGCGTATGCAGAACCAAGGGCCAAA	4875
N R H S F L A D P R M Q N Q G Q	
TGAACGAGACGGCGCCTACGAACCATGAACTATCAAACGCGTATCAT	4925
M N E T G A Y E P M N Y Q N A Y H	
CCGCATCAACAAACAATACGAATCTGAAGACGGGGGGAGCAGACTCAGCGG	4975
P H Q Q Q Y E S E D G G S R L S G	

FIG. 17G

24/51

CCCCGTGCCAGACGTGCTCGGGCCGGTCCTTCATCCGGGTCCATAGAGC	5025
P V P D V L R P G P S S G S I E	
AGCACGACCAAGCTAACAAACGACAACAATATGTGGAATAATCGCGAGTAC	5075
Q H D Q A N N D N N M W N N R E Y	
TATGGTAACAGCCCATCGTATGCAGGCGGATACACGCAAGATGGCAATAT	5125
Y G N S P S Y A G G Y T Q D G N I	
CCACGAGCAGCAACAAACACGATGAGTACACGAGTAATGCGTCATATGGCG	5175
H E Q Q Q H D E Y T S N A S Y G	
GAAATCAAGGAGCAGGCGGAGGCAGCGGCGGCGGTGGCGGTCTCCGAGTT	5225
G N Q G A G G G S G G G G G G L R V	
GCAAATCGTACAGCTCCGACAGCGAGGGTGCAGATGACGACGCTTGGAG	5275
A N R D S S D S E G A D D D A W R	
ACGTGATGCCCTTGCTCAGATCAATTTGCAGGCGGCGCTGCTGCTGCCT	5325
R D A L A Q I N F A G G A A A A A	
CCGCTGGAGCACCTGCTGCTGGTCTTCTTCGAGCCGGCCATGCG	5375
S A G A P A A G A S S S Q P G H A	
CAG <u>I</u> AGACGGATATGCGTGAGTTTTAAATTGCTACATAGAGAC	5425
Q END	
CGTTGTATACGCAGGTTCAAATTAGAAGAGCGAATATGCATATCAGCTG	5475
TTGTTCAATGTTCTAGTTGGAAAGGTTAACCCCCCCCCCTTCCCTTCC	5525
AAGACTTTCACTGTTGTGTGATTAAATCTGGAGATTCAAATCT	5575
ACATCTCGCTACATACAGGTGTTGATAACGTAGGGGGCAGAAGGGT	5625
ATCTCGTGTATTAGACTGGAGTTGATGAATCAAGGTGTTGAGAAAA	5675
AAAGAGAGAGCGGTGAAGGGCGGGGGGATAGGTGGTGTGACGTGGCTG	5725

FIG. 17H

25/51

AAGCCAGGTACGCATCTGCGCAGGGCCCGCAAGCCCAGTATGACGTTT	-286
TACCCAGAGCCAGCCTGCTCCTCGGCTTGGCGCCGCCCTGGCTAGTCAG	-236
CCCCCATCAGTCAGCGGCCAGTCACGTGTTGGCGCGACAAGCTCCCACA	-186
TGGCCGATGATCATTGTGCCCTGGCTATGCCGTCTGCCGTGCTTGTAC	-136
GTGCCCGGAAATGACGACGCCAAAGTCTCACACAGCCGCTCGCTGCTG	-86
***** CTGCTTGCATCTGGTCTTGCTACACTGGCTCCTTTCGCAGAACACAGA	-36
ACGATCCACCAACAGGAAA <u>ACTATAGCCACCACCATGGCCACCC</u> TCATC M A T L I	15
CGCCCCCGCCCGCCGACCACTCCCAGTCGGCATTGAGAACGTCTTGA R P P P A D H S Q S A I E N V L E	65
ACTGACGCAGCTGGCCGACATTGATCCAAACATCTCACAAACACTCGTC L T Q L A D I D P N I F T N T R	115
CCCTATGGCATCCCCCGGTGCGCGCCGATCTTGGCGGTGCTGCCATT P L W H P P G A R G I F G G A A I	165
GCCCAGACACTCAGCGCCGCCAGAACGAGCGTTGACCCGACTTCACTGT A Q T L S A A Q K T V D P D F T [REDACTED]	215
GCATTGATGCACTGCTACTTCATCCTTGCAGGCAACTCCGAGATCCCCG [REDACTED] M H C Y F I L A G N S E I P	265
TCATATAACCATGTAGAAAGGGTGCCTCGGGAAAGTCGTTGCAACCAGG V I Y H V E R V R S G K S F A T R	315
ACGGTGCAGGCCGGCAAAGGGGCAACGTCATCTTCACTACGACCATGAG T V Q A R Q R G N V I' F T T T M S	365
TTTGTTGCCAGAACAGTGGAGGCCTCAAAAGGTGAGCACATCTACC F V R Q N S G G A Q K V E H I Y	415
CGATGCCAGACGTCCCAGGCCAAGGAAGGTAGCGACGACTTGAAGACG P M P D V P A P K E G S D D L K T	465
CCCAACGATGGCCAGAGTCCCTTCCAGACCCAGCTGTTGCCATTGAAAA P N D G Q S P F Q T Q L L P I E N	515
CGgtgagtgacttcgcataccat <u>ccttaagcctgccttagctcatatcta</u>	565

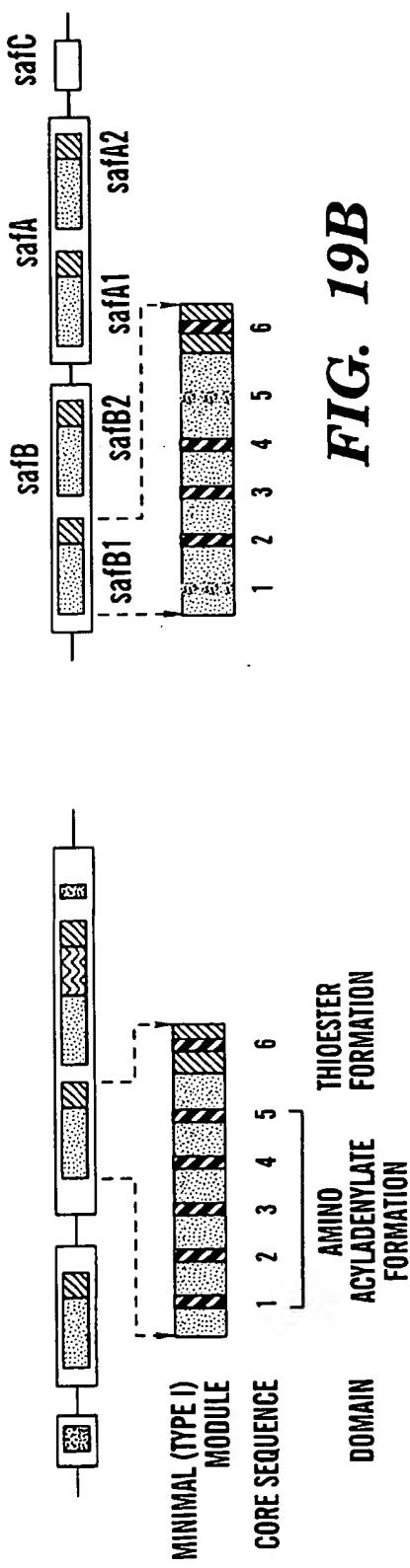
FIG. 18A

26/51

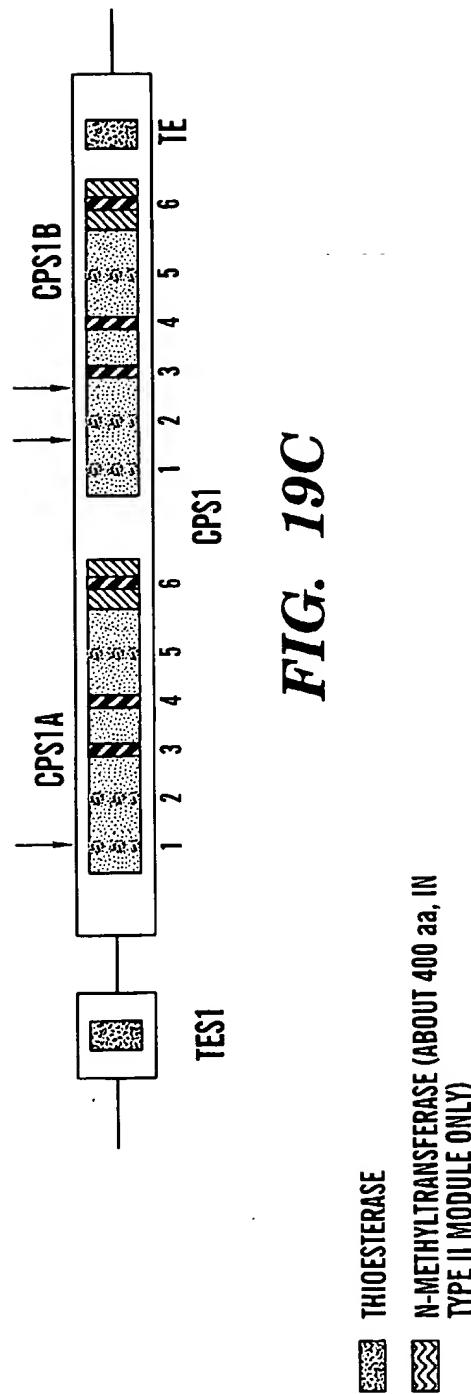
gCAGACGACTCCGACAAGCCCCACACCAAGAAATGCCGACAATGGATAAA	615
A D D S D K P H T K K C R Q W I K	
GGCTCGCGGTAAAATCTCCCCCGCTGGCGGTACGAAGCCCATCTCTCCG	665
A R G K I S P A G G H E A H L S	
CCATCGCATACTATGTCCGATAGCTACTTCATCGGCACCCTCGCGCGTGC	715
A I A Y M S D S Y F I G T V A R A	
CACAAGCTTCTGCGCTACTCGAACCGAGCGCAAGAGCAGGGCCAGGTCGAG	765
H K L L R Y S N Q R K S R A R S S	
CATCGACGAGGACGTACTTAAGAACGCTGCTGAGATGGATGATGCCGAGT	815
I D E D V L K K L L E M D D A E	
TACAGCGCCAAGCTTTGTCAACGAATCAGACAAGCAGCGCATACTGAA	865
L Q R O S F V N E S D K Q R I R E	
TTGAGGAAAGCAGAACCTGGCAAAGAGCAGGAGACGCAAAGCCTGAGAT	915
L R K A E D L A K S G D A K P E I	
TGGCATGATGGTTAGTCTGGACCACACCATTACTTACAATCCTCGCA	965
G M M V S L D H T I Y F H N P R	
GTTTCCGCGCAGATGAATGGATCTTCACCGAGATGGAGACCCCTGGGCT	1015
S F R A D E W I F T E M E T P W A	
GGTGATGGTCGGCCCTCGTTCCCAGAGAACGATGTATACCAAGGGATGGCAC	1065
G D G R G L V S Q R M Y T K D G T	
GCTCATTGCCAGCTCGTCCAAGAGGTAAAGCAGCTTCTTATGTCTGCTA	1115
L I A S C V Q E V S S L L M S A	
TAGTAGACTGTGCTTACATGAATAGGGCGTCATACGAT <u>I</u> GAAGCAGAACG	1165
I V D C A Y M N R A S Y D END	
AAAGTAAGCTATGCCATGACCGTGTAGATTGCTCGTATGTCCGTGTA	1215
CTTGATTGATTGACTTTATGCCGCCCTTCACTTGACCTTGTGCG	1265
AGTAGACTCCGCTTTCTATTGTTAGTAGCTTAGATTGGAGGTCT	1315
GGAACACACGTACCTCACGTATA <u>CTTTAA</u> ATACAAGGTTCTCGTTA	1365
CAAATAGCAACTCGCACTTGTGTAGTACACGACTTGGCACAGCAAGTGCA	1415
TGCAGATGATGCGCACTGCATGCAGCCCTAGAACGCTGCACCTGTGCTGT	1465
TCGTCCCTCCCACGTTACAGGCCAGGGTCCAATTAGCGCAATGCCGGTCT	1515
CGGGCTAGCATGGCCCTCACACTTGGATTCTCCGAAGCTACCGTCGGCAG	1565

FIG. 18B

27/51

**FIG. 19A****FIG. 19B**

SUBSTITUTE SHEET (RULE 26)

**FIG. 19C**

■ THIOESTERASE
 ■ N-METHYLTRANSFERASE (ABOUT 400 aa, IN
 TYPE II MODULE ONLY)

28/51

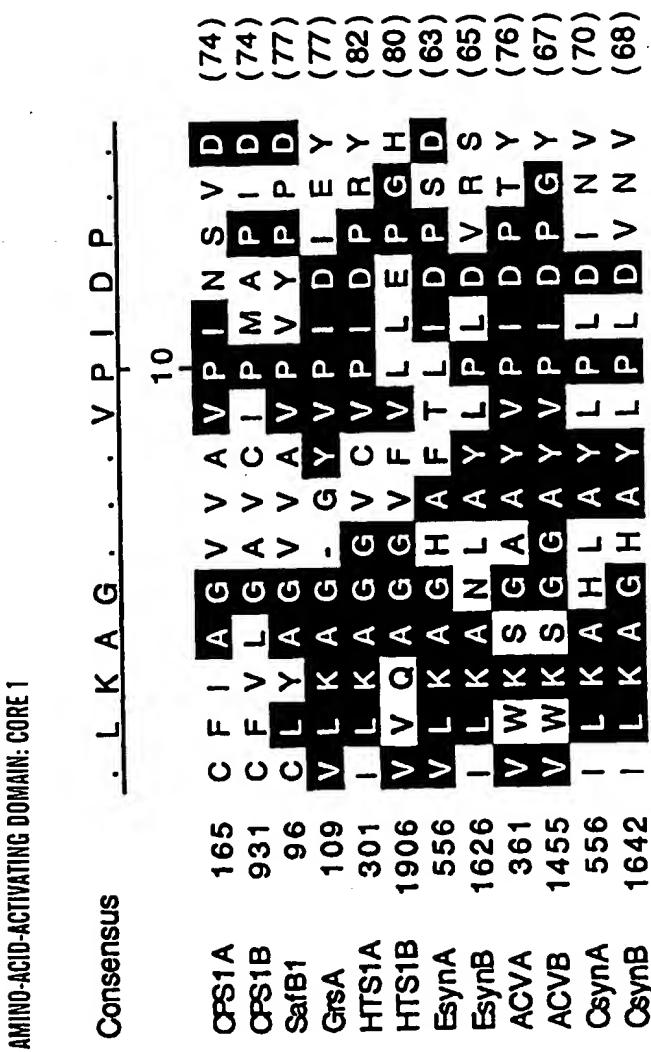


FIG. 20A

29/51

Consensus	F	T	S	G	.	T	G	.	P	K	G	V	.	.	H	R	.	I	
CPS1A	253	F	S	R	A	P	T	G	D	L	R	G	V	V	L	S	H	R	-
CPS1B	1019	W	T	Y	W	.	T	P	D	Q	R	A	V	V	Q	S	H	R	-
Saf1B1	187	Y	T	S	G	S	T	A	D	P	K	G	T	M	L	E	H	R	N
GrsA	190	Y	T	S	G	T	T	G	N	P	P	K	C	-	V	V	S	A	Y
HTS1A	397	F	T	S	G	S	T	G	-	P	K	G	A	V	M	-	E	H	R
HTS1B	2000	F	T	S	G	-	T	G	-	F	T	S	G	S	T	G	Q	H	T
EsynA	633	F	T	S	G	T	T	G	-	F	T	S	G	T	T	V	E	H	G
EsynB	1706	F	T	S	G	-	T	G	-	Y	T	S	G	T	T	M	-	E	H
ACVA	451	Y	T	S	G	T	T	G	-	Y	T	S	G	S	T	G	R	G	I
ACVB	1538	F	T	S	G	-	T	G	-	F	T	S	G	T	T	V	P	K	G
CsynA	640	F	T	S	G	-	T	G	-	Y	T	S	G	S	T	G	K	P	G
CsynB	1724	F	T	S	G	-	T	G	-	F	T	S	G	S	T	G	K	P	G

* AN INSERTION (2 RESIDUES BETWEEN R AND A) IS NOT SHOWN.

FIG. 20B

30/51

	<u>G E L . V . G . G L A R G Y</u>									
	Consensus									
CPS1A	583	G	E	I	W	V	D	S	P	S
CPS1B	1209	G	E	I	W	V	Q	S	A	G
SafB1	418	G	E	I	W	V	R	G	P	F
GsA	374	G	E	L	C	I	G	G	V	(32)
HTS1A	569	G	E	L	L	I	E	S	G	(25)
HTS1B	2184	G	E	L	L	I	E	G	S	(23)
EsynA	816	G	E	L	V	V	T	G	I	(23)
EsynB	1893	G	E	L	V	H	I	G	D	(31)
ACVA	640	G	E	L	H	I	G	G	R	(26)
ACVB	1728	G	E	L	Y	L	G	G	K	(30)
OsynA	830	G	E	L	V	V	S	G	D	(23)
OsynB	1916	G	E	L	V	V	T	G	D	(23)

FIG. 20C

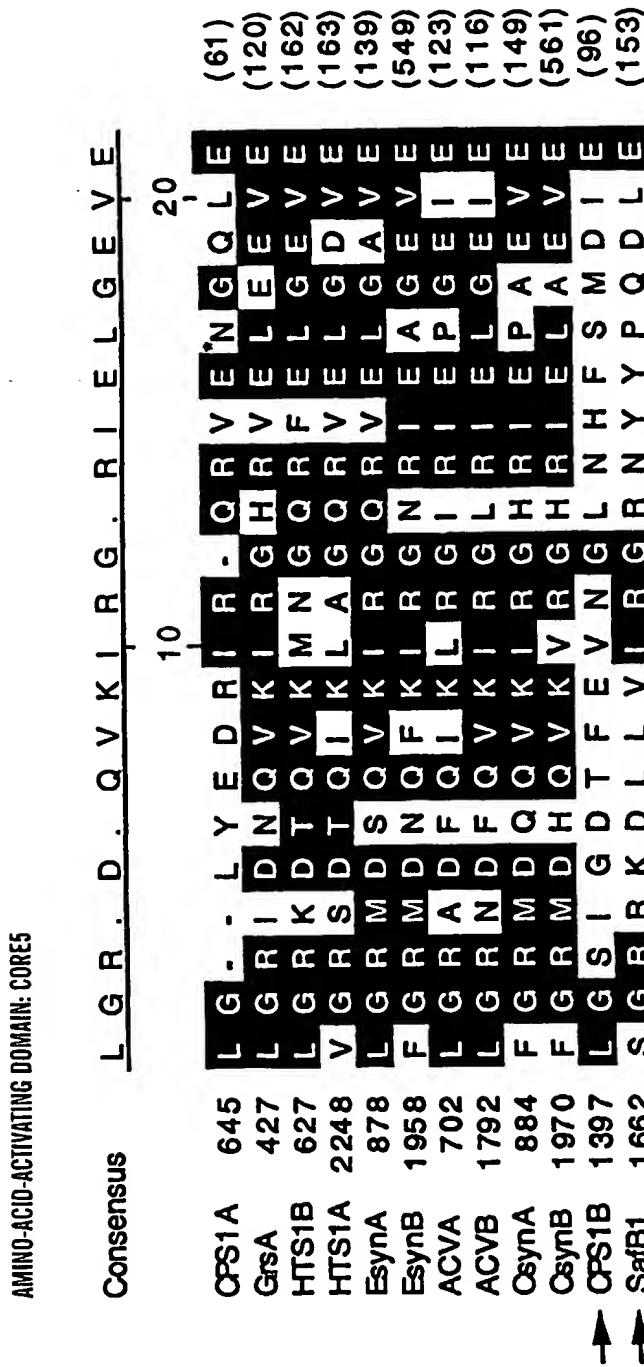
AMINO-ACID-ACTIVATING DOMAIN: CORE 3

31/51

	<u>Y - R T G D L . R</u>	
Consensus		
CPS1A	628	F L R T G L L G F (13)
CPS1B	1301	Y V R T G D L G F (9)
SafB1	454	W L R T G D L G F (11)
GsA	410	Y - K T Q A R (8)
HTS1A	609	Y - R T V A R (8)
HTS1B	2223	Y - K T D L V R (8)
EsynA	860	Y - R T G D L A C (9)
EsynB	1939	Y - R T G D R M R (10)
ACVA	684	Y - K T G D L A R (9)
ACVB	1772	Y - K T G D L V R (11)
CsynA	866	Y - R T G D R A R (10)
CsynB	1956	Y - R T G D R A R (10)

FIG. 20D

32/51



*AN INSERTION (TWO AMINO ACID) BETWEEN E AND N IN CPS1A IS NOT SHOWN.
THE POORLY CONSERVED CORES 5 IN CPS1B AND SafB1 ARE INDICATED BY ARROWS

FIG. 20E

33/51

THIOLATION DOMAIN: CORE 6

Consensus	F F . . G G D S L . A . .	10	
CPS1A	L D I P F L D S L S E R C	574	(193)
CPS1B	R D P N G Q D S L M I T E	530	
SafB1	L P D L G L D S L V E	562	(590)
GrsA	F Y A L G G D S I K A I Q	471	
HTS1A	F I H A G G D S I T A M Q	524	(1082)
HTS1B	F F S S G G N S M A I A	529	
EsynA	F F E M G G H S I I A - K	497	(906)
EsynB	F F Q L G G H S I I A T K	917*	
ACVA	F F R L G G H S I T C I Q	500	(595)
ACVB	F F S L G G D S L K S T K	489	
CsynA	F F D L G G H S L T A M K	510	(577)
CsynB	F F N V G G H S L L A T K	922**	

* ACTIVE SITE FOR 4-PHOSPHOPANTETHINE BINDING.

** TYPE II MODULES CONTAINING A METHYLTRANSFERASE DOMAIN (ABOUT 400 AMINO ACIDS) BETWEEN CORES 5 AND 6. ALL OTHERS ARE TYPE I MODULES WITHOUT THIS INSERTION.

FIG. 20F

34/51

Consensus		1619	V L R P S G S E Q H D Q A	(125)
OVS1-TE		Y H V Y F A L T Y T F A F	F G W S S M G A L I S C S L G N L M T L Y G H F	(168) (186) (157) (173) (163) (168)
ACVA-TE		Y V L I F L T L T L	G Y S H G Y S G Y S M G G L V	- V E L A V F E L V A F E L
GrsB-TE		Y F L I F L T L T L	G Y S H G Y S G Y S M G G L V	- V E L A V F E L V A F E L
GrsT-TE		F A L I F L T L T L	G Y S H G Y S G Y S M G G L V	- V E L A V F E L V A F E L
SrfA-TE		L T L T L T L T L	G Y S H G Y S G Y S M G G L V	- V E L A V F E L V A F E L
TycC-TE		93 Y T L T L T L T L	G Y S H G Y S G Y S M G G L V	- V E L A V F E L V A F E L
TycF-TE		76 F A F F F F F F F	G Y S H G Y S G Y S M G G L V	- V E L A V F E L V A F E L

FIG. 21

34/51

FIG. 21

35/51

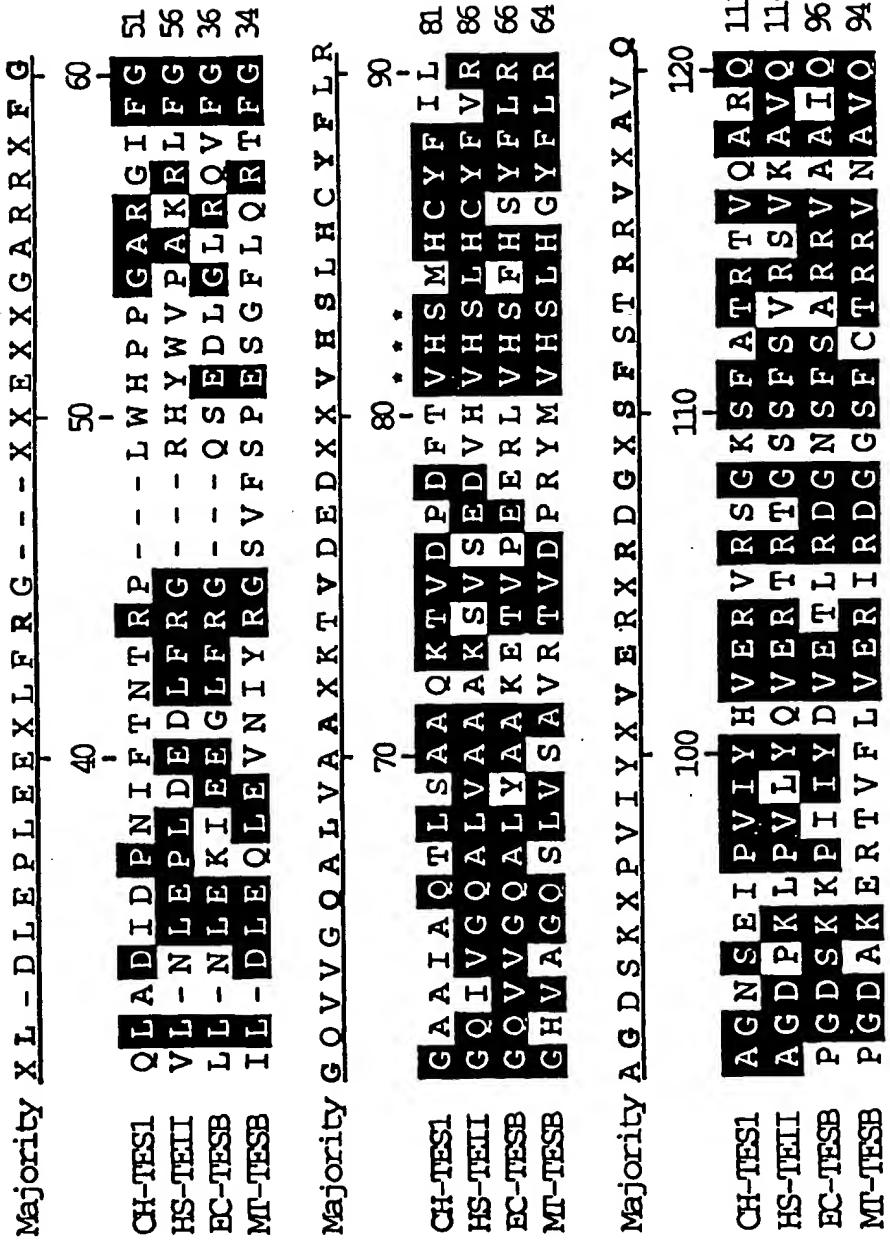


FIG. 22A

36/51

<u>Majority H G K P I F X M T A S F O X X Q X G X X O H O X X M P X A P</u> CH-TES1 R G N V I F T T M S F V R Q N S G G A Q K V E H I Y P M P 141 HS-TEII H G K P I F I C O A S F Q Q A Q P S P M Q H Q F S M P T V P 146 EC-TESB N G K P I F Y M T A S F Q Q A P E A G F E - H Q K T M P S A P 125 MT-TESB H G E T I F S M A A S F Q Q T E Q E G I T - H Q D V M P A A P 123	<u>Majority - - - - E P X O Q V W X R A G S - X P G D X X X H X C A</u> CH-TES1 - - - - K C R Q W I K A R G K I S - P A G G H E A H L S A 205 HS-TEII Q L Q R M E P K Q M F W V R A R G Y I G E G D M K M H C C V 226 EC-TESB - - - - E P H R Q V W I R A N G S - V P D D D L R V H Q Y L 198 MT-TESB - - - - A S Q Q Q V W L R H R D P - L P D D P V L H I C A 188	<u>Majority L A Y M S D L X F L G T A L Q P H X L</u> CH-TES1 I A Y M S D S Y F I G T V A R A H K L L R Y S N Q R K S R A 235 HS-TEII A A Y T S D Y A F L G T A L L P H Q W Q H K V H - - - - - 250 EC-TESB L G Y A S D L N F L P V A L Q P H G I - - - - - 217 MT-TESB L A Y M S D L T L L G S A - Q V N H L - - - - - 206
---	---	---

FIG. 22B

37/51

<u>Majority</u> CH-TESI HS-TEII EC-TESB MT-TESB	- - - - - D X X X X - X Q M A S L D 310 K Q R I R K A E D L A K S G D A K P E I G M M V S L D 320 - - - - - F M V S L D - - - - - G F L E P G I Q I A T I D - - - - - D V R D Q - L Q V A S L D 295 256 230 218
<u>Majority</u> HS M W F H R - P F R A D E W L L Y E X E S P S A G G G R	330 340 H T I Y F H N P R S F R A D E W I F T E M E T P W A G D G R 350 - - P F R A D H W M L Y E C E S P W A G G S R - - P F N L N E W L L Y S V E S T S A S S A R - - P F R A D E W L L Y D Q S S P S A S G G R 325 284 258 246
<u>Majority</u> GL V R G E X Y T Q D G V L V A S C V Q E G V X R X X X X	360 370 G L V S O R M Y T K D G T L I A S C V Q E V S S L L M S A I 380 - - G L V H G R V W S Q D G V L A V T C A Q E G V I R V K P Q V - - G F V R G E F Y T Q D G V L V A S T V Q E G V M R N H N - - A L T R G E I F T R S G E M V A A V M Q E G L T R H R R G H 355 314 286 276

FIG. 22C

38/51

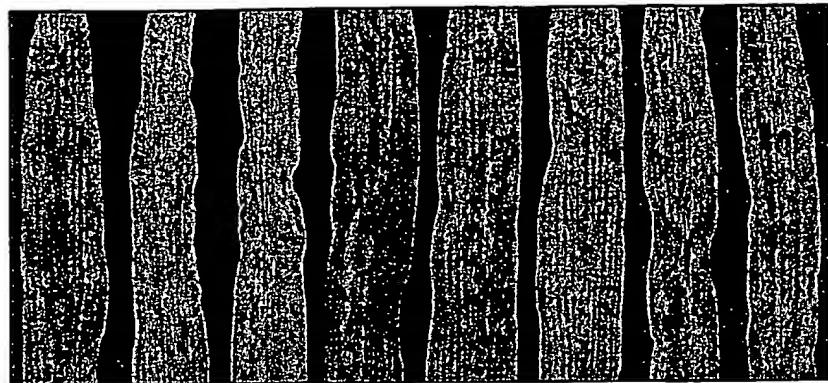


FIG. 23A

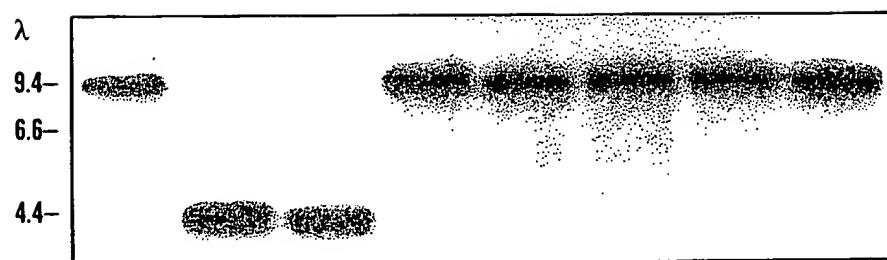


FIG. 23B

39/51

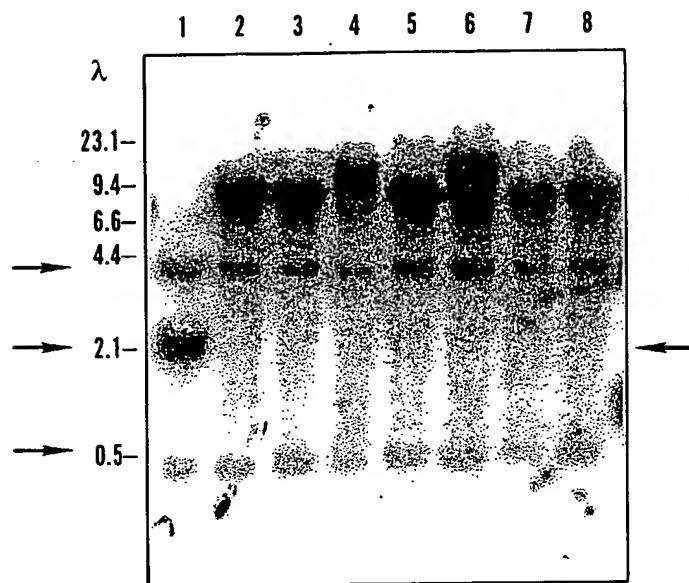


FIG. 24

40/51

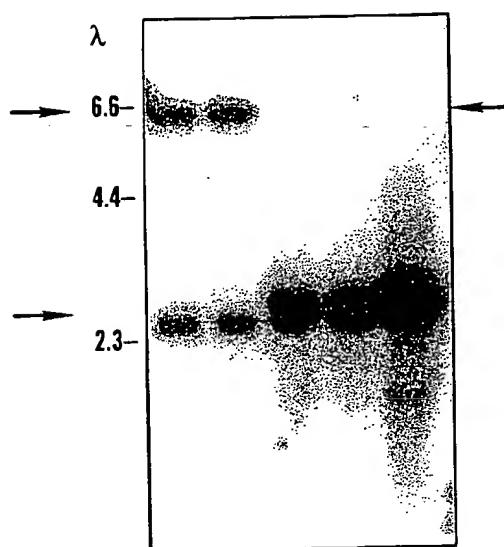


FIG. 25

41/51

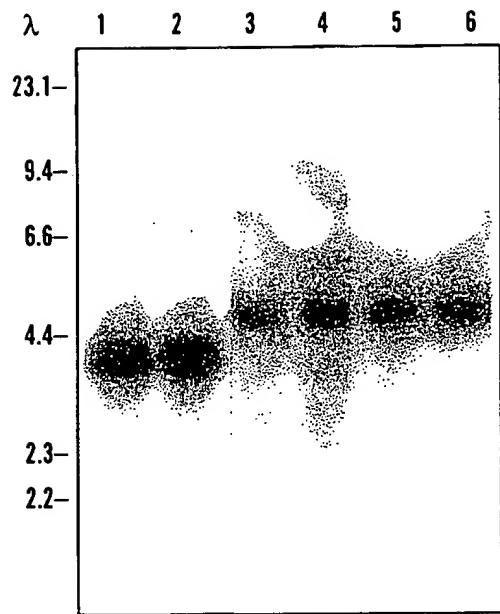
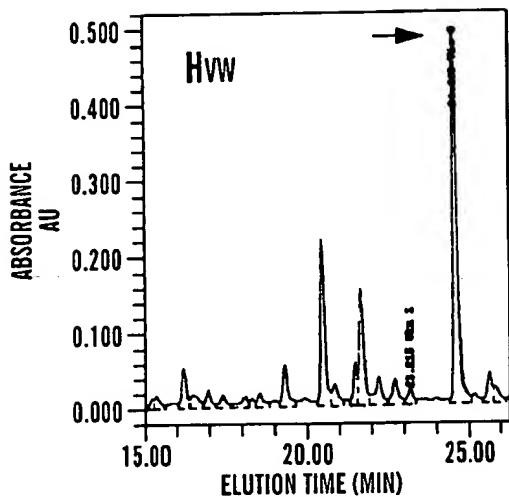
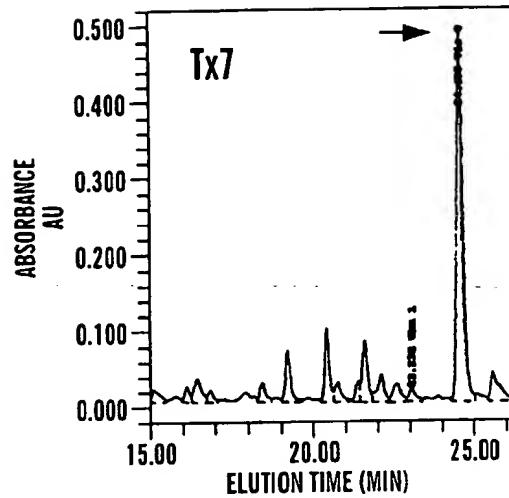
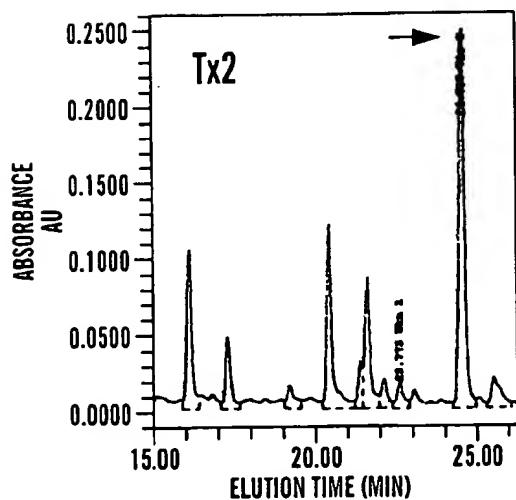
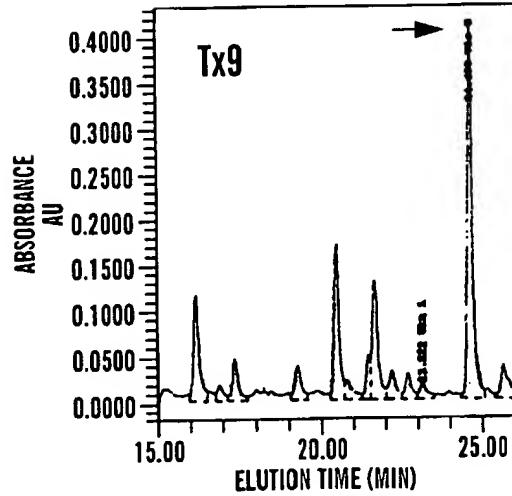
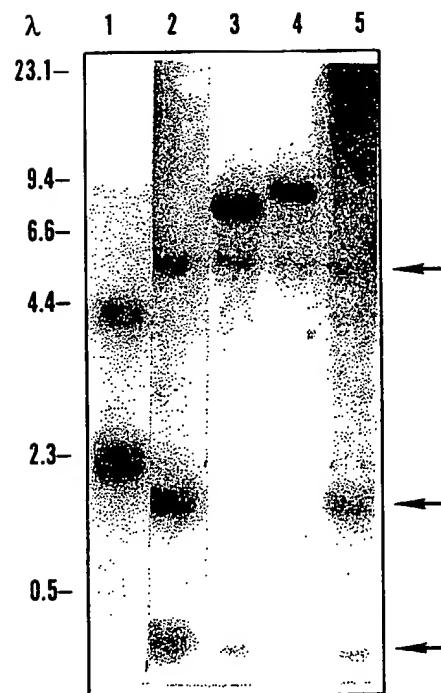


FIG. 26

42/51

**FIG. 27A****FIG. 27B****FIG. 27C****FIG. 27D**

43/51

**FIG. 28A****FIG. 28B**

SUBSTITUTE SHEET (RULE 26)

44/51

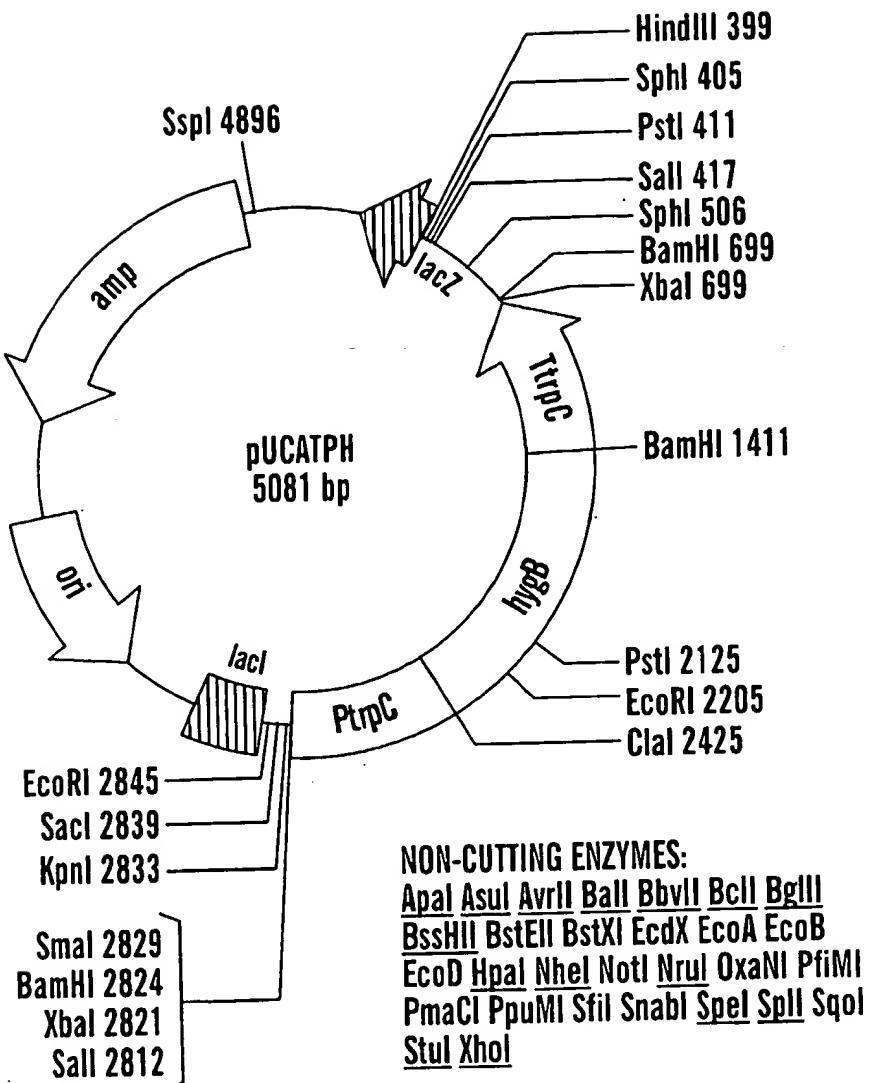


FIG. 29

45/51

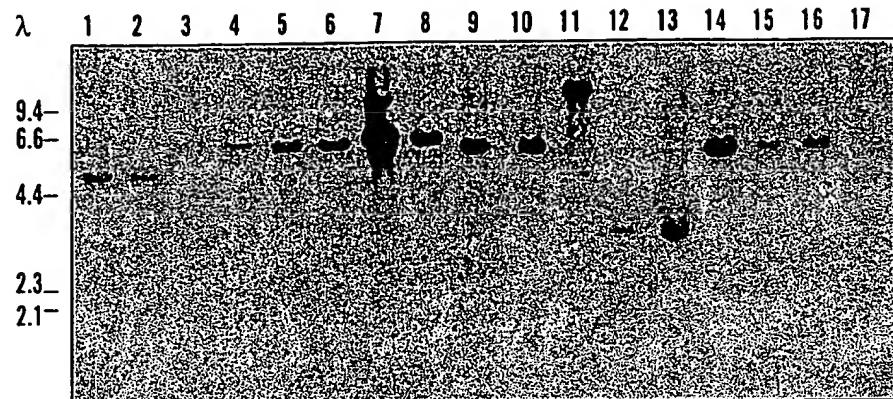


FIG. 30A

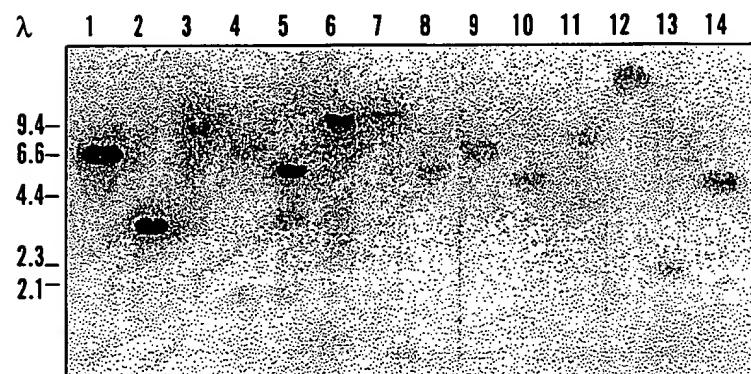


FIG. 30B

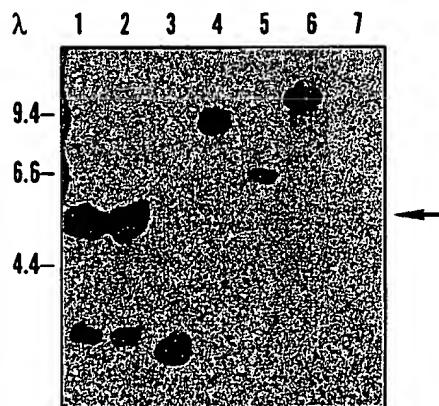
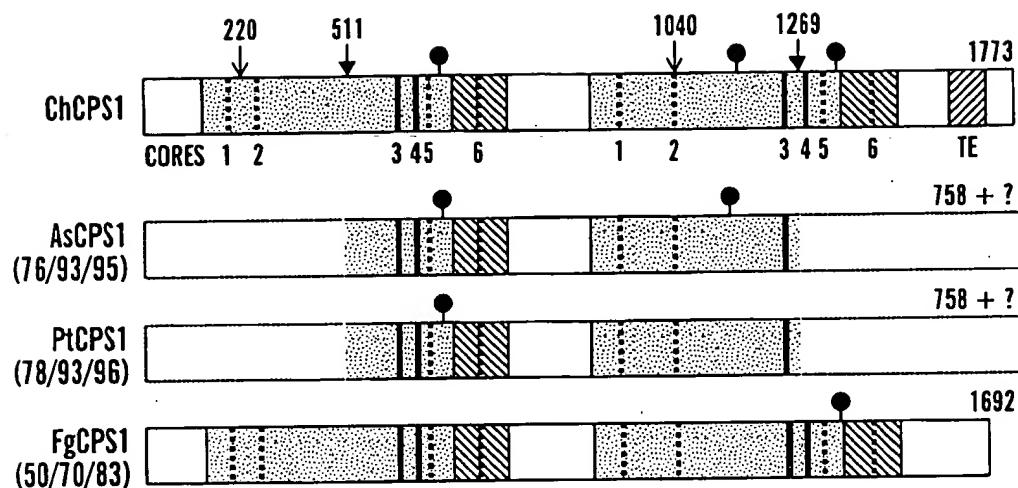


FIG. 30C

SUBSTITUTE SHEET (RULE 26)

46/51

**FIG. 31A**

47/51

SUBSTITUTE SHEET (RULE 26)

48/51

FIG. 32

SUBSTITUTE SHEET (RULE 26)

49/51

FIG. 33

50/51

FIG. 34

51/51

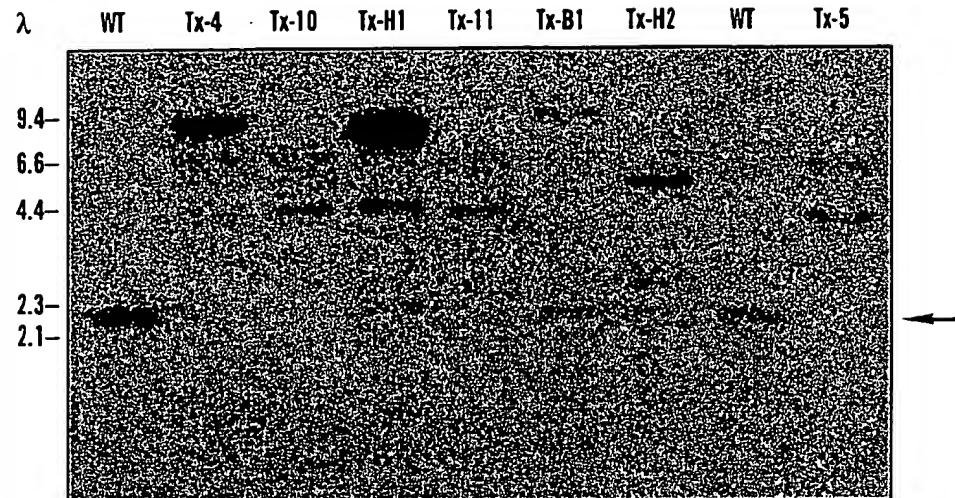


FIG. 35A

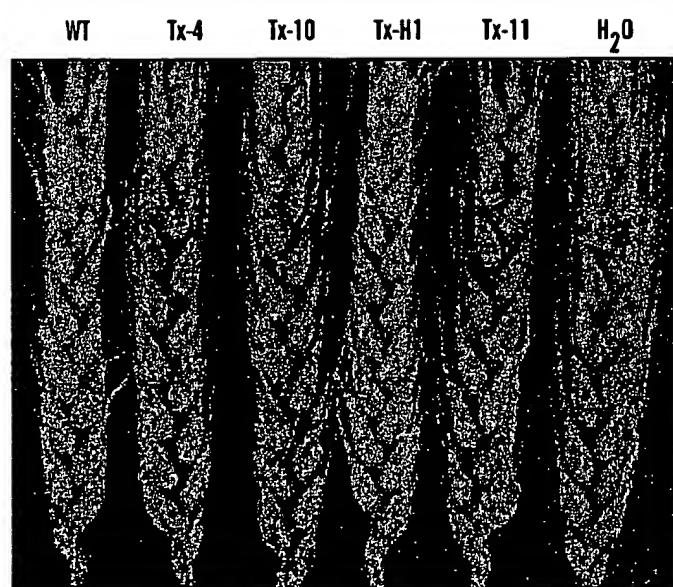


FIG. 35B

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
31 May 2001 (31.05.2001)

PCT

(10) International Publication Number
WO 01/38489 A3

- (51) International Patent Classification²: A01H 5/00, C12N 5/14, 15/29, 15/52, 15/82
- (21) International Application Number: PCT/US00/32227
- (22) International Filing Date: 22 November 2000 (22.11.2000)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data: 09/448,215 23 November 1999 (23.11.1999) US
- (71) Applicant: CORNELL RESEARCH FOUNDATION, INC. [US/US]; Suite 105, 20 Thornwood Drive, Ithaca, NY 14850 (US).
- (72) Inventors: YODER, Olen, C.; 44 Hungerford Road, Ithaca, NY 14850 (US). TURGEON, Barbara, C.; 608 Mitchell Street, Ithaca, NY 14850 (US). LU, Shun-Wen; 604 Winston Court, Apt. #4, Ithaca, NY 14850 (US).
- (74) Agents: GOLDMAN, Michael, L. et al.; Nixon Peabody LLP, Clinton Square, P.O. Box 31051, Rochester, NY 14603 (US).

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

— with international search report

(88) Date of publication of the international search report:
31 January 2002

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

WO 01/38489 A3

(54) Title: PEPTIDE SYNTHETASE GENE CPS1

(57) Abstract: The present invention relates to genes cloned from the plant pathogens *Cochliobolus heterostrophus*, *Alternaria solani*, *Fusarium graminearum*, and *Pyrenophora teres*, that encode a CPS1 peptide synthetase required for fungal pathogenesis. The nucleic acid molecules in a vector, a host cell, or a plant is also disclosed. The invention further provides a protein or polypeptide encoded by the CPS1 genes. Other aspects of the invention relate to a method of imparting disease resistance to a plant by overexpressing a protein of the present invention in a plant and a method for identifying inhibitors of a CPS1 protein in a sample.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/32227

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : A01H 6/00; C12N 5/14, 15/89, 15/52, 15/82

US CL : Please See Extra Sheet.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 495/320.1, 419, 419, 468; 556/23.2, 23.6; 800/279, 306, 314, 315, 316, 317.2, 317.4, 320, 320.1, 320.2, 320.3

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

STN, AGRICOLA, CAPLUS, BIOSIS, EMBASE, USPAT
search terms: peptide synthase, peptide synthetase, DNA, cDNA, gene, nucleic, disease, fungal, bacterial, pathogen, resistance

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	NIKOLSKAYA et al. Identification of Peptide Synthetase-Encoding Genes from Filamentous Fungi Producing Host-Selective Phytotoxins or Analogs. Gene. 1995. Vol. 165, pages 207-211.	49-53

Further documents are listed in the continuation of Box C. See patent family annex.

* Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"Z"	document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means		
"P" document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

25 MAY 2001

Date of mailing of the international search report

26 JUL 2001

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231
Facsimile No. (703) 305-3230

Authorized officer

AMY NELSON

Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/32227

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:

2. Claims Nos.: 1-5,25-44 because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
applicant did not submit a computer readable form of the sequence listing, and hence SED ID NO:41 could not be searched.

3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
1-5,25-44,49-53

Remark on Protest

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/32227

A. CLASSIFICATION OF SUBJECT MATTER:

US CL :

435/320.1, 419, 419, 468; 536/23.2, 23.6; 800/279, 306, 314, 315, 316, 317.2, 317.4, 320, 320.1, 320.2, 320.3

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

Group I, claim(s) 1-5, 25-44, 49-53, drawn to DNA of SEQ ID NO:41, vector, transformed host cell, transgenic plant, and plant transformation method. Claims 25-44 and 49-53 will be examined to the extent they read on SEQ ID NO:41.

Group II, claim(s) 6-8, drawn to protein of SEQ ID NO:42.

Group III, claim(s) 9-15, 25-44, 49-53, drawn to DNA of SEQ ID NO:45, vector, transformed host cell, transgenic plant, and plant transformation method. Claims 25-44 and 49-53 will be examined to the extent they read on SEQ ID NO:45.

Group IV, claim(s) 14-18, drawn to protein of SEQ ID NO:44.

Group V, claim(s) 17-21, 25-44, 49-53, drawn to DNA of SEQ ID NO:46, vector, transformed host cell, transgenic plant, and plant transformation method. Claims 25-44 and 49-53 will be examined to the extent they read on SEQ ID NO:46.

Group VI, claim(s) 22-24, drawn to protein of SEQ ID NO:46.

Group VII, claim(s) 45-48, drawn to inhibitor selection method.

The inventions listed as Groups I, II, III, IV, V, VI and VII do not relate to a single inventive concept under PCT Rule 15.1 because, under PCT Rule 15.2, they lack the same or corresponding special technical features for the following reasons:

The phrase "CPS1" and "stringent conditions" are not well defined in the disclosure, and hence the claims of Group I encompass essentially any DNA from a plant pathogen encoding a cyclic peptide synthetase. Nikolskaya (Gene 165: 207-211, 1995) teaches various CPS genes isolated from plant pathogens (see entire article). Hence, there is no special technical feature under PCT Rule 15.2 which links the DNA of Group I with the protein of Group II.

The DNAs of Group III and V, and the proteins of Group IV and VI differ in composition and structure from the DNA of Group I and the protein of Group II, respectively, and hence are not so linked by a special technical feature. Separate searches and considerations would be required for examination of each of the nucleic acid sequences or amino acid sequences.

Also, the inhibitor selection method of Group VII is distinct from the plant transformation method of Group I in starting materials, method steps, and end products.

Therefore, the inventions of Groups I, II, III, IV, V, VI and VII do not relate to a single inventive concept under PCT Rule 15.1.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/32227

A. CLASSIFICATION OF SUBJECT MATTER:

US CL :

435/320.1, 418, 419, 468; 536/23.2, 23.6; 800/279, 308, 314, 315, 316, 317.2, 317.4, 320, 320.1, 320.2, 320.3

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

Group I, claim(s) 1-5, 25-44, 49-53, drawn to DNA of SEQ ID NO:41, vector, transformed host cell, transgenic plant, and plant transformation method. Claims 25-44 and 49-53 will be examined to the extent they read on SEQ ID NO:41.

Group II, claim(s) 6-8, drawn to protein of SEQ ID NO:42.

Group III, claim(s) 9-13, 25-44, 49-53, drawn to DNA of SEQ ID NO:43, vector, transformed host cell, transgenic plant, and plant transformation method. Claims 25-44 and 49-53 will be examined to the extent they read on SEQ ID NO:43.

Group IV, claim(s) 14-16, drawn to protein of SEQ ID NO:44.

Group V, claim(s) 17-21, 25-44, 49-53, drawn to DNA of SEQ ID NO:45, vector, transformed host cell, transgenic plant, and plant transformation method. Claims 25-44 and 49-53 will be examined to the extent they read on SEQ ID NO:45.

Group VI, claim(s) 22-24, drawn to protein of SEQ ID NO:46.

Group VII, claim(s) 45-48, drawn to inhibitor selection method.

The inventions listed as Groups I, II, III, IV, V, VI and VII do not relate to a single inventive concept under PCT Rule 15.1 because, under PCT Rule 15.2, they lack the same or corresponding special technical features for the following reasons:

The phrase "CPS1" and "stringent conditions" are not well defined in the disclosure, and hence the claims of Group I encompass essentially any DNA from a plant pathogen encoding a cyclic peptide synthetase. Nikolskaya (Gene 165: 207-211, 1995) teaches various CPS genes isolated from plant pathogens (see entire article). Hence, there is no special technical feature under PCT Rule 15.2 which links the DNA of Group I with the protein of Group II.

The DNAs of Group III and V, and the proteins of Group IV and VI differ in composition and structure from the DNA of Group I and the protein of Group II, respectively, and hence are not so linked by a special technical feature. Separate searches and considerations would be required for examination of each of the nucleic acid sequences or amino acid sequences.

Also, the inhibitor selection method of Group VII is distinct from the plant transformation method of Group I in starting materials, method steps, and end products.

Therefore, the inventions of Groups I, II, III, IV, V, VI and VII do not relate to a single inventive concept under PCT Rule 15.1.